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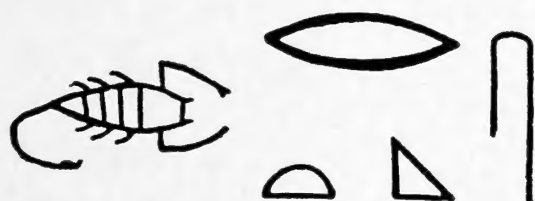
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1996-2000

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v. 5
part 1
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SERKET

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VOLUME **5**
PART **1**

CAIRO - EGYPT
1996

SERKET

Volume 5

Part 1

December, 1996

Cairo, Egypt

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Subscription for volume 5 (1996-1997) :
US \$ 25.00 (personal rate)
US \$ 35.00 (institutional rate)

Back issues :
Volume 1 (1987-1990),
Volume 2 (1990-1992),
Volume 4 (1994-1996):
US \$ 25.00 (p.r.) per volume
US \$ 35.00 (i.r.) per volume

Volume 3 (1992-1993):
US \$ 35.00 (p.r.), US \$ 45.00 (i.r.)

Correspondence concerning subscription, back issues, publication, etc. should be addressed to the editor.

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41, El-Manteqa El-Rabia St.,
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PREFACE

In the 5th volume of SERKET, I hope you will find new additions with the continuation of bibliographic works on spiders and other arachnids. Your comments are very important to evaluate these works. Your contributions for publication are also requested.

My sincere gratitude for everyone who shared in pushing this bulletin ahead and forward.

The Editor

Hisham El-Hennawy

Ecological studies on spiders in Giza governorate

By

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Introduction

Among about 35 000 species of spiders, only 366 species were recorded from Egypt (El-Hennawy, 1990). The studies on Egyptian spiders are few, sparse, and mostly taxonomic. There are two ecological studies on Egyptian spiders in cultivated areas in Assiut (Negm et al., 1976) and El-Fayoum (Rahil, 1988).

This study was carried out to reveal some ecological aspects of spiders in Giza governorate.

Materials and Methods :

The present study was conducted in Giza Governorate. The study was aimed towards surveying terrestrial spiders in different habitats (i.e. horticulture, field crops and ornamental plants).

Survey studies were conducted in two fields at Cairo University, Faculty of Agriculture Experimental Station and four plastic houses belong to Ministry of Agriculture. Collections were carried out randomly in the selected habitats. Collecting methods included : beating net (branch shaking) and pitfall traps. Biweekly samples were collected over two years. The collected spiders were preserved in 70 % ethanol in glass vials and transferred to the laboratory for counting and classification. Plants were surveyed for the durations indicated in Table (1).

A- Habitat conditions :

1- Apple trees, Malus sylvestris Mill were about 2 m height with dense foliage, and a lot of weeds around the plants especially milky weeds. These trees were weeded out from time to time. No chemical control was applied. The presence of weeds caused a good source of different types of insects to inhabit this ecosystem.

2- Grape, Vitis quadrangularis Vall was ground type which grow up horizontally with dense foliage. This crop was replanted yearly. Different types of weeds existed in the field and only mineral oil was used to control grape pests.

3- Pear, Pyrus communis L. were 2-3 m height and 10 years of age. Different weeds existed specially milky weeds. Pear defoliates during late January to February each year.

4- Peach, Prunus persica Stokes were 1.5-2 m height and 3 years of age. They were few trees planted on an irrigation canal side. No weeds or ground cover existed. No chemical control was applied.

5- Olive, Olea europea L. were 2-2.5 m height and 10-15 years of age. No ground cover or weeds associated with these trees.

6- Citrus (sour orange), Citrus aurantium L. was somewhat abundant grove. Trees were 2-3 m height and 7-10 years of age. Foliage was dense with no chemical control application. Little horticultural procedures were applied including weeding out the grove with harvesting the over-mature fruits and burning them.

7- Guava, Psidium guajava L. were few in numbers and existed next to buildings. No chemical control was applied and few weeds existed.

8- Mango, Mangifera indica L. were few in number, 3 m height and 7 years of age. No chemical control was applied and few weeds was observed.

9- Cotton, Gossypium barbadense L. were planted yearly and left in the field until the whole plants dry out in the fall. No chemical control was applied. Weeds existed between the cotton plants.

10- Maize, Zea mays L. were planted yearly during summer season and left with no chemical control over the growing season. Plants usually were left until the end of the year. In the plastic house, it was maintained around the year by continuous replantation. These plants were cultivated to rear grasshoppers on them (in glass cages).

11- Soybean, Glycine soja Sieb & Zucc. was planted yearly in the summer season. No chemical or weed control was conducted. Plants were usually left until drying in the fall.

12- Dadhi, Hypecorum perforatum L. were a dense fence on wire. No chemical control was applied. The fence was hedged from time to time and was cut back in January and allowed to grow back yearly.

13- Mulberry, Morus nigra Plam were few in number with dense foliage and existed near buildings. Few weeds grow up under these trees which was 3-3.5 m height and about 5 years old. No chemical control was applied to these trees.

14- Diafla, Nerium oleander Poir. were small bushes arranged as fence with 1.5 m height around buildings. No weeds existed around these bushes. No chemical control was applied.

15- Daisy, Camomille puante Mar. were flowering plants with about 0.5 m height and were planted in groups. These plants had a dense foliage. No weeds or chemical control existed.

B- Sampling using branch shaking :

1- Fruit trees : For fruit trees, five to six branches were shaken for each sample. Branches were sampled randomly. For grapes, a large amount of leaves were shaken over the shaking cloth.

2- Field crops : For field crops which included cotton and soybean, 10-15 plants were shaken for each sample, while for maize leaves of 5-6 plants were shaken over the shakig cloth.

3- Ornamental plants : Ten to twenty plants were shaken regardless of their flowering status.

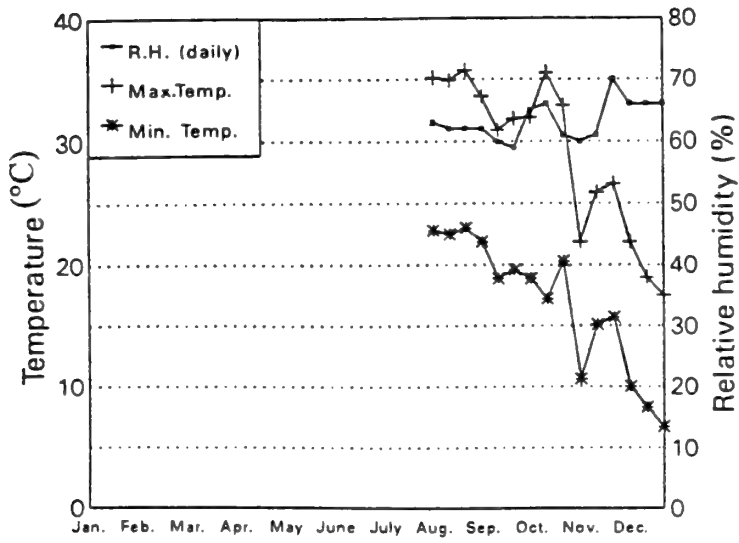
C- Sampling using pitfall traps :

Pitfall traps were used for all types of plantations. The pitfall traps consisted of glass bowls (5.5 cm diameter and 13 cm depth) into which 6 ml of foamy soap solution was used. Five traps were set at different places in each sampling site and checked biweekly during the surveying period.

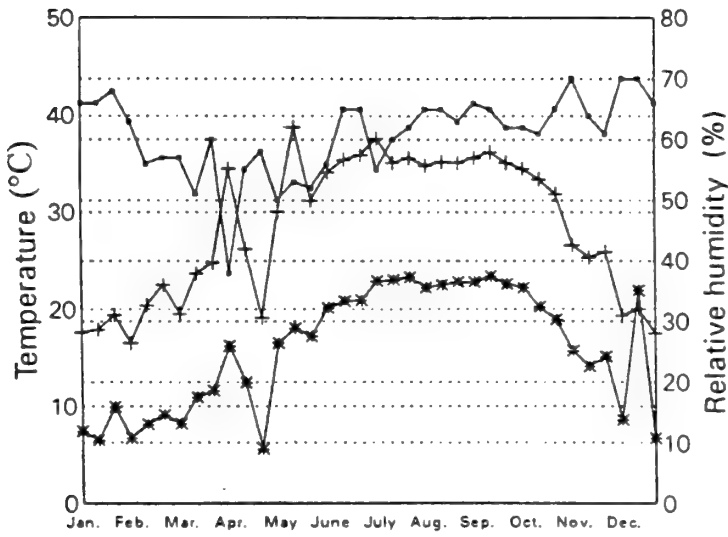
Table (1) : Sampling duration of different host plants used for spiders survey study.

Common	Scientific name	Sampling dates		
		1992	1993	1994
Apple	<i>Malus sylvestris</i> Mill	Aug. to Dec.	May to Dec.	May to Dec.
Grapes	<i>Vitis quadrangularis</i> Vall.	Aug. to Dec.	Jan., Apr. - Dec.	Jan., Apr. - Dec.
Pear	<i>Pyrus communis</i> L.	Aug. to Dec.	Jan., Mar. - Dec.	Jan., Mar. - Dec.
Peach	<i>Prunus persica</i> Stokes	Aug. to Dec.	Feb. to Dec.	Feb. to Dec.
Olive	<i>Olea europea</i> L.	Aug. to Dec.	Jan. to Dec.	Jan. to Dec.
Sour orange	<i>Citrus aurantium</i> L.	Aug. to Dec.	Jan. to Dec.	Jan. to Dec.
Guava	<i>Psidium guajava</i> L.	Aug. to Dec.	Feb. to Dec.	Feb. to Dec.
Mango	<i>Mangifera indica</i> L.	Aug. to Dec.	Jan. to Dec.	Jan. to Dec.
Cotton	<i>Gossypium barbadense</i> L.	Aug. to Dec.	Aug. to Dec.	Aug. to Dec.
Maize	<i>Zea mays</i> L.			
(open field)		Aug. to Dec.	June to Dec.	June to Dec.
(plastic houses)		-----	Feb. to Dec.	Feb. to Dec.
Soybean	<i>Glycine soja</i> Sieb & Zucc.	Sept. to Nov.	Sept. to Nov.	Sept. to Nov.
Dadhi	<i>Hypecorum perforatum</i> L.	Aug. to Dec.	Feb. to Dec.	Feb. to Dec.
Mulberry	<i>Morus nigra</i> Plam.	Aug. to Dec.	Feb. to Dec.	Feb. to Dec.
Diafla	<i>Nerium oleander</i> Poir.	Aug. to Dec.	Sept. to Dec.	Sept. to Dec.
Daisy	<i>Camomille puante</i> Mar.	Aug. to Dec.	Feb. to Mar.	Feb. to Mar.

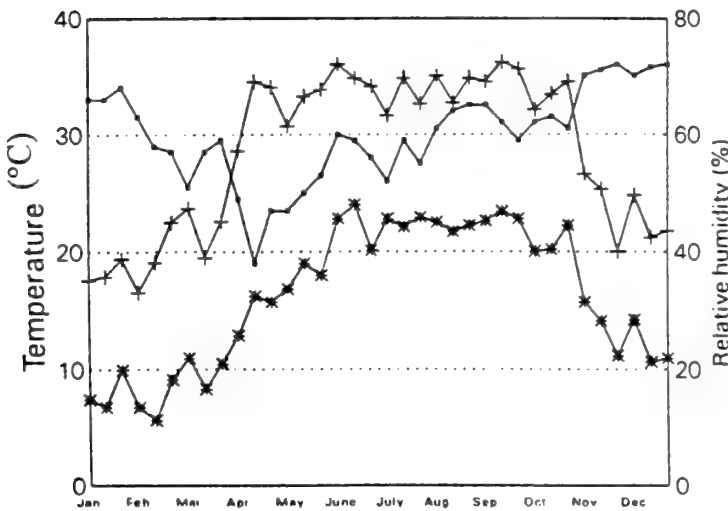
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1992



1993



1994

Fig. (1) : Mean weather (air) factors in Giza governorate during the survey study.

Results

Survey results are presented for the obtained and identified specimens regardless of gender or maturity stage. Reference was made for the most abundant families and species over each crop. The monthly variance of the most abundant families per crop are presented graphically. No graph was made when the total number collected was low (i.e. lower than 10 individuals). Variance of mean weather (air) factors in Giza Governorate during the survey study are illustrated in Fig. (1).

1- Apple trees :

Results of the survey study is shown in Table (2) and the monthly variance is illustrated in Fig. (2). Family Thomisidae was the most abundant one (Table 2). Thomisus spinifer Cambridge, 1872 was the most abundant species followed by Cheiracanthium jovium Denis, 1947. Monthly occurrence indicated that the most population was present in the second half of the year, although apple foliage occurs most around the year. No pitfall samples were obtained.

2- Grape vine :

Grape survey provided the highest number of spider population over the whole study (Table 2). Family Theridiidae and the species Euryopis acuminata (Lucas, 1846) were the most abundant ones. Family Thomisidae presented by T. spinifer and Xysticus sp. occupied the second most abundant category. Family Clubionidae represented by C. jovium occupied the third category (Table 2). It is obvious that spiders population occurred in grape during September to December yearly (Fig. 3).

3- Pear trees :

Survey results are presented in Table (2). Spiders population were second to grape vine. Family Clubionidae represented by C. jovium was the most abundant one, followed by family Thomisidae (i.e. T. spinifer and Xysticus sp.). Family Theridiidae was third in abundance (Table 2). Monthly variance indicated the same phenomena of presence of spiders mainly during September to December during the study duration (Fig. 4). Considerable numbers of family Lycosidae presented by Lycorma ferox (Lucas, 1838) were collected in pitfall traps (Table 2).

4- Peach trees :

Few individuals were collected from surveying peach trees (Table 2). All individuals belong to family Salticidae.

5- Olive trees :

Moderate spider numbers were obtained by surveying olive trees (Table 2). Family Clubionidae was represented by C. jovium, the most abundant.

6- Citrus trees :

Moderate spider populations were obtained by surveying citrus trees (Table 2). Family Clubionidae and family Uloboridae were the most abundant, presented by C. jovium and Uloborus walckenaerius Latreille, 1806, respectively.

7- Guava trees :

Few spider individuals were obtained by surveying trees (Table 2) where C.jovium was the most abundant species.

8- Mango trees :

Similar results to guava trees were obtained by surveying mango trees (Table 2).

9- Cotton plants :

Cotton plants which were surveyed over August to December over the three year period indicated that family Salticidae was the most abundant with Plexippus paykulli (Audouin, 1825) as the most abundant species (Table 2). Family Lycosidae which was collected using pitfall traps was the second in abundance represented by L.ferox. Fewer individuals were collected of family Philodromidae (Table 2). The species C.jovium also present but less abundant than in fruit trees (Table 2). Monthly population variance is illustrated in Fig.(5).

10- Maize plants :

Similar trend of spider species presence on cotton plants was obtained on maize in the open field (Table 2). Family Salticidae was the most abundant one with most numbers represented by P.paykulli. The spider C.jovium contributed as the second most abundant (Family : Clubionidae). Fewer numbers represented by family Thomisidae. Pitfall trapped individuals belong to three families (Table 2), with L.ferox (Family : Lycosidae) and Lycosoides coarctata (Dufour, 1831) (Family : Agelenidae) as the most abundant species. Similar results were obtained for maize under plastic house conditions (Table 2). Few individuals were present in the first half of the year compared with the second one (Figs 6 & 7).

11- Soybean :

Relative to cotton and maize smaller population of spiders occurred in soybean (Table 2). The species L.ferox (Family : Lycosidae) trapped in the pitfalls was the most abundant. Family Philodromidae is represented by Philodromus sp. and Thanatus albin (Audouin, 1825) were second in abundance (Fig.8).

12- Dadhi plants :

Dadhi plants provided moderate population of spiders (Table 2) and occurred mainly from August to October (Fig.9). Family Salticidae represented by Synageles sp. was the most abundant followed by Pholcidae and Uloboridae represented by Pholcus sp. & U.walckenaerius respectively.

13- Mulberry trees :

Mulberry trees was lightly occupied by spiders. A total of thirty six individuals was obtained during the study, twenty one of which were C.jovium (Table 2).

14- Diafla plants :

Diafla plants provided a total of fourteen individuals over the survey period (Table 2).

Table (2) : Spider individuals obtained by surveying different host plants in Giza.

Host plant	Family	Scientific name	Survey duration			Total
			1992	1993	1994	
1- <i>Malus sylvestris</i> (Apple)	1- Thomisidae	<i>Thomisus spinifer</i> Cambridge	30	30	25	85
		<i>Xysticus</i> sp.	17	10	15	42
	2- Clubionidae	<i>Cheiracanthium jovium</i> Denis	20	28	27	75
		<i>Synageles</i> sp.	1	2	4	7
	3- Salticidae	<i>Plexippus paykulli</i> (Audouin)	3	10	11	24
		<i>Thyene imperialis</i> (Rossi)	2	5	3	10
	4- Theridiidae	<i>Cyrtulina conspicua</i> (Cambridge)	-	2	1	3
		<i>Euryopsis acuminata</i> (Lucas)	8	9	12	29
		<i>Theridion</i> sp.	4	1	2	7
		<i>Cyrtophora citricola</i> (Forskål)	6	8	12	26
	5- Araneidae	<i>Lycorma ferox</i> (Lucas)	19	20	16	55
	6- Lycosidae*	<i>Erigone dentipalpis</i> (Wider)	2	1	2	5
		<i>Prinerigone vagans</i> (Savigny)	-	1	1	2
	7- Linyphiidae	<i>Dictyna</i> sp.	1	1	1	3
		<i>Philodromus</i> sp.	-	-	1	1
	8- Dictynidae	<i>Thanatus albi</i> (Audouin)	1	1	1	3
		<i>Uloborus walckenaerius</i> Latreille	2	1	-	3
	9- Philodromidae	-----	-	1	-	1
	10- Uloboridae					
	11- Filistatidae					

Table (2) : Cont'd.

Host plant	Family	Scientific name	Survey duration			Total
			1992	1993	1994	
2- <i>Vitis quadrangularis</i> (Grapes)	12- Agelenidae*	<i>Lycosoides coarctata</i> (Dufour)	-	1	-	1
	13- Gnaphosidae*	<i>Trachyzelotes</i> sp.	-	-	1	1
		<i>Zelotes</i> sp.	1	1	-	2
	1- Thomisidae	<i>Thomisus spinifer</i> Cambridge	30	40	45	115
		<i>Xysticus</i> sp.	24	21	30	75
	2- Clubionidae	<i>Cheiracanthium jovium</i> Denis	31	21	35	87
	3- Salticidae	<i>Synageles</i> sp.	2	3	4	9
		<i>Plexippus paykulli</i> (Audouin)	6	10	9	25
		<i>Thyene imperialis</i> (Rossi)	4	3	4	11
		<i>Cyrtulina conspicua</i> (Cambridge)	10	10	10	30
5- Araneidae 6- Dictynidae 7- Philodromidae	4- Theridiidae	<i>Euryopsis acuminata</i> (Lucas)	40	46	40	126
		<i>Theridion</i> sp.	12	10	14	36
	5- Araneidae	<i>Cyrtophora citricola</i> (Forskål)	4	9	8	21
	6- Dictynidae	<i>Dictyna</i> sp.	10	15	14	39
	7- Philodromidae	<i>Philodromus</i> sp.	1	-	2	3
		<i>Thanatus albini</i> (Audouin)	3	2	3	8

Table (2) : Cont'd.

Host plant	Family	Scientific name	Survey duration			Total
			1992	1993	1994	
3- <i>Pyrus communis</i> (Pear)	8- Gnaphosidae *	<i>Trachyzelotes sp.</i>	-	-	1	1
		<i>Zelotes sp.</i>	1	1	2	4
	9- Linyphiidae	<i>Erigone dentipalpis</i> (Wider)	1	2	1	4
		<i>Prinerigone vagans</i> (Savigny)	-	-	1	1
	10- Uloboridae	<i>Uloborus walckenaerius</i> Latreille	1	1	-	2
	11- Filistatidae	-----	-	1	1	2
	1- Thomisidae	<i>Thomisus spinifer</i> Cambridge	20	20	30	70
		<i>Xysticus sp.</i>	6	14	14	34
	2- Clubionidae	<i>Cheiracanthium jovium</i> Denis	58	66	67	191
	3- Salticidae	<i>Synageles sp.</i>	5	4	2	11
		<i>Plexippus paykulli</i> (Audouin)	4	5	4	13
		<i>Thyene imperialis</i> (Rossi)	9	10	12	31
4- Theridiidae		<i>Cyrtulina conspicua</i> (Cambridge)	2	4	2	8
		<i>Euryopsis acuminata</i> (Lucas)	10	15	22	57
		<i>Theridion sp.</i>	9	4	4	17
		<i>Cyrtophora citricola</i> (Forskål)	6	8	8	22
		<i>Lycorma ferox</i> (Lucas)	16	27	20	63

Table (2) : Cont'd.

Host plant	Family	Scientific name	Survey duration			Total
			1992	1993	1994	
	7- Linyphiidae	<i>Erigone dentipalpis</i> (Wider)	2	6	3	11
		<i>Prinerigone vagans</i> (Savigny)	-	2	2	4
	8- Dictynidae	<i>Dictyna</i> sp.	3	2	1	6
	9- Philodromidae	<i>Philodromus</i> sp.	-	1	1	2
		<i>Thanatus albini</i> (Audouin)	2	1	1	4
	10- Uloboridae	<i>Uloborus walckenaerius</i> Laterille	3	1	-	4
	11- Agelenidae*	<i>Lycosoides coarctata</i> (Dufour)	-	1	1	2
	12- Gnaphosidae *	<i>Trachyzelotes</i> sp.	-	-	1	1
		<i>Zelotes</i> sp.	1	1	1	3
	1- Salticidae	<i>Synageles</i> sp.	-	2	1	3
4- <i>Prunus persica</i> (Peach)		<i>Plexippus paykulli</i> (Audouin)	2	3	5	10
5- <i>Olea europea</i> (Olive)		<i>Thyene imperialis</i> (Rossi)	1	1	3	5
	1- Clubionidae	<i>Cheiracanthium jovium</i> Denis	4	8	7	19
	2- Philodromidae	<i>Philodromus</i> sp.	-	1	2	3
		<i>Thanatus albini</i> (Audouin)	2	1	4	7
	3- Salticidae	<i>Synageles</i> sp.	-	-	1	1
		<i>Plexippus paykulli</i> (Audouin)	-	1	2	3
		<i>Thyene imperialis</i> (Rossi)	5	2	2	9

Table (2) : Cont'd.

Host plant	Family	Scientific name	Survey duration			Total
			1992	1993	1994	
6- <i>Citrus aurantium</i> (Sour orange)	4- Linyphiidae	<i>Erigone dentipalpis</i> (Wider)	1	1	1	3
		<i>Prinerigone vagans</i> (Savigny)	-	1	-	1
	5- Uloboridae	<i>Uloborus walckenaerius</i> Latreille	1	5	6	12
	6- Theridiidae	<i>Cyrtulina conspicua</i> (Cambridge)	-	-	1	1
		<i>Euryopsis acuminata</i> (Lucas)	1	1	2	4
		<i>Theridion</i> sp.	-	-	1	1
	7- Araneidae	<i>Cyrtophora citricola</i> (Forskål)	-	1	3	4
	8- Oecobiidae	<i>Oecobius templi</i> Cambridge	1	-	1	2
		<i>Oecobius annulipes</i> Lucas	-	1	-	1
	9- Pholcidae	<i>Pholcus</i> sp.	1	-	1	2
	1- Clubionidae	<i>Cheiracanthium jovium</i> Denis	8	10	12	20
	2- Uloboridae	<i>Uloborus walckenaerius</i> Latreille	3	6	10	19
	3- Salticidae	<i>Synageles</i> sp.	-	1	1	2
		<i>Plexippus paykulli</i> (Audouin)	3	2	4	9
	<i>Thyene imperialis</i> (Rossi)	1	1	2	4	
4- Theridiidae	<i>Cyrtulina conspicua</i> (Cambridge)	-	-	2	2	
	<i>Euryopsis acuminata</i> (Lucas)	2	1	2	5	
	<i>Theridion</i> sp.	-	1	2	3	

Table (2) : Cont'd.

Host plant	Family	Scientific name	Survey duration			Total
			1992	1993	1994	
7- <i>Psidium guajava</i> (Guava)	5- Linyphiidae	<i>Erigone dentipalpis</i> (Wider)	2	1	1	4
		<i>Prinerigone vagans</i> (Savigny)	-	-	1	1
	6- Araneidae	<i>Cyrtophora citricola</i> (Forskål)	-	1	1	2
	7- Thomisidae	<i>Thomisus spinifer</i> Cambridge	-	1	1	2
		<i>Xysticus</i> sp.	-	1	1	2
	8- Philodromidae	<i>Philodromus</i> sp.	-	-	1	1
		<i>Thanatus albini</i> (Audouin)	1	1	1	3
	9- Dictynidae	<i>Dictyna</i> sp.	-	1	-	1
	10- Lycosidae*	<i>Lycorma ferox</i> (Lucas)	1	-	1	2
	1- Salticidae	<i>Synageles</i> sp.	-	-	1	1
		<i>Plexippus paykulli</i> (Audouin)	1	2	1	4
		<i>Thyene imperialis</i> (Rossi)	2	2	2	6
	2- Clubionidae	<i>Cheiracanthium jovium</i> Denis	2	5	8	15
	3- Philodromidae	<i>Philodromus</i> sp.	-	-	1	1
		<i>Thanatus albini</i> (Audouin)	1	1	-	2
	4- Araneidae	<i>Cyrtophora citricola</i> (Forskål)	1	-	-	1

Table (2) : Cont'd.

Host plant	Family	Scientific name	Survey duration			Total
			1992	1993	1994	
8- <i>Mangifera indica</i> (Mango)	1- Clubionidae	<i>Cheiracanthium jovium</i> Denis	2	5	8	15
	2- Thomisidae	<i>Thomisus spinifer</i> Cambridge	1	-	1	2
		<i>Xysticus</i> sp.	1	-	-	1
	3- Uloboridae	<i>Uloborus walckenaerius</i> Latreille	-	1	1	2
	4- Philodromidae	<i>Philodromus</i> sp.	-	1	-	1
		<i>Thanatus albinus</i> (Audouin)	1	-	1	2
		<i>Synageles</i> sp.	-	1	-	1
	5- Salticidae	<i>Plexippus paykulli</i> (Audouin)	1	1	-	2
		<i>Thyene imperialis</i> (Rossi)	-	1	1	2
		<i>Lycorma ferox</i> (Lucas)	20	23	24	67
9- <i>Gossypium</i> <i>barbadense</i> (Cotton)	1- Lycosidae*	<i>Cheiracanthium jovium</i> Denis	2	9	7	18
	2- Clubionidae	<i>Thomisus spinifer</i> Cambridge	2	1	-	3
	3- Thomisidae	<i>Xysticus</i> sp.	1	-	1	2
	4- Philodromidae	<i>Philodromus</i> sp.	5	3	4	12
		<i>Thanatus albinus</i> (Audouin)	6	10	8	24
		<i>Synageles</i> sp.	11	15	10	36
	5- Salticidae	<i>Plexippus paykulli</i> (Audouin)	40	30	30	100
		<i>Thyene imperialis</i> (Rossi)	20	20	10	50

Table (2) : Cont'd.

Host plant	Family	Scientific name	Survey duration			Total
			1992	1993	1994	
10- <i>Zea mays</i> (Maize) a- Open field	6- Araneidae	<i>Cyrtophora citricola</i> (Forskāl)	2	3	2	7
	7- Gnaphosidae*	<i>Trachyzelotes</i> sp.	-	1	1	2
		<i>Zelotes</i> sp.	1	1	3	5
	8- Theridiidae	<i>Cyrtulina conspicua</i> (Cambridge)	-	-	1	1
		<i>Euryopsis acuminata</i> (Lucas)	1	2	3	6
		<i>Theridion</i> sp.	-	1	-	1
		<i>Castianeira antinori</i> (Pavesi)	-	1	-	1
	9- Corinnidae*	<i>Lycorma ferox</i> (Lucas)	13	13	16	42
	1- Lycosidae*	<i>Cheiracanthium jovium</i> Denis	14	22	22	58
	2- Clubionidae	<i>Thomisus spinifer</i> Cambridge	2	6	8	16
		<i>Xysticus</i> sp.	-	4	5	9
	3- Thomisidae	<i>Lycosoides coarctata</i> (Dufour)	7	21	19	47
		<i>Synageles</i> sp.	10	8	10	28
	4- Agelenidae*	<i>Plexippus paykulli</i> (Audouin)	30	40	40	110
		<i>Thyene imperialis</i> (Rossi)	10	20	19	49
	5- Salticidae	<i>Cyrtulina conspicua</i> (Cambridge)	-	-	1	1
		<i>Euryopsis acuminata</i> (Lucas)	-	3	1	4
		<i>Theridion</i> sp.	-	1	-	1
	6- Theridiidae					

Table (2) : Cont'd.

Host plant	Family	Scientific name	Survey duration			Total
			1992	1993	1994	
b- Plastic houses	7- Dictynidae	<i>Dictyna sp.</i>	3	1	1	5
	8- Linyphiidae	<i>Erigone dentipalpus</i> (Wider)	2	1	3	6
		<i>Prinerigone vagans</i> (Savigny)	-	1	-	1
	9- Uloboridae	<i>Uloborus walckenaerius</i> Latreille	3	2	2	7
	10- Gnaphosidae*	<i>Trachyzelotes sp.</i>	-	-	1	1
		<i>Zelotes sp.</i>	2	1	1	4
	11- Filistatidae	- - - - -	-	1	-	1
	12- Philodromidae	<i>Philodromus sp.</i>	-	-	1	1
		<i>Thanatus albini</i> (Audouin)	1	-	1	2
	13- Dysderidae *	<i>Dysdera sp.</i>	-	-	1	1
	1- Lycosidae*	<i>Lycorma ferox</i> (Lucas)	15	23	32	70
	2- Clubionidae	<i>Cheiracanthium jovium</i> Denis	12	33	36	81
	3- Thomisidae	<i>Thomisus spinifer</i> Cambridge	2	15	16	33
<i>Xysticus sp.</i>		1	5	10	16	
4- Agelenidae*	<i>Lycosoides coarctata</i> (Dufour)	6	22	22	50	

Table (2) : Cont'd.

Host plant	Family	Scientific name	Survey duration			Total
			1992	1993	1994	
5- Salticidae		<i>Synageles sp.</i>	10	10	15	35
		<i>Plexippus paykulli</i> (Audouin)	20	50	60	130
		<i>Thyene imperialis</i> (Rossi)	13	30	15	58
6- Theridiidae		<i>Cyrtulina conspicua</i> (Cambridge)	-	-	1	1
		<i>Euryopsis acuminata</i> (Lucas)	1	1	1	3
		<i>Theridion sp.</i>	-	1	1	2
		<i>Dictyna sp.</i>	3	3	2	8
7- Dictynidae		<i>Erigone dentipalpus</i> (Wider)	2	3	2	7
8- Linyphiidae		<i>Prinerigone vagans</i> (Savigny)	-	-	1	1
		<i>Uloborus walckenaerius</i> Latreille	2	3	3	8
9- Uloboridae		<i>Trachyzelotes sp.</i>	-	-	1	1
10- Gnaphosidae *		<i>Zelotes sp.</i>	1	2	3	6
		<i>Dysdera sp.</i>	-	1	1	2
11- Dysderidae *		-----	-	-	1	1
12- Filistatidae		<i>Philodromus sp.</i>	-	-	1	1
13- Philodromidae		<i>Theridion albini</i> (Audouin)	-	1	1	2
			-	1	1	2

Table (2) : Cont'd.

Host plant	Family	Scientific name	Survey duration			Total
			1992	1993	1994	
11- <i>Glycine soja</i> (Soybean)	1- Lycosidae*	<i>Lycorna ferox</i> (Lucas)	7	18	20	45
	2- Clubionidae	<i>Cheiracanthium jovium</i> Denis	3	3	4	10
	3- Philodromidae	<i>Philodromus</i> sp.	3	6	9	17
		<i>Thanatus albini</i> (Audouin)	6	9	10	25
	4- Theridiidae	<i>Cyrtulina conspicua</i> (Cambridge)	1	1	-	2
		<i>Euryopsis acuminata</i> (Lucas)	3	4	7	14
		<i>Theridion</i> sp.	1	1	1	3
		<i>Synageles</i> sp.	-	2	1	3
	5- Salticidae	<i>Plexippus paykulli</i> (Audouin)	3	5	6	14
		<i>Thyene imperialis</i> (Rossi)	3	3	2	8
12- <i>Hypecorum perforatum</i> (Dadhi)	6- Thomisidae	<i>Thomisus spinifer</i> Cambridge	3	6	4	13
		<i>Xysticus</i> sp.	-	1	2	3
	7- Araneidae	<i>Cyrtophora citricola</i> (Forskål)	5	4	2	11
		<i>Synageles</i> sp.	17	19	16	52
	1- Salticidae	<i>Pholcus</i> sp.	8	11	8	27
	3- Clubionidae	<i>Cheiracanthium jovium</i> Denis	7	4	3	14
	4- Uloboridae	<i>Uloborus walckenaerius</i> Latreille	5	6	10	21

Table (2) : Cont'd.

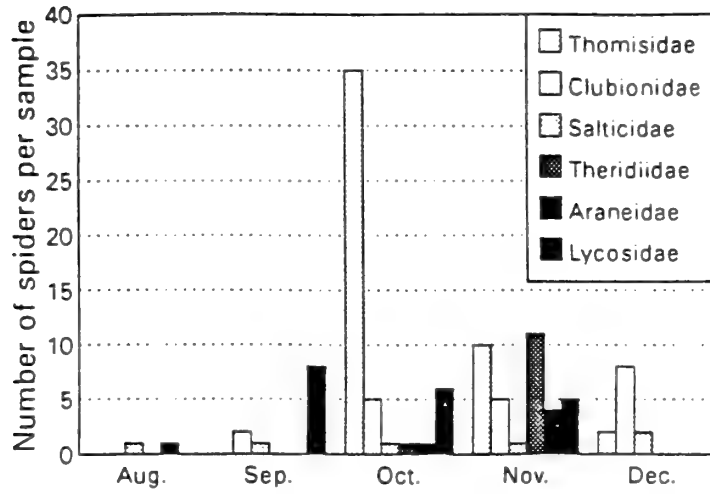
Host plant	Family	Scientific name	Survey duration			Total
			1992	1993	1994	
13- <i>Morus nigra</i> (Mulberry)	5- Philodromidae	<i>Thanatus albini</i> (Audouin)	2	4	5	11
	6- Theridiidae	<i>Euryopsis acuminata</i> (Lucas)	1	-	1	2
	7- Thomisidae	<i>Thomisus spinifer</i> Cambridge	1	-	-	1
	8- Dictynidae	<i>Dictyna</i> sp.	-	1	-	1
	9- Araneidae	<i>Cyrtophora citricola</i> (Forskål)	-	-	1	1
	10- Pisauridae	-----	-	1	-	1
	1- Clubionidae	<i>Cheiracanthium jovium</i> Denis	5	9	7	21
	2- Philodromidae	<i>Philodromus</i> sp.	-	2	4	6
	3- Araneidae	<i>Cyrtophora citricola</i> (Forskål)	1	3	2	6
	4- Linyphiidae	<i>Erigone dentipalpis</i> (Wider)	1	1	1	3
	5- Thomisidae	<i>Thomisus spinifer</i> Cambridge	-	1	-	1
		<i>Xysticus</i> sp.	-	-	1	1
	6- Oecobiidae	<i>Oecobius templi</i> Cambridge	1	-	1	2
	7- Lycosidae*	<i>Lycorma ferox</i> (Lucas)	-	1	1	2

Table (2) : Cont'd.

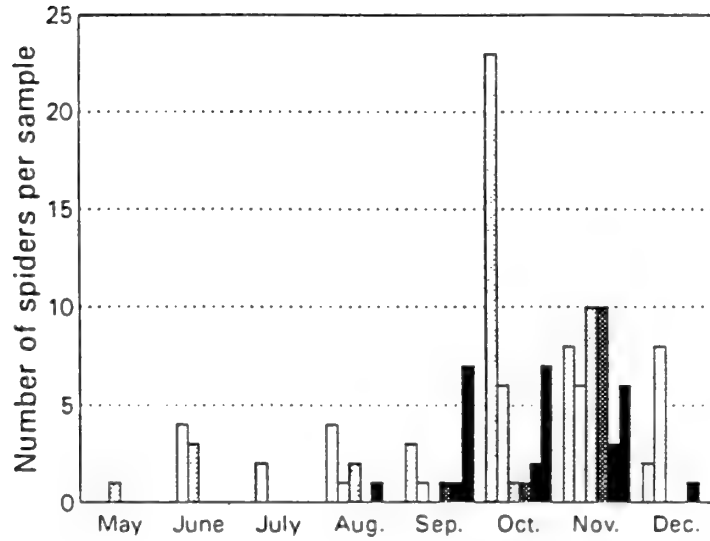
Host plant	Family	Scientific name	Survey duration			Total
			1992	1993	1994	
14- <i>Nerium oleander</i> (Diafla)	1- Salticidae	<i>Plexippus paykulli</i> (Audouin)	2	1	1	4
		<i>Thyene imperialis</i> (Rossi)	-	-	2	2
	2- Clubionidae	<i>Cheiracanthium jovium</i> Denis	1	2	-	3
	3- Theridiidae	<i>Cyrustulina conspicua</i> (Cambridge)	-	1	-	1
		<i>Euryopsis acuminata</i> (Lucas)	-	1	-	1
15- <i>Camomille</i> <i>puante</i> (Daisy)	4- Philodromidae	<i>Thanatus albini</i> (Audouin)	-	-	1	1
	5- Araneidae	<i>Cyrtophora citricola</i> (Forskål)	-	1	-	1
	1- Thomisidae	<i>Thomisus spinifer</i> Cambridge	1	5	4	10
	2- Clubionidae	<i>Cheiracanthium jovium</i> Denis	-	1	2	3
	3- Salticidae	<i>Synageles</i> sp.	5	2	-	7
	4- Pisauridae	- - - - -	-	1	1	2

* Pitfall traps

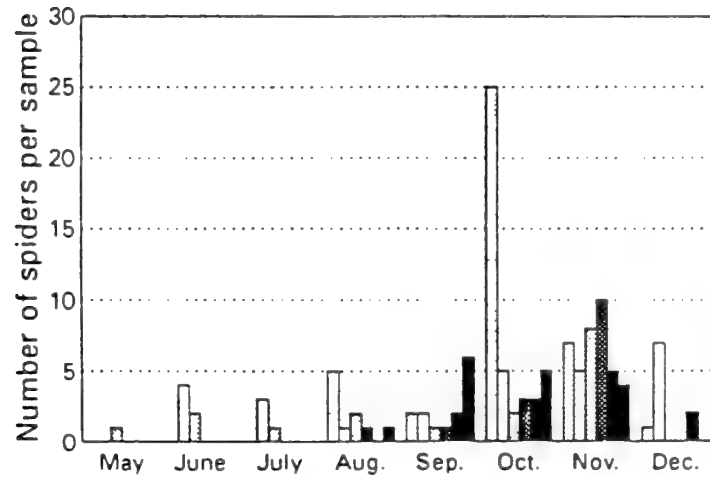
Apple



1992



1993



1994

Fig. (2): Monthly variance of different spider families occurred on apple trees during the survey study.

Grape

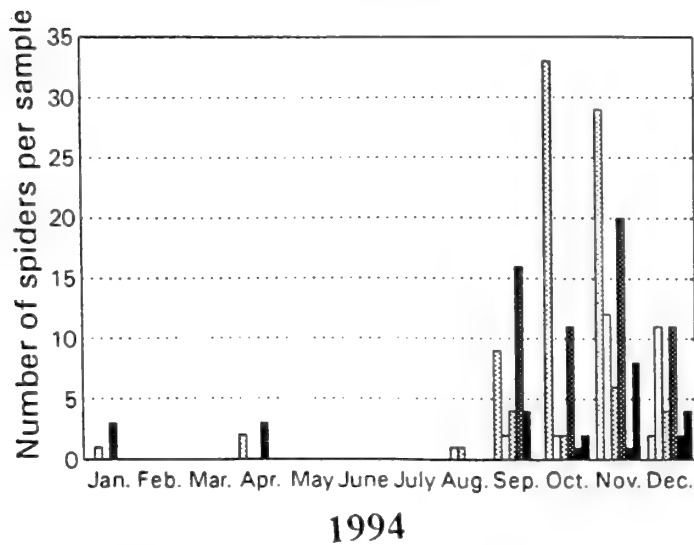
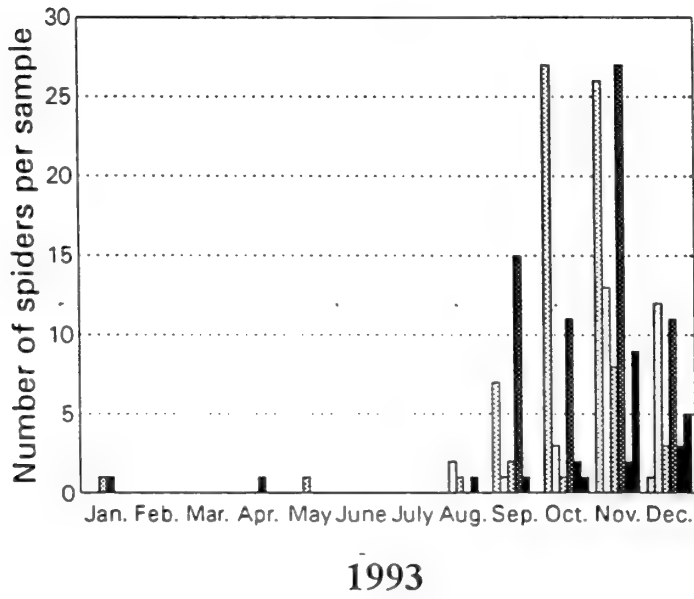
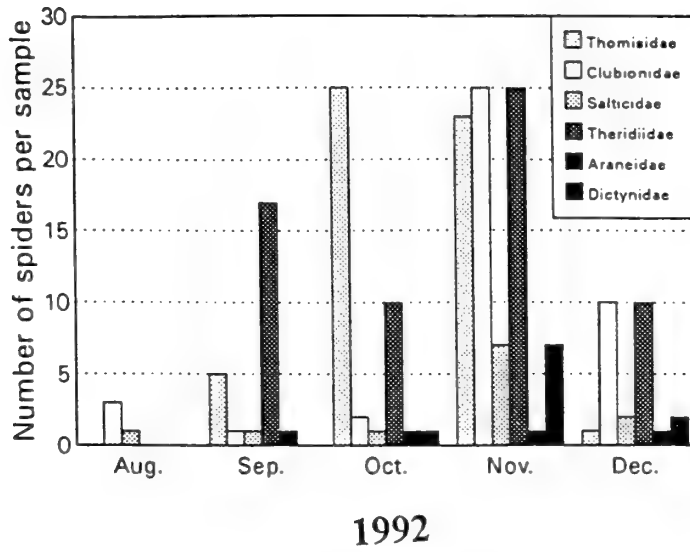
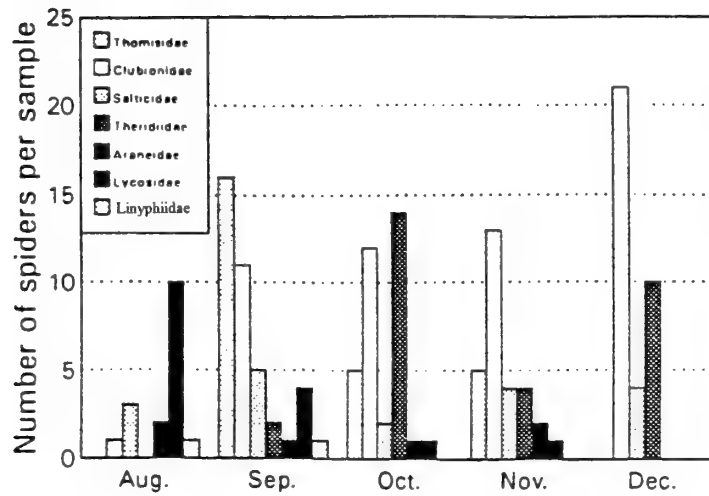
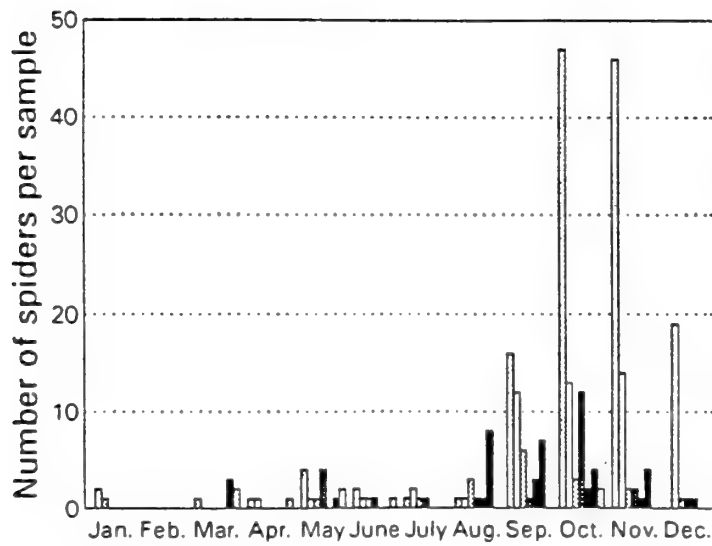


Fig. (3): Monthly variance of different spider families occurred on grape trees during the survey study.

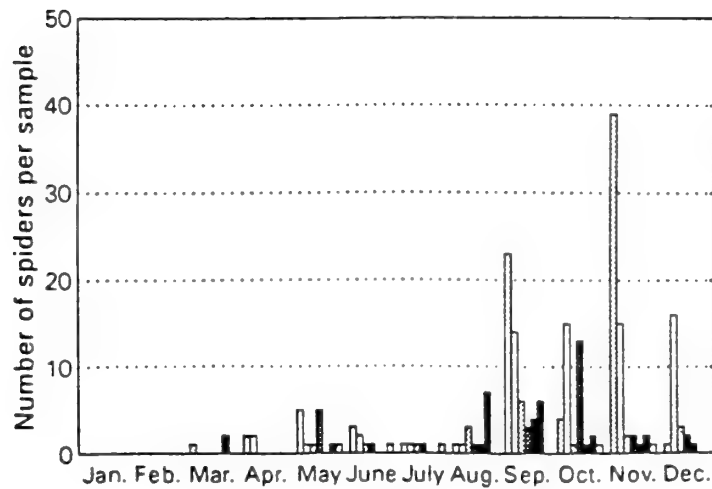
Pear



1992



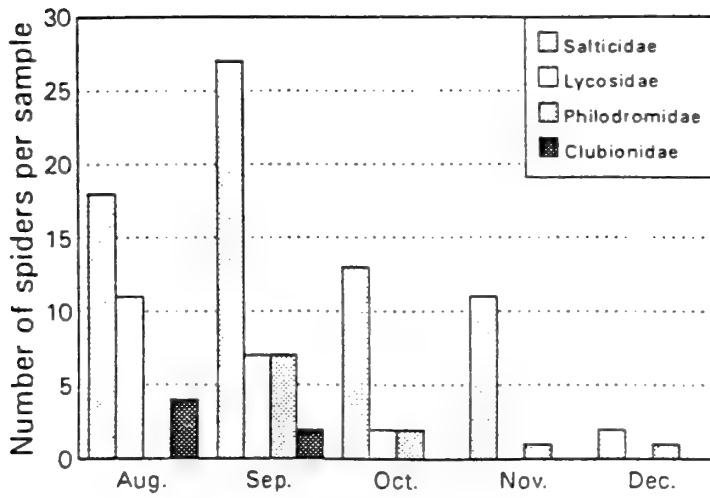
1993



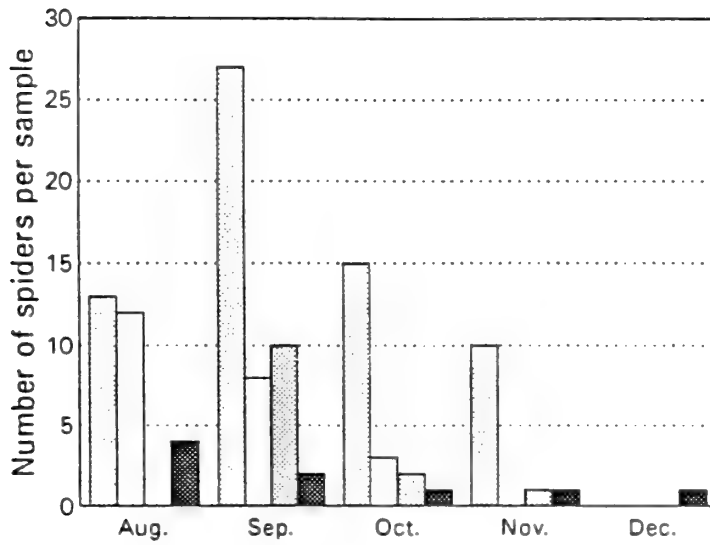
1994

Fig. (4): Monthly variance of different spider families occurred on pear trees during the survey study.

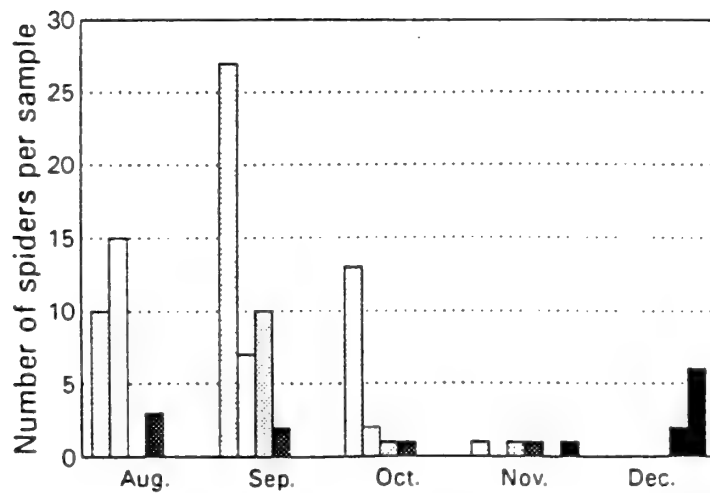
Cotton



1992



1993



1994

Fig. (5): Monthly variance of different spider families occurred on cotton plants during the survey study.

Maize (open field)

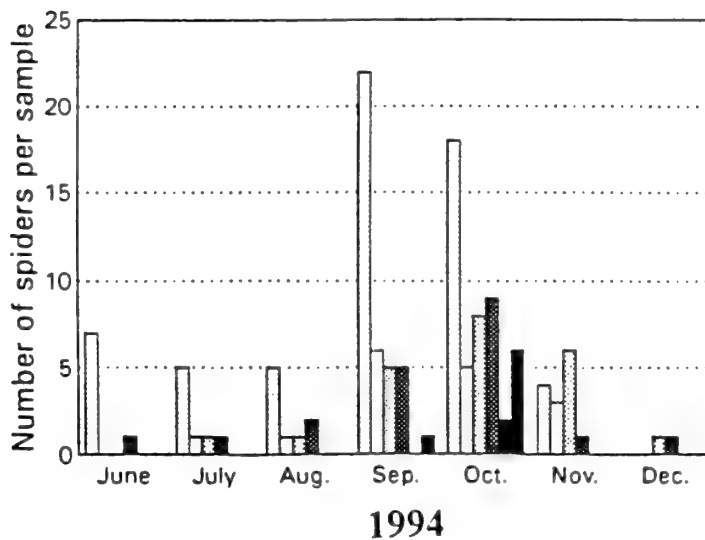
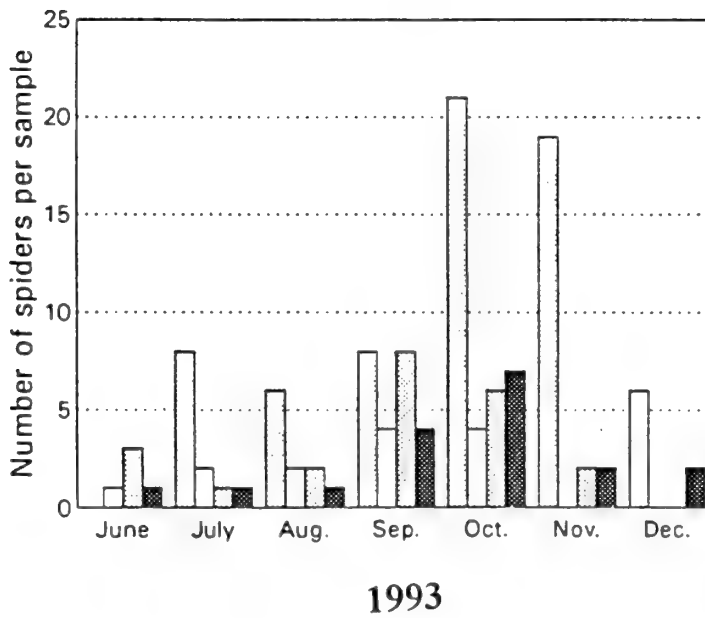
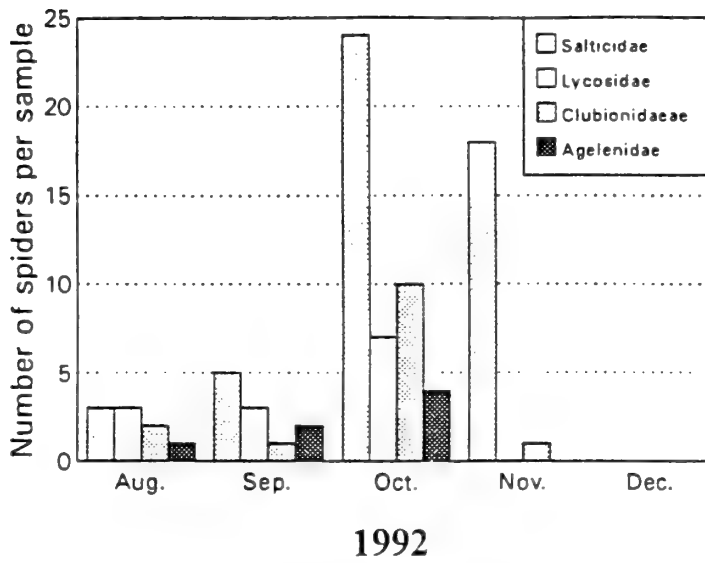
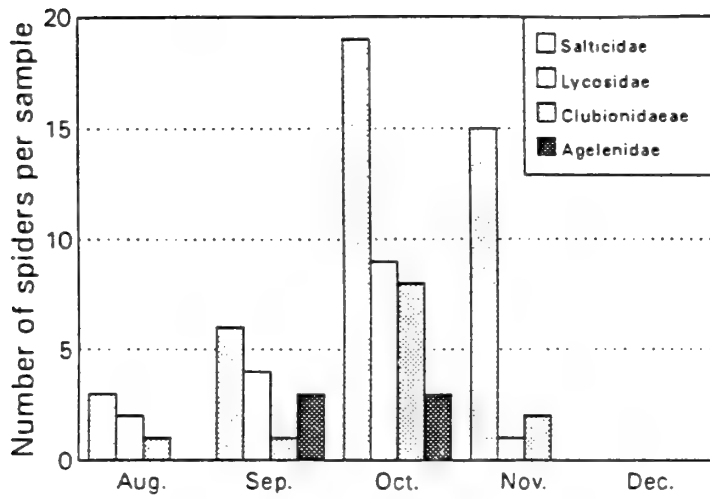
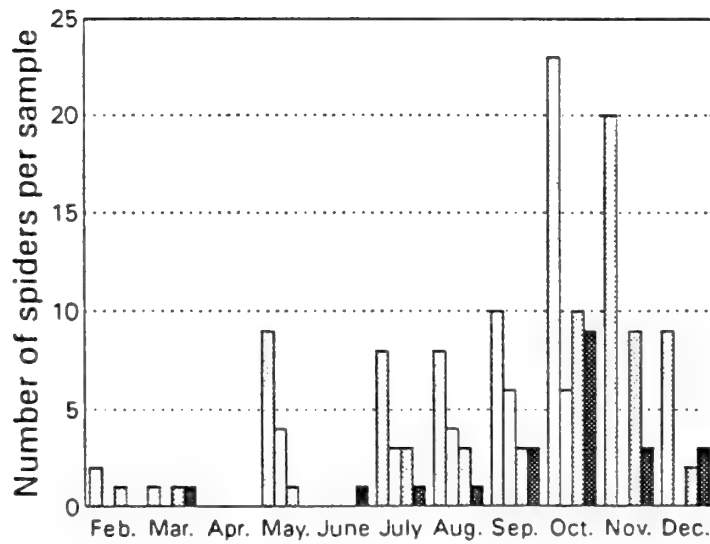


Fig. (6): Monthly variance of different spider families occurred on maize (open field) plants during the survey study.

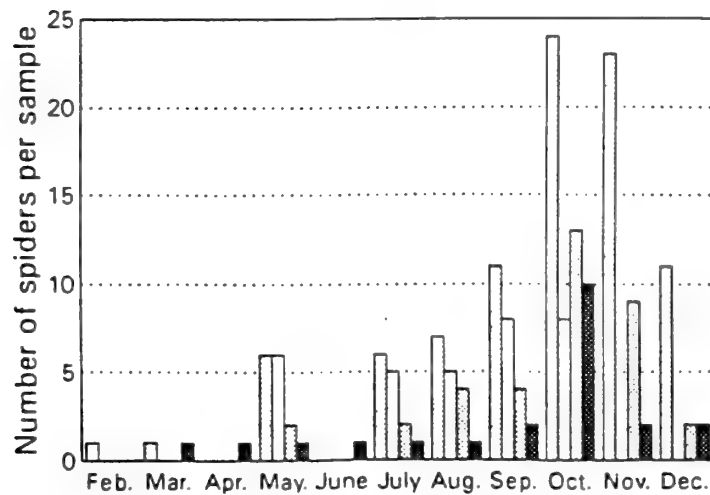
Maize (plastic house)



1992



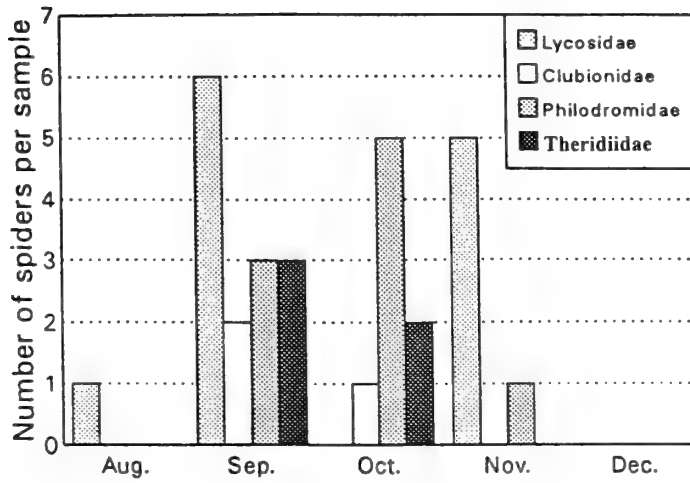
1993



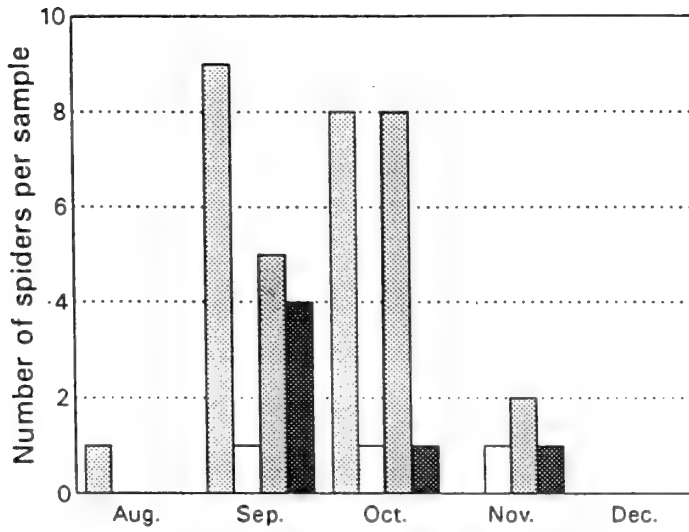
1994

Fig. (7): Monthly variance of different spider families occurred on maize (plastic houses) plants during the survey study.

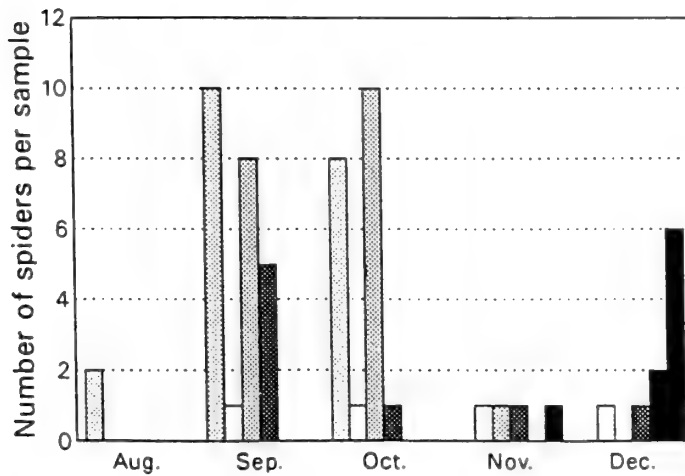
Soybean



1992



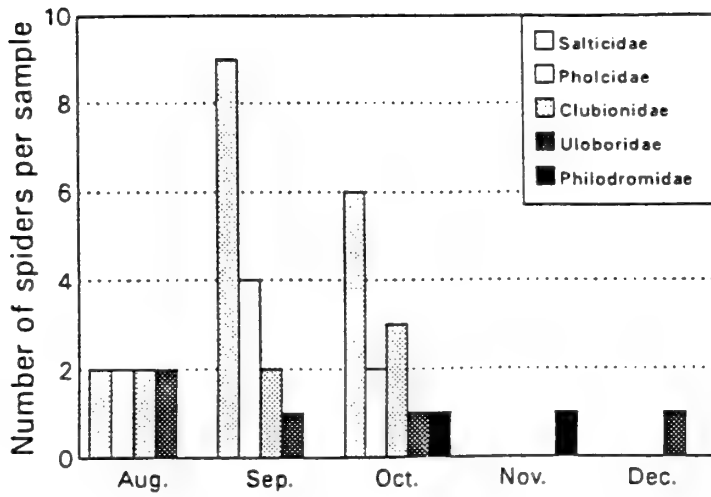
1993



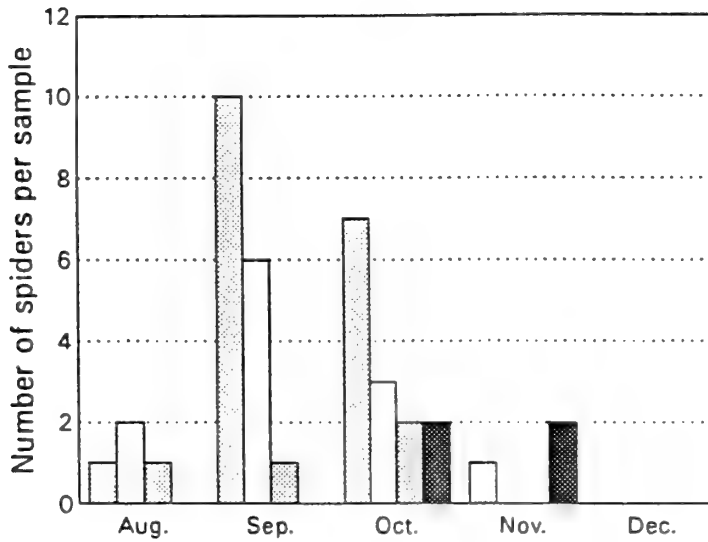
1994

Fig. (8): Monthly variance of different spider families occurred on soybean plants during the survey study.

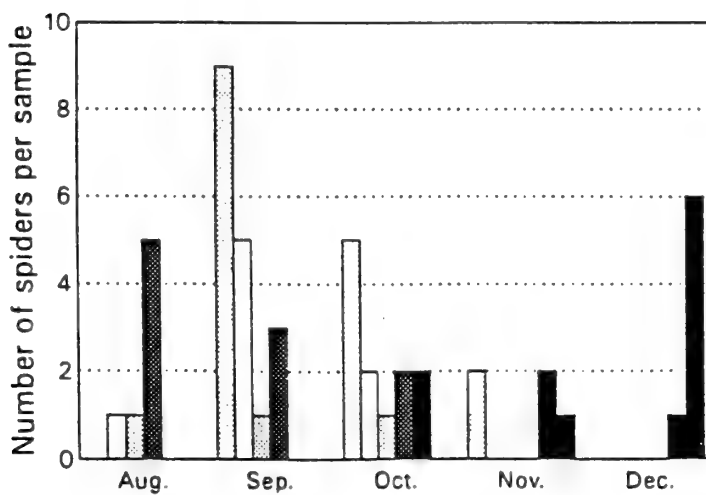
Dadhi



1992



1993



1994

Fig. (9): Monthly variance of different spider families occurred on dadhi plants during the survey study.

Table (3) : Occurrence of spiders (+) on different host plants.

Family	Scientific name	Host plants														
		1	2	3	4	5	6	7	8	9	10	11	12	13	14	15
Agelenidae	<i>Lycosoides coarctata</i>	+		+							+					
Araneidae	<i>Cyrtophora citreocola</i>	+	+	+		+	+	+		+		+	+		+	
Clubionidae	<i>Cheiracanthium jovium</i>	+	+	+		+	+	+	+	+	+	+	+	+	+	+
Corinnidae	<i>Castianeira antinori</i>									+						
Dictynidae	<i>Dictyna sp.</i>	+	+	+			+				+		+			
Dysderidae	<i>Dysdera sp.</i>										+					
Filistatidae	————	+	+								+					
Gnaphosidae	<i>Trachyzelotes sp.</i>	+	+	+						+	+					
	<i>Zelotes sp.</i>	+	+	+						+	+					
Linyphiidae	<i>Erigone dentipalpis</i>		+	+		+	+				+			+		
	<i>Prinerigone vagans</i>		+	+		+	+				+			+		
Lycosidae	<i>Lycorma ferox</i>	+		+			+			+	+	+		+		
Oecobiidae	<i>Oecobius templi</i>					+								+		
	<i>Oecobius annulipes</i>					+								+		
Philodromidae	<i>Philodromus sp.</i>	+	+	+		+	+	+	+	+	+	+	+			
	<i>Thanatus albini</i>	+	+	+		+	+	+	+	+	+	+	+	+	+	
Pholcidae	<i>Pholcus sp.</i>					+							+			
Pisauridae													+			+
Salticidae	<i>Synageles sp.</i>	+	+	+	+	+	+	+	+	+	+	+	+			+
	<i>Plexippus paykulli</i>	+	+	+	+	+	+	+	+	+	+	+			+	
	<i>Thyene imperialis</i>	+	+	+	+	+	+	+	+	+	+	+			+	
Theridiidae	<i>Cyrtulina conspicua</i>	+	+	+		+	+			+	+	+			+	
	<i>Euryopsis acuminata</i>	+	+	+		+	+			+	+	+	+		+	
	<i>Theridion sp.</i>	+	+	+		+	+			+	+	+				
Thomisidae	<i>Thomisus spinifer</i>	+	+	+			+		+	+	+	+	+	+		+
	<i>Xysticus sp.</i>	+	+	+			+		+	+	+	+		+		

Host plants :

- 1- Apple
- 2- Grape
- 3- Pear
- 4- Peach
- 5- Olive
- 6- Citrus
- 7- Guava
- 8- Mango

- 9- Cotton
- 10- Maize
- 11- Soybean
- 12- Dadhi
- 13- Mulberry
- 14- Diafla
- 15- Daisy

15- Daisy plants :

Daisy plants were inhabited by very few spiders. T. spinifer was the most collected species with total of ten individuals over the survey duration (Table 2).

Discussion

The survey study indicated diversity of spider fauna inhabiting surveyed plants. A summary of species occurrence is presented in Table (3). The thomisid Xysticus tristrani (Cambridge, 1872) was recorded in Egypt for the first time. Xysticus was found on nine out of fifteen plant crops surveyed (Table 3). Maize was the richest source of spiders (i.e. 21 species) followed by apple, grape and pear (i.e. 19 species). Ornamental plants provided the least source of spiders. The arboreal spiders belong to families: Thomisidae, Clubionidae and Salticidae were in order the most abundant on fruit trees, while the webbing family Theridiidae was less abundant. Family Lycosidae (i.e. wolf spiders) which inhabit the lower plant parts and ground surface and litter was the most abundant on cotton, maize and soybean with relative presence in apple and pear surveyed plants. In field crops (i.e. disturbed agriculture systems), family Salticidae was the most dominant followed by Lycosidae and Clubionidae.

Similar results were obtained in regard to stable and disturbed agriculture ecosystems (Dondale, 1956; Negm et al., 1976; Young & Edwards, 1990). No direct relation was obtained between a crop and a definite species, which reflects these spiders range of habitat and food sources. A relation could be established much more between plant foliage density and hence higher humidity and enclosed hiding places, occurrence of weeds, time of the year and spiders presence.

Unfortunately, the presence of other fauna was not considered in this study which represent an important part of the food chain. Aphids, fruitflies, lepidopteran and coleopteran larvae and tetranychid mites were noticed on the plants in association with spiders abundance during sampling. Seasonally, the fall had the highest contribution. This season was associated with higher temperature and relative humidity which also contributes to higher insect populations.

The role of these spiders was determined as non specific predators, which may have an important role in the agriculture ecosystem stability (Muma, 1975; Negm et al., 1976; Mansour et al., 1980). The slow rate of development of spiders reduces their ability of fast response to arthropod pest species in the agriculture ecosystem. Their long survival and diversity of species and habitat may mask their importance.

Acknowledgments

We thank Mr. H.K. El-Hennawy, for identifying spider specimens.

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J.Arachnol., 18(1): 1-27

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Serket = Sarkat
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Notes on Pseudopompilus humboldti (Dahlbom, 1845)

[Hymenoptera : Pompilidae] and Stegodyphus lineatus

(Latreille, 1817) [Araneida : Eresidae]

Hisham K. El-Hennawy

41, El-Manteqa El-Rabia St., Heliopolis,
Cairo, Egypt

Abstract

Pseudopompilus humboldti is a pompilid wasp which was recorded before from different localities in Egypt. Its relation with the eresid spider Stegodyphus dufouri was also studied.

Here, its relation with Stegodyphus lineatus in Egypt is dealt with. Also, the specificity of this wasp in attacking spiders is discussed trying to fill the gaps in information about its life cycle.

Key Words: Eresidae - Stegodyphus - Pompilidae - Pseudopompilus - Egypt - Distribution - Behaviour - Predation.

Introduction

In 1985, my first observations on the pompilid wasp Pseudopompilus humboldti (Dahlbom, 1845) had been published in a summarized form. The same observations had been published again entirely in 1987. Additional observations on the same species and its relation with the eresid spider Stegodyphus dufouri (Audouin, 1825) were published in 1986.

The wasp's prey was always S.dufouri, except in one experimental case when it was Stegodyphus lineatus (Latreille, 1817). Thence, the specificity of P.humboldti in predation was doubtful. Also, the life cycle along the year, or what the adult wasps do from June (time of cocoons hatching) until September (season of adult S.dufouri in Egypt), is a question which needs an answer.

This paper was presented in the XIIIth International Congress of Arachnology, Geneva, 3-8 September 1995.

The following notes are of two kinds : experimental (in laboratory) and observational (in nature). They will not solve the problem. They answer only a part of the question. They are dealing with the relation between P. humboldti and S. lineatus. A distribution map of P. humboldti, S. dufouri, and S. lineatus in Egypt is presented here.

Materials and Methods

Eight females and one male S. lineatus and two females P. humboldti were used as prey spiders and hunting wasps. Collecting specimens, keeping them alive in room temperature for biological study and rearing adult female wasps in a wooden box (50 x 50 x 50 cm) with two sides of glass for studying their predatory behaviour. Diluted bee honey was used for feeding the wasps. Illumination and temperature were increased a few minutes before putting a spider's nest (containing the spider within it) inside the wasp's cage by means of a 300 W lamp. A simple magnifying lens (3X) was used to observe the wasp's behaviour through it.

Results

I. Experimental notes :

1. July 4, 1980. A female wasp (hatched in captivity from a cocoon collected from Cairo) attacked a female S. lineatus (collected from Ras El-Barr, near Damietta). The wasp's egg was laid during night. It was devoured by the mother in the following day ?!

2. A female wasp hatched on June 18th 1982 (from a cocoon collected from Assiut, 23rd of December 1981) and lived until the 9th of July 1982. That wasp had made 14 attacks on 8 spiders. The results of those attacks are summarized below :

a. First attack (1st of July) on a male spider : stung, paralyzed; 4th leg not in position; no egg was laid.

b. Thirteen attacks on 7 females (of which : 6 attacks on 4 already paralyzed spiders). Eleven eggs were laid (of which : 3 were devoured, 1 became dry, and only 7 hatched and completed the life cycle until adults hatching).

II. Observational notes :

A. Locality : Ras El-Barr (about 500m from the Mediterranean sea coast).

1. Date : June 10, 1989

An area of 20 x 50 m was surveyed (10:30 - 13:15 h) to find :

- 17 female S. lineatus alive in their nests; some with egg sacs and only one with newly hatched spiderlings.

- 1 hatched cocoon and 1 unhatched cocoon (a female wasp hatched in captivity, 28th of June 1989) were found inside two S. lineatus nests with remnants of the spiders.
- Ratio of attacked spider nests to the total number of the adult spider nests : 2 : 19 (10.53 %).

2. Date : July 7, 1989

The same area was surveyed (17:15 - 18:30 h), [The reflection of sun-light on spiders' nests and webs, in that time of day, helped in finding the nests hidden among dry and green herbs.], to find :

- 13 nests of female S. lineatus with their spiderlings (6 of them with alive mother and 7 with a dead one).

[Two nests were nearly empty after the emergence of the spiderlings out of the nest. There was a gnaphosid spider inside one of those two nests. It's a predator on the spiderlings.]

- 2 empty cocoons; 1 deformed (in shape) unhatched cocoon [never hatched]; a 4th instar larva [completed its cocoon, July 10].

- 1 dry dead female spider with fourth leg in position (a small gnaphosid spider was found inside its moulting chamber beside the dead spider).

- Also, I could see, twice, a flying female P. humboldti ? But, only once, I could see it something near (2 m distant) and standing on the ground, but I could not catch it.

- Ratio : 4 : 18 (22.22 %).

[Total ratio : 6 : 37 (16.22 %)].

3. Date : August 19, 1989

- 1 empty cocoon was found in an old nest of S. lineatus in a place very near to the surveyed area. [Not included in the total ratio.]

B. Locality : St. Katherine (southern Sinai)

Date : June 24, 1989

During collecting samples of S. lineatus, found in nests attached to a stony wall of a building, Mr. Mobarak (of the Ecology Research Centre, Suez Canal University) discovered a pompilid cocoon inside one of the nests. He brought it to me thinking that it is a strange egg. The cocoon hatched on June 27th to give me my first sample of P. humboldti from Sinai.

Discussion

The experimental results are doubtful because the wasp was under obligatory conditions. Therefore, it attacked even paralyzed spiders (6 cases). But, these results lead us to say that it is probable that P. humboldti attacks S. lineatus in nature.

The specimens collected from Ras El-Barr and St. Katherine make it sure that P. humboldti attacks S. lineatus in nature. The yielded cocoons hatched in late June and early July. The female wasps attack S. lineatus again in early July. The wasps' activity increases during that time.

Finding P.humboldti in Ras El-Barr and St.Katherine extends the scope of distribution of this wasp in two directions, towards north until the mediterranean sea and towards east (in Sinai). P.humboldti was recorded formerly from southern Europe until Asia and Egypt (Haupt, 1927). It was considered rare in Egypt and recorded only from Suez and Luxor (Priesner, 1955 & 1960). Its distribution in Egypt was plotted on a map in my work of 1987. It is plotted here again (Fig. 1) with the distribution of S.dufouri and S.lineatus in Egypt for comparison (El-Hennawy, 1990 & 1992).

The relation between P.humboldti and S.lineatus was recorded before by Ward & Henschel (1992) in the Negev desert. They noticed that P.humboldti "stores its paralyzed host, the spider S.lineatus, at the entrance of the spider's nest...to prevent the wasp's host from overheating in this hot desert". It's the contrast in Ras El-Barr, near the Mediterranean sea coast, where the spider (Fig. 2) is always in the middle of her tubular nest. They described the attack of the wasp but unfortunately, they did not notice what the wasp had done by the fourth leg of the spider. A characteristic behaviour which was described and recorded in my works of 1985 and 1987. That behaviour which obliged me to add a new behavioural unit "Positioning" ("R") to Iwata's system of 1942 (Iwata, 1976) to say that the ethological type of P.humboldti is VPTR0. A modification which is recently approved by Shimizu (1994), who applied it on the similar case of Allochaes azureus and Filistata hibernalis (Deyrup et al., 1988).

Ward & Henschel (1992) found during 1990, that 25.6 % of the spiders in natural populations were parasitized. Figure which is near to that recorded from Ras El-Barr in early July, 1989 (22.22 %). That's in spite of the great difference between the two faunas of the arid Negev desert and the Mediterranean sea coast. Both their figure and mine are higher than that of S.dufouri (19.3 %) in the Nile valley (El-Hennawy, 1986). But, it is almost the same for the victim. The spider is devoured by the wasp's larva (Fig. 3) and her egg sac will not be opened. Then, the wasp do not only kill the mentioned ratio of spiders, but also kill a similar ratio of the next generation of that spider species (El-Hennawy, 1986).

Grout & Brothers (1982) mentioned that a male and a female of P.humboldti were reared (in the collections of the British Museum of Natural History) "from spiders nest" (Spain) and "ex cocoon on rose bush" (India). I think that the Spanish spider was S.lineatus and the Indian one was also a Stegodyphus sp. [4 species were recorded from India, two of them, S.pacificus and S.sarasinorum, are taxonomically related to S.dufouri and the other two, S.mirandus and S.tibialis are related to S.lineatus (Kraus & Kraus, 1988)].

Thence, P.humboldti is a predator of both S.dufouri and S.lineatus in Egypt. Also, it may be a predator of other species of the same genus "Stegodyphus" in India and other countries between southern Europe and Asia. In other words, it is not a species-specific pompilid wasp. It may be a "genus-specific" predator? The words of Wasbauer & Powell (1962) in their introduction, are almost applied in this case

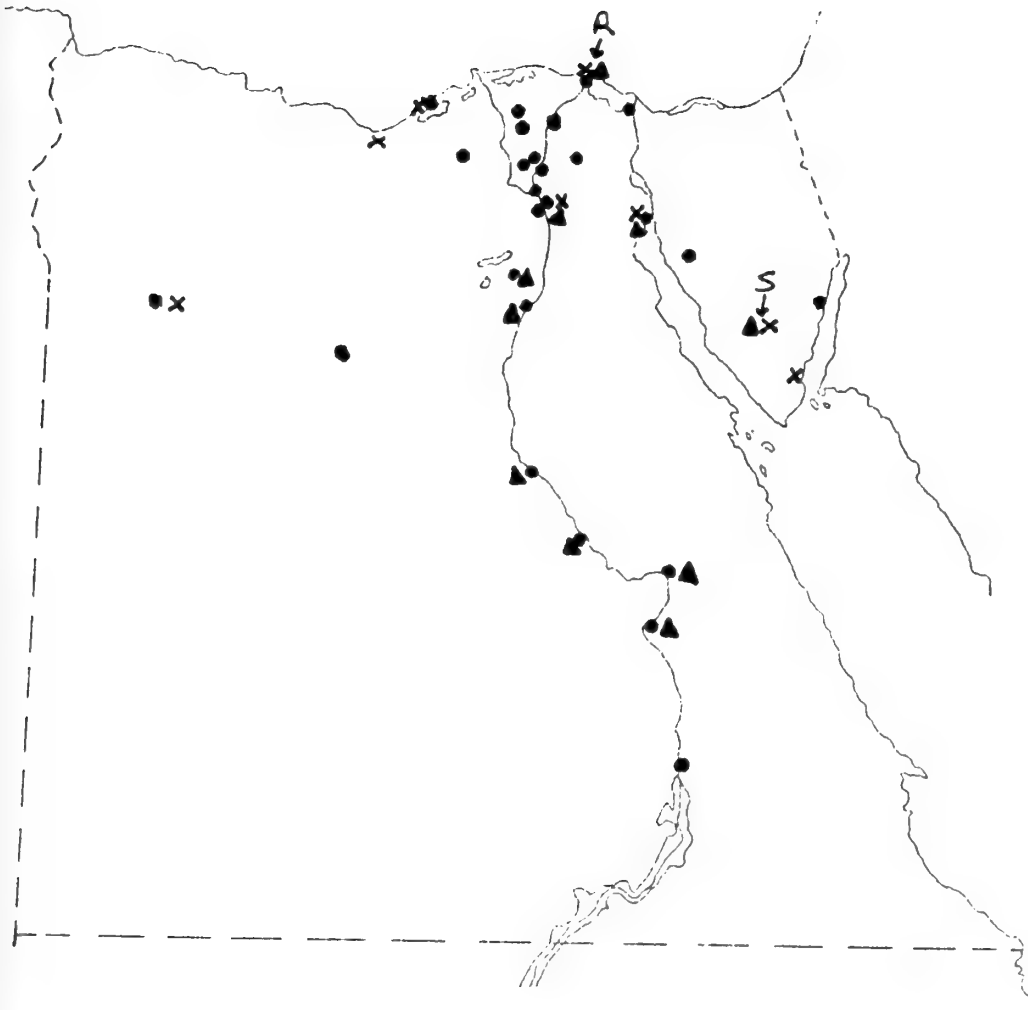


Fig. 1. Distribution map of Pseudopompilus humboldti (triangle), Stegodyphus lineatus (x), and Stegodyphus dufouri (circle) in Egypt. [R = Ras El-Barr; S = St. Katherine].

Fig. 2



Fig. 3



Fig. 2. Stegodyphus lineatus ♀ in her nest (Ras El-Barr, Egypt).

Fig. 3. 5th instar larva of Pseudopompilus humboldti devouring a female Stegodyphus lineatus inside her nest (St. Katherine).

"in general there is not much specificity, but often there is a tendency to specialize either on orb-weavers or on wandering spiders, or occasionally on one species of spider".

P.humboldti attacks S.dufouri in September-December and the next generation hatches, after long pupation period, in June to attack S.lineatus and continues actively the attack activity in early July. Hence, it is possible that S.lineatus is found all over the Nile valley (no records until now), or that those wasps attack other spider species in different regions. Also, what the wasps do after hatching in late July? Is there another victim in the period of July-September? Or is there another long pupation after devouring S.lineatus spiders?

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Introduction

This short bibliography is prepared to cover the years 1990-1995 and partly 1996. It is a complement to my "Bibliography of Pseudoscorpionida 1980-1989" (Serket 2(2), 1990) and also to Harvey's "Catalogue of the Pseudoscorpionida" (1990) [Bibliography: pp.10-128].

It includes 231 papers. Eighteen papers of the years 1987-1989 are here included to complete the previous bibliographies. The main source is the C.I.D.A.'s "Liste des Travaux Arachnologiques" (1991-1996), with reviewing the following periodicals and books:

Bulletin of the British Arachnological Society

	1990-1996	vol.8(4) - 10(6)
The Journal of Arachnology	1990-1996	vol.18(1) - 24(2)
Revue Arachnologique	1990-1996	vol.8(13) - 11(13)
Korean Arachnology	1990-1995	vol.5(2) - 11(2)
Arthropoda Selecta	1992-1995	vol.1(1) - 4(2)
Klapalekiana	1993-1996	vol.29(1-2) - 32(1-2)

Proceedings of the XIth International Congress of Arachnology,
Turku, Finland, 7-12 August 1989

Comptes rendus du XIIème Colloque européen d'Arachnologie,
Paris, 2-4 juillet 1990

Comptes rendus du XIIIème Colloque européen d'Arachnologie,
Neuchâtel, 2-6 septembre 1991

Proceedings of the 15th European Colloquium of Arachnology,
České Budějovice, Czech Republic, 11-15 July 1994

Proceedings of the XIIIth International Congress of Arachnology,
Geneva, 3-8 September 1995

The titles are arranged here alphabetically by the author name (and chronologically within this arrangement). Key words are bold typed to facilitate looking for papers of a special topic.

Acknowledgments

I thank all friends and colleagues who sent me their papers : Dr. Mark Harvey (Perth), Dr. Jacqueline Heurtault (Paris), and Dr. Volker Mahnert (Genève).

I thank too, my wife Rana who helped me so much and typed the material used in this work.

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 -- Another pseudoscorpion from Empire Cave, Santa Cruz County, California (Chthoniidae)
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pseudoscorpion, Cordylorchernes scorpioides
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On the function of harlequin beetle-riding in the
pseudoscorpion, Cordylorchernes scorpioides
(Pseudoscorpionida: Chernetidae).
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Dispersal-generated sexual selection in a beetle-riding
pseudoscorpion.
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Emergence of a giant fly triggers phoretic dispersal in the
neotropical pseudoscorpion, Semeiochernes armiger (Balzan)
(Pseudoscorpionida: Chernetidae).
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Failed predation or transportation ? Causes and consequences of
phoretic behavior in the pseudoscorpion Dinocheirus arizonensis
(Pseudoscorpionida : Chernetidae).
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pseudoscorpion (Cordylorchernes scorpioides).
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Cooperative foraging for large prey by Paratemnus elongatus
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armiger (Balzan) (Pseudoscorpionida: Chernetidae).
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AM. MUS. NAT. HIST. LIBRARY
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SERKET

Volume 5

Part 3

September, 1997

Cairo , Egypt

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I N D E X A R A N E A R U M

Part 5

(Hypochilidae, Austrochilidae, Gradungulidae, Filistatidae,
Sicariidae, Scytodidae, Drymusidae, Leptonetidae)

Hisham K. El-Hennawy

41, El-Manteqa El-Rabia St., Heliopolis,
Cairo, Egypt

I N D E X A R A N E A R U M

Part 5

Family **Hypochilidae**

Genus **Ectatosticta** Simon, 1892

E. davidi (Simon, 1888)

R2b p.1278 [2], P1 p.112 [2]

{1 sp.}

Genus **Hypochilus** Marx, 1888

H. bonneti Gertsch, 1964

B p.142 [1], P1 p.112 [1]

H. coylei Platnick, 1987

P1 p.112 [1], P2 p.115 [1]

H. gertschi Hoffman, 1963

B p.142 [2], P1 p.112 [3], P2 p.115 [1]

H. kastoni Platnick, 1987

P1 p.112 [1]

H. petrunkevitchi Gertsch, 1958

B p.142 [1], P1 p.112 [1]

H. pococki Platnick, 1987

P1 p.112 [1]

H. sheari Platnick, 1987

P1 p.112 [1]

H. thorelli Marx, 1888

R2b p.1278 [5], P1 p.112 [2]

{8 spp.}

Family **Austrochilidae**

Genus **Austrochilus** Gertsch & Zapfe, 1955

A. franckei Platnick, 1987

P1 p.113 [1]

A. manni Gertsch & Zapfe, 1955

B p.143 [1] [Thaida peculiaris] (F.Thaididae), P1 p.113 [1]

A. melon Platnick, 1987

P1 p.113 [1]

- A. newtoni Platnick, 1987
P1 p.113 [1]
A. schlingeri Platnick, 1987
P1 p.113 [1]

{5 spp.}

Genus **Hickmania** Gertsch, 1958

- H. troglodytes (Higgins & Petterd, 1883)
R2b p.1278 [4] [Ectatosticta t.] (F.Hypochilidae),
B p.143 [1] (F.Hickmaniidae), P1 p.113 [2], P2 p.116 [1]
{1 sp.}

Genus **Thaida** Karsch, 1880

- T. chepu Platnick, 1987
P1 p.113 [1]
T. peculiaris Karsch, 1880
R2a p.98 [2] (F.Agelenidae), P1 p.113 [2], P2 p.116 [1]
{2 spp.}

Family **Gradungulidae**

Genus **Gradungula** Forster, 1955

- G. soerenseni Forster, 1955
B p.143 [2]

{1 sp.}

Genus **Kaiya** Gray, 1987

- K. bemboka Gray, 1987
P1 p.113 [1]
K. brindabella (Moran, 1985)
P1 p.113 [2]
K. parnabyi Gray, 1987
P1 p.114 [1]
K. terama Gray, 1987
P1 p.114 [1]

{4 spp.}

Genus **Macrogradungula** Gray, 1987

- M. moonya Gray, 1987
P1 p.114 [1]

{1 sp.}

Genus **Pianoa** Forster, 1987

- P. isolata Forster, 1987
P1 p.114 [1]

{1 sp.}

Genus **Progradungula** Forster & Gray, 1979

- P. carraiensis Forster & Gray, 1979
P1 p.114 [3]

{1 sp.}

Genus **Spelungula** Forster, 1987

- S. cavernicola Forster, 1987
P1 p.114 [1]

{1 sp.}

Genus **Tarlina** Gray, 1987

- T. daviesae Gray, 1987
P1 p.114 [1]

- T. millidgei Gray, 1987
P1 p.114 [1]

- T. noorundi Gray, 1987
P1 p.114 [1]

- T. simipes Gray, 1987
P1 p.114 [1]

- T. smithersi Gray, 1987
P1 p.114 [1]

- T. woodwardi (Forster, 1955)
B p.143 [2] [Gradungula w.l], P1 p.114 [1]

{6 spp.}

Family **Filistatidae**Genus **Andoharano** Lehtinen, 1967

- A. decaryi (Fage, 1945)
R2b p.1280 [2] [Filistata d.], P2 p.116 [1]

- A. grandidieri (Simon, 1901)
R2b p.1280 [2] [Filistata g.], P2 p.116 [1]

- A. milloti Legendre, 1971
B p.144 [1]

- A. monodi Legendre, 1971
B p.144 [1]

{4 spp.}

Genus *Filistata* Latreille, 1810

- F. afghana* Roewer, 1962
B p.144 [1]
- F. annulipes* Kulczyński, 1908
R2b p.1278 [1]
- F. brignolii* Alayón, 1981
B p.673 [1], P1 p.115 [1]
- F. canariensis* Schmidt, 1976
B p.144 [1], P1 p.115 [1]
- F. chiardolae* Caporiacco, 1934
R2b p.1279 [1]
- F. delimbata* Strand, 1914
R2b p.1279 [1]
- F. fuscata* Kishida, 1943
B p.144 [1]
- F. gibsonhilli* Savory, 1943
R2b p.1281 [1]
- F. hebraea* Strand, 1914
R2b p.1279 [1]
- F. hebraea limbomaculata* Strand, 1914
R2b p.1279 [1]
- F. hirsuta* Cambridge, 1872
R2b p.1279 [1]
- F. insidiatrix* (Forskål, 1775)
R2b p.1280 [15], P1 p.115 [2]
- F. longiventris* Yaginuma, 1967
B p.144 [1]
- F. marginata* Kishida, 1936
B p.144 [2], P1 p.115 [1]
- F. napadensis* Patel, 1975
B p.144 [1]
- F. puta* Cambridge, 1876
R2b p.1279 [2]
- F. rufa* Caporiacco, 1934
R2b p.1279 [1]
- F. seclusa* Cambridge, 1885
R2b p.1279 [2]
- F. tarimuensis* Hu & Wu, 1989
P2 p.117 [1]
- F. xizanensis* Hu, Hu & Li, 1987
P1 p.115 [1], P2 p.117 [1]

{19 spp., 1 ssp.}

Genus *Filistatinella* Gertsch & Ivie, 1936

- F. crassipalpus* (Gertsch, 1935)
R2b p.1282 [2]

{1 sp.}

Genus **Filistatoides** F.O.P.-Cambridge, 1899

- F. fasciatus (Banks, 1902)
R2b p.1281 [1] [Filistata f.], P1 p.115 [1], P2 p.117 [1]
- F. insignis (O.P.-Cambridge, 1896)
R2b p.1281 [4] [Filistata i.], B p.144 [1] [Filistata polita],
P2 p.117 [2]
- F. mendensis (Mello-Leitão, 1920)
P2 p.117 [2]
- F. metamerica (Mello-Leitão, 1940)
R2b p.1281 [1] [Filistata m.], P2 p.117 [1]
- F. milloti (Zapfe, 1961)
B p.144 [1] [Filistata m.], P2 p.117 [1]

(5 spp.)

Genus **Kukulcania** Lehtinen, 1967

- K. arizonica (Chamberlin & Ivie, 1935)
R2b p.1282 [1] [Filistata a.], P2 p.118 [1]
- K. brevipes (Keyserling, 1882)
R2b p.1281 [1] [Filistata b.], P2 p.118 [1]
- K. geophila (Chamberlin & Ivie, 1935)
R2b p.1282 [1] [Filistata g.], P2 p.118 [1]
- K. geophila wawona (Chamberlin & Ivie, 1942)
R2b p.1282 [1] [Filistata g.w.]
- K. hibernalis (Hentz, 1842)
R2b pp.1281-1282 [16] [Filistata h.], P1 p.115 [4],
P2 p.118 [4]
- K. hurca (Chamberlin & Ivie, 1942)
R2b p.1282 [1] [Filistata h.], P2 p.118 [1]
- K. isolinae (Alayón, 1972)
B p.144 [2]
- K. tractans (O.P.-Cambridge, 1896)
R2b p.1281 [3] [Filistata t.], P2 p.118 [1]
- K. utahana (Chamberlin & Ivie, 1935)
R2b p.1282 [1] [Filistata u.], P2 p.118 [1]

(8 spp., 1 ssp.)

Genus **Malalistata** Mello-Leitão, 1946

- M. patagonica (Mello-Leitão, 1938)
R2b p.1282 [2]

(1 sp.)

Genus **Microfilistata** Zonstein, 1990

- M. tyshchenkoi Zonstein, 1990
P2 p.118 [1]

(1 sp.)

Genus *Pikelinia* Mello-Leitão, 1946

- P. kiliani* Müller, 1987
 P1 p.115 [1]
P. tambilloi (Mello-Leitão, 1941)
 R2b p.1282 [2] [*Pikeliana* t.]

(2 spp.)

Genus *Pritha* Lehtinen, 1967

- P. albimaculata* (O.P.-Cambridge, 1872)
 R2b p.1278 [1] [*Filistata* a.], P2 p.118 [1]
P. ampulla Wang, 1987
 P1 p.115 [1]
P. australiensis (L.Koch, 1873)
 R2b p.1281 [1] [*Filistata* a.], P2 p.118 [1]
P. bakeri (Berland, 1938)
 R2b p.1281 [1] [*Filistata* b.], P2 p.118 [1]
P. beijingensis Song, 1986
 P1 p.115 [2], P2 p.118 [2]
P. condita (O.P.-Cambridge, 1873)
 R2b p.1279 [2] [*Filistata* c.], P2 p.118 [1]
P. crosbyi (Spassky, 1938)
 R2b p.1279 [1] [*Filistata* c.], P2 p.118 [1]
P. dharmakumarsinhji Patel, 1978
 P1 p.115 [1]
P. fradei (Berland & Millot, 1940)
 R2b p.1280 [1] [*Filistata* f.], P2 p.118 [1]
P. garciai (Simon, 1892)
 R2b p.1280 [1] [*Filistata* g.], p.1281 [*F.pulchella*][1],
 P2 p.119 [1]
P. hasselti (Simon, 1906)
 R2b p.1281 [1] [*Filistata* h.], P2 p.119 [1]
P. heikki Saaristo, 1978
 B p.145 [1]
P. insularis (Thorell, 1881)
 R2b p.1281 [1] [*Filistata* i.], P2 p.119 [1]
P. lindbergi (Roewer, 1962)
 B p.144 [1] [*Filistata* l.], P2 p.119 [1]
P. littoralis (Roewer, 1938)
 R2b p.1281 [1] [*Filistata* l.], P2 p.119 [1]
P. nana (Simon, 1868)
 R2b p.1279 [4] [*Filistata* n.], [*F.debilis*][1], p.1280
 [*F.vestita*][3], P1 p.115 [1], P2 p.119 [6]
P. nicobarensis (Tikader, 1977)
 B p.144 [1] [*Filistata* n.], P1 p.115 [1]
P. pallida (Kulczyński, 1897)
 R2b p.1279 [3] [*Filistata* p.], P1 p.115 [1], P2 p.119 [1]
P. poonaensis (Tikader, 1963)
 B p.144 [1] [*Filistata* p.], P2 p.119 [1]

- P. sechellana Benoit, 1978
B p.145 [1]
P. spinula Wang, 1987
P1 p.116 [1]
P. sundaica (Kulczyński, 1908)
R2b p.1281 [1] [Filistata s.], P2 p.119 [1]
P. tenuispina (Strand, 1914)
R2b p.1279 [1] [Filistata t.], P1 p.116 [1]
P. zebrata (Thorell, 1895)
R2b p.1281 [1] [Filistata z.], P2 p.119 [1]
(24 spp.)

Genus **Sabastata** Benoit, 1968

- S. ashapuriae Patel, 1978
P1 p.116 [1]
S. nigra (Simon, 1897)
R2b p.1280 [4] [Filistata n.], [Filistata infuscata][1],
P2 p.119 [1]
S. sabaea Brignoli, 1982
B p.145 [1], P1 p.116 [1]

(3 spp.)

Genus *Tricalamus* Wang, 1987

- T. albidulus Wang, 1987
P1 p.116 [1]
T. longimaculatus Wang, 1987
P1 p.116 [1]
T. menglaensis Wang, 1987
P1 p.116 [1]
T. meniscatus Wang, 1987
P1 p.116 [1]
T. papilionaceus Wang, 1987
P1 p.116 [1]
T. papillatus Wang, 1987
P1 p.116 [1]
T. tetragonius Wang, 1987
P1 p.116 [1]
- (7 spp.)

Genus *Zaitunia* Lehtinen, 1967

- Z. alexandri Brignoli, 1982
B p.145 [1], P1 p.116 [1]
Z. beshkentica (Andreeva & Tyschchenko, 1969)
B p.144 [2] [Filistata b.], P2 p.119 [1]
Z. maracandica (Charitonov, 1946)
B p.144 [2] [Filistata m.], P2 p.119 [1]

- Z. martynovae (Andreeva & Tyschchenko, 1969)
B p.144 [1] [Filistata m.], P2 p.119 [1]
Z. medica Brignoli, 1982
B p.145 [1], P1 p.116 [1]
Z. monticola (Spassky, 1941)
R2b p.1279 [2] [Filistata m.], P2 p.119 [1]
Z. persica Brignoli, 1982
B p.145 [1], P1 p.116 [1]
Z. schmitzi (Kulczyński, 1911)
R2b p.1279 [1] [Filistata s.], P2 p.119 [1]
(8 spp.)

Family Sicariidae

Genus *Sicarius* Walckenaer, 1847

- S. albospinosus Purcell, 1908
R1 p.317 [2]
S. crustosus (Nicolet, 1849)
R1 p.317 [3]
S. damarensis Lawrence, 1928
R1 p.317 [1]
S. deformis (Nicolet, 1849)
R1 p.317 [2]
S. dolichocephalus Lawrence, 1928
R1 p.317 [1]
S. fumosus (Nicolet, 1849)
R1 p.318 [4]
S. gracilis (Keyserling, 1880)
R1 p.318 [3]
S. hahni (Karsch, 1878)
R1 p.317 [4]
S. lanuginosus (Nicolet, 1849)
R1 p.318 [2]
S. minoratus (Nicolet, 1849)
R1 p.318 [2]
S. nicoleti (Keyserling, 1880)
R1 p.318 [2]
S. patagonicus Simon, 1919
R1 p.318 [1], B p.145 [1] [*S. irregularis*], P1 p.116 [1]
S. peruensis (Keyserling, 1880)
R1 p.318 [2], P1 p.116 [1]
S. rubipes (Nicolet, 1849)
R1 p.318 [2]
S. rugosus (F.O.P.-Cambridge, 1899)
R1 p.318 [2], P1 p.116 [1]
S. rupestris (Holmberg, 1881)
R1 p.318 [3], P1 p.116 [1]
S. spatulatus Pocock, 1900
R1 p.317 [1]

- S. terrosus (Nicolet, 1849)
 R1 p.318 [4], P1 p.117 [1]
S. terrosus yurensis Strand, 1908
 R1 p.318 [1]
S. testaceus Purcell, 1908
 R1 p.317 [1]
S. tropicus (Mello-Leitão, 1936)
 R1 p.319 [1], P1 p.117 [1]
S. utrifomis (Butler, 1877)
 R1 p.319 [3]

{21 spp., 1 ssp.}

Genus Loxosceles Heineken & Lowe, 1835

- L. accepta Chamberlin, 1920
 R1 p.320 [1]
L. adelaida Gertsch, 1967
 B p.146 [2] (F.Scytodidae)
L. alamosa Gertsch & Ennik, 1983
 P1 p.118 [1] (F.Loxoscelidae)
L. alicia Gertsch, 1967
 B p.146 [1] (F.Scytodidae)
L. amazonica Gertsch, 1967
 B p.146 [1] (F.Scytodidae), P1 p.118 [1] (F.Loxoscelidae)
L. anomala (Mello-Leitão, 1917)
 R1 p.311 [1] [Calheirosia a.] (F.Leptonetidae)
L. apachea Gertsch & Ennik, 1983
 P1 p.118 [1] (F.Loxoscelidae)
L. aranea Gertsch, 1973
 B p.146 [1] (F.Scytodidae), P1 p.118 [1] (F.Loxoscelidae)
L. arizonica Gertsch & Mulaik, 1940
 B p.146 [2] (F.Scytodidae), P1 p.118 [1] (F.Loxoscelidae)
L. aurea Gertsch, 1973
 B p.146 [1] (F.Scytodidae), P1 p.118 [1] (F.Loxoscelidae)
L. baja Gertsch & Ennik, 1983
 P1 p.118 [1] (F.Loxoscelidae)
L. barbara Gertsch & Ennik, 1983
 P1 p.118 [1] (F.Loxoscelidae)
L. belli Gertsch, 1973
 B p.146 [1] (F.Scytodidae), P1 p.118 [1] (F.Loxoscelidae)
L. bettyae Gertsch, 1967
 B p.146 [1] (F.Scytodidae)
L. blancasi Gertsch, 1967
 B p.146 [1] (F.Scytodidae)
L. blanda Gertsch & Ennik, 1983
 P1 p.118 [1] (F.Loxoscelidae)
L. boneti Gertsch, 1958
 B p.146 [2] (F.Scytodidae), P1 p.118 [1] (F.Loxoscelidae)
L. candela Gertsch & Ennik, 1983
 P1 p.118 [1] (F.Loxoscelidae)

- L. caribbaea Gertsch, 1958
 B p.147 [1] (F.Scytodidae), P1 p.118 [1] (F.Loxoscelidae),
 P2 p.120 [1]
- L. carmena Gertsch & Ennik, 1983
 P1 p.118 [1] (F.Loxoscelidae)
- L. chinateca Gertsch & Ennik, 1983
 P1 p.119 [1] (F.Loxoscelidae)
- L. colima Gertsch, 1958
 B p.147 [1] (F.Scytodidae), P1 p.119 [1] (F.Loxoscelidae)
- L. conococha Gertsch, 1967
 B p.147 [1] (F.Scytodidae)
- L. coquimbo Gertsch, 1967
 B p.147 [1] (F.Scytodidae)
- L. coyote Gertsch & Ennik, 1983
 P1 p.119 [1] (F.Loxoscelidae)
- L. cubana Gertsch, 1958
 B p.147 [1] (F.Scytodidae), P1 p.119 [1] (F.Loxoscelidae)
- L. decemdentata Franganillo, 1926
 R1 p.319 [1]
- L. deserta Gertsch, 1973
 B p.147 [2] (F.Scytodidae), P1 p.119 [1] (F.Loxoscelidae)
- L. devia Gertsch & Mulaik, 1940
 B p.147 [2] (F.Scytodidae), p.146 [L.bolivarii][2],
 P1 p.119 [1] (F.Loxoscelidae)
- L. fontainei Millot, 1941
 B p.148 [1] (F.Scytodidae)
- L. foutadjalloni Millot, 1941
 B p.148 [1] (F.Scytodidae)
- L. francisca Gertsch & Ennik, 1983
 P1 p.119 [1] (F.Loxoscelidae)
- L. frizzelli Gertsch, 1967
 B p.147 [1] (F.Scytodidae)
- L. gaucho Gertsch, 1967
 B p.147 [2] (F.Scytodidae)
- L. gloria Gertsch, 1967
 B p.147 [1] (F.Scytodidae)
- L. guatemala Gertsch, 1973
 B p.147 [1] (F.Scytodidae), P1 p.119 [1] (F.Loxoscelidae)
- L. harrietae Gertsch, 1967
 B p.147 [1] (F.Scytodidae)
- L. herreri Gertsch, 1967
 B p.147 [1] (F.Scytodidae)
- L. hirsuta Mello-Leitão, 1931
 R1 p.320 [2]
- L. huasteca Gertsch & Ennik, 1983
 P1 p.119 [1] (F.Loxoscelidae)
- L. immodesta (Mello-Leitão, 1917)
 R1 p.311 [1] [Calheirosia i.] (F.Leptonetidae)
- L. inca Gertsch, 1967
 B p.147 [1] (F.Scytodidae)
- L. insula Gertsch & Ennik, 1983
 P1 p.119 [1] (F.Loxoscelidae)

- L. intermedia Mello-Leitão, 1934
R1 p.320 [1], p.321 [L.ornata][1], P2 p.120 [1]
- L. jaca Gertsch & Ennik, 1983
P1 p.119 [1] (F.Loxoscelidae)
- L. jamaica Gertsch & Ennik, 1983
P1 p.119 [1] (F.Loxoscelidae)
- L. jarmila Gertsch & Ennik, 1983
P1 p.119 [1] (F.Loxoscelidae)
- L. julia Gertsch, 1967
B p.147 [1] (F.Scytodidae)
- L. kaiba Gertsch & Ennik, 1983
P1 p.119 [1] (F.Loxoscelidae)
- L. lacroixi Millot, 1941
B p.148 [1] (F.Scytodidae)
- L. laeta (Nicolet, 1849)
R1 p.320 [6], [L.longipalpis][1], p.321 [L.nesophila][1],
[Omosita bicolor, under L.rufipes], B p.146 [L.yura][1],
P1 p.119 [2] (F.Loxoscelidae), P2 p.120 [1]
- L. lawrencei Caporiacco, 1955
B p.147 [3] (F.Scytodidae)
- L. lutea Keyserling, 1877
R1 p.321 [1], [L.pictithorax][1], P2 p.120 [1]
- L. luteola Gertsch, 1973
B p.147 [1] (F.Scytodidae), P1 p.119 [1] (F.Loxoscelidae)
- L. manuela Gertsch & Ennik, 1983
P1 p.119 [1] (F.Loxoscelidae)
- L. martha Gertsch & Ennik, 1983
P1 p.119 [1] (F.Loxoscelidae)
- L. meruensis Tullgren, 1910
R1 p.320 [1]
- L. misteca Gertsch, 1958
B p.147 [1] (F.Scytodidae), P1 p.119 [1] (F.Loxoscelidae)
- L. mulege Gertsch & Ennik, 1983
P1 p.119 [1] (F.Loxoscelidae)
- L. nahuana Gertsch, 1958
B p.147 [1] (F.Scytodidae), P1 p.119 [1] (F.Loxoscelidae)
- L. neuvillei Simon, 1909
R1 p.320 [2]
- L. olmea Gertsch, 1967
B p.147 [1] (F.Scytodidae)
- L. pallidecolorata (Strand, 1906)
R1 p.321 [2] [Loxoscella p.]
- L. palma Gertsch & Ennik, 1983
P1 p.119 [1] (F.Loxoscelidae)
- L. panama Gertsch, 1958
B p.147 [2] (F.Scytodidae), P1 p.119 [1] (F.Loxoscelidae)
- L. parrami Newlands, 1981
P1 p.119 [1] (F.Loxoscelidae)
- L. piura Gertsch, 1967
B p.148 [1] (F.Scytodidae)
- L. pucara Gertsch, 1967
B p.148 [1] (F.Scytodidae)

- L. reclusa Gertsch & Mulaik, 1940
B p.148 [2] (F.Scytodidae), P1 p.119 [1] (F.Loxoscelidae)
- L. rica Gertsch & Ennik, 1983
P1 p.120 [1] (F.Loxoscelidae)
- L. rosana Gertsch, 1967
B p.148 [1] (F.Scytodidae)
- L. rothi Gertsch & Ennik, 1983
P1 p.120 [1] (F.Loxoscelidae)
- L. rufescens (Dufour, 1820)
R1 p.319 [17], [L.compactilis][1], [L.distincta][4],
B p.146 [L.indrabeles][1], [L.marylandica][1],
P1 p.120 [6] (F.Loxoscelidae), P2 p.120 [4]
- L. rufescens lucifuga Simon, 1910
R1 p.319 [1]
- L. rufipes (Lucas, 1834)
R1 p.321 [9], P1 p.120 [1] (F.Loxoscelidae), P2 p.120 [1]
- L. russelli Gertsch & Ennik, 1983
P1 p.120 [1] (F.Loxoscelidae)
- L. sabina Gertsch & Ennik, 1983
P1 p.120 [1] (F.Loxoscelidae)
- L. seri Gertsch & Ennik, 1983
P1 p.120 [1] (F.Loxoscelidae)
- L. similis Moenkhaus, 1898
R1 p.321 [3] [L.surata], P2 p.120 [1] (not L.laeta)
- L. smithi Simon, 1902
R1 p.320 [1]
- L. sonora Gertsch & Ennik, 1983
P1 p.120 [1] (F.Loxoscelidae)
- L. spadicea Simon, 1907
R1 p.321 [1]
- L. speluncarum Simon, 1893
R1 p.320 [2]
- L. spinulosa Purcell, 1904
R1 p.320 [1], [L.pilosa][1], [L.simillima][1],
p.319 [L.bergeri][1], B p.146 [L.spiniceps][1], P2 p.120 [1]
- L. surca Gertsch, 1967
B p.148 [1] (F.Scytodidae)
- L. taeniopalus Simon, 1907
R1 p.321 [1]
- L. taino Gertsch & Ennik, 1983
P1 p.120 [1] (F.Loxoscelidae)
- L. tehuana Gertsch, 1958
B p.148 [1] (F.Scytodidae), P1 p.120 [1] (F.Loxoscelidae)
- L. tenango Gertsch, 1973
B p.148 [1] (F.Scytodidae), P1 p.120 [1] (F.Loxoscelidae)
- L. teresa Gertsch & Ennik, 1983
P1 p.120 [1] (F.Loxoscelidae)
- L. tlacolula Gertsch & Ennik, 1983
P1 p.120 [1] (F.Loxoscelidae)
- L. unicolor Keyserling, 1887
P2 p.121 [1] (not L.rufipes)

- L. valdosa Gertsch, 1973
B p.148 [2] (F.Scytodidae), P1 p.120 [1] (F.Loxoscelidae)
L. valida Lawrence, 1964
B p.148 [2] (F.Scytodidae)
L. variegata Simon, 1897
R1 p.321 [1]
L. virgo Gertsch & Ennik, 1983
P1 p.120 [1] (F.Loxoscelidae)
L. vonwredei Newlands, 1980
B p.674 [2], P1 p.120 [1] (F.Loxoscelidae)
L. weyrauchi Gertsch, 1967
B p.148 [1] (F.Scytodidae)
L. yucatana Chamberlin & Ivie, 1938
R1 p.321 [1], P1 p.120 [1] (F.Loxoscelidae)
L. zapoteca Gertsch, 1958
B p.148 [1] (F.Scytodidae), P1 p.120 [1] (F.Loxoscelidae)
(99 spp., 1 ssp.)

Family Scytodidae

[In R1 it is considered a subfamily of the Sicariidae.]

Genus **Scytodes** Latreille, 1804

- S. aethiopica Simon, 1907
R1 p.325 [1]
S. affinis Kulczyński, 1901
R1 p.325 [2]
S. aguapeyana Mello-Leitão, 1945
B p.149 [1]
S. aharoni Strand, 1914
R1 p.324 [1]
S. alayoi Alayón, 1977
B p.149 [1]
S. albiapicalis Strand, 1907
R1 p.327 [1]
S. annulata Keyserling, 1891
R1 p.329 [1]
S. arabica (Simon, 1890)
R1 p.325 [1]
S. arboricola Millot, 1946
B p.149 [1]
S. arenacea Purcell, 1904
R1 p.325 [2]
S. aruensis Strand, 1911
R1 p.329 [1]
S. bergeri Strand, 1915
R1 p.325 [1]
S. bertheloti Lucas, 1838
R1 p.324 [2]

- S. bertheloti annulipes Simon, 1907
R1 p.324 [1]
- S. blanda Bryant, 1940
B p.149 [1]
- S. broomi Pocock, 1902
R1 p.325 [2]
- S. caffra Purcell, 1904
R1 p.325 [2]
- S. camerunensis Strand, 1906
R1 p.325 [1]
- S. canariensis Wunderlich, 1987
P1 p.117 [1]
- S. cavernarum Roewer, 1962
B p.149 [1]
- S. cedri Purcell, 1904
R1 p.325 [1]
- S. cellularis Simon, 1907
R1 p.325 [1]
- S. chamberlini Caporiacco, 1955
B p.149 [1]
- S. championi F.O.P.-Cambridge, 1899
R1 p.329 [1], P1 p.117 [1]
- S. clavata Benoit, 1965
B p.149 [1]
- S. concolor Mello-Leitão, 1918
R1 p.329 [1]
- S. congoanus Strand, 1908
R1 p.325 [1]
- S. constellata Lawrence, 1938
R1 p.325 [1]
- S. coronata Thorell, 1899
R1 p.325 [2]
- S. cubensis Alayón, 1977
B p.149 [1]
- S. darlingtoni Alayón, 1977
B p.149 [1]
- S. depressiventris Mello-Leitão, 1916
R1 p.329 [2]
- S. diminuta Valerio, 1981
B p.674 [1], P1 p.117 [1]
- S. dissimulans Petrunkevitch, 1929
R1 p.329 [1]
- S. dollfusi Millot, 1941
B p.149 [1]
- S. dorothea Gertsch, 1935
R1 p.329 [1]
- S. drakensbergensis Lawrence, 1947
B p.149 [1]
- S. elizabethae Purcell, 1904
R1 p.325 [1]
- S. farri Alayón, 1985
P1 p.117 [1]

- S. flagellata Purcell, 1904
R1 p.325 [1]
- S. fourchei Lessert, 1939
R1 p.326 [1]
- S. fusca Walckenaer, 1837
R1 p.323 [14], p.329 [S.bajulal[1], [S.campinensis][1],
[S.discolor][1], [S.hebraical[2], p.330 [S.nannipes][1],
B p.150 [S.torquatal[1], P1 p.117 [3], P2 p.121 [2]
- S. gertschi Valerio, 1981
B p.674 [2], P1 p.117 [1]
- S. gilva (Thorell, 1887)
R1 p.327 [2]
- S. globula Nicolet, 1849
R1 p.329 [2]
- S. gooldi Purcell, 1904
R1 p.326 [1]
- S. grammocephala Simon, 1909
R1 p.328 [1]
- S. guttipes Simon, 1892
R1 p.329 [1]
- S. humilis L.Koch, 1873
R1 p.326 [2]
- S. iguassuensis Mello-Leitão, 1918
R1 p.330 [1]
- S. immaculata L.Koch, 1875
R1 p.324 [1]
- S. insperata Soares & Camargo, 1948
B p.149 [1]
- S. intricata Banks, 1909
R1 p.330 [2], P1 p.117 [1]
- S. itzana Chamberlin & Ivie, 1938
R1 p.330 [1]
- S. jousseau mei Simon, 1907
R1 p.326 [1]
- S. kaokoensis Lawrence, 1928
R1 p.326 [1]
- S. karrooica Purcell, 1904
R1 p.326 [1]
- S. kinsukus Patel, 1975
B p.149 [1]
- S. lanceolata Purcell, 1904
R1 p.326 [1]
- S. lawrencei Lessert, 1939
R1 p.326 [1]
- S. leipoldti Purcell, 1904
R1 p.326 [1]
- S. leopoldi Giltay, 1935
R1 p.328 [1], P1 p.117 [1]
- S. leprosula Strand, 1913
R1 p.326 [1]
- S. lesserti Millot, 1941
B p.149 [1]

- S. lewisi Alayón, 1985
P1 p.117 [1]
- S. lineatipes Taczanowski, 1874
R1 p.330 [4]
- S. longipes Lucas, 1844
R1 p.323 [12], p.329 [*S. penicillata*] [1],
P1 p.117 [3], P2 p.121 [1]
- S. longipes simplex Franganillo, 1926
P2 p.121 [1]
- S. lorenzoi Alayón, 1977
B p.150 [1]
- S. lugubris (Thorell, 1887)
R1 p.328 [3]
- S. luteola Simon, 1892
R1 p.330 [1]
- S. lycosella Purcell, 1904
R1 p.326 [1]
- S. lyriformis Purcell, 1904
R1 p.326 [1]
- S. maculata Holmberg, 1876
R1 p.330 [3]
- S. magna Bristowe, 1952
B p.149 [2], P1 p.117 [1]
- S. major Simon, 1885
R1 p.324 [2]
- S. maritima Lawrence, 1938
R1 p.326 [1]
- S. marshalli Pocock, 1902
R1 p.326 [1]
- S. mawphlongensis Tikader, 1966
B p.149 [2]
- S. meridiana Chamberlin & Ivie, 1938
R1 p.330 [1]
- S. mexicana Banks, 1898
R1 p.330 [1]
- S. montana Purcell, 1904
R1 p.326 [1]
- S. multilineata Thorell, 1899
R1 p.326 [2]
- S. nigristeris Simon, 1907
R1 p.326 [1]
- S. noeli Alayón, 1977
B p.150 [1]
- S. obelisci Denis, 1947
B p.148 [3]
- S. oswaldi Lenz, 1891
R1 p.326 [1]
- S. pallida Doleschall, 1859
R1 p.328 [2]
- S. perfecta Banks, 1898
R1 p.330 [2]

- S. perimensis (Simon, 1890)
R1 p.326 [2]
- S. pholcoides Simon, 1897
R1 p.327 [1]
- S. plumbea Mello-Leitão, 1929
R1 p.330 [1]
- S. pnocitens Chamberlin, 1924
R1 p.330 [1]
- S. propinqua Stoliczka, 1869
R1 p.328 [2]
- S. pulchella Berland, 1914
R1 p.327 [1]
- S. punctipes Simon, 1907
R1 p.327 [1]
- S. quarta Lawrence, 1927
R1 p.327 [1]
- S. quatuordecemmaculata Strand, 1907
R1 p.328 [1]
- S. quatuordecemmaculata clarior Strand, 1907
R1 p.328 [1]
- S. quinqua Lawrence, 1927
R1 p.327 [1]
- S. redempta Chamberlin, 1924
R1 p.330 [1]
- S. reticulata Jézéquel, 1964
B p.149 [1]
- S. robertoi Alayón, 1977
B p.150 [1]
- S. romiti Caporiacco, 1947
B p.150 [2]
- S. rubra Lawrence, 1937
R1 p.327 [1]
- S. rufipedata Roewer, 1942
R1 p.324 [1]
- S. ruizensis Strand, 1914
R1 p.330 [1]
- S. sansibarica Strand, 1907
R1 p.327 [2]
- S. scholaris Toledo Piza, 1944
B p.150 [1]
- S. schultzei Purcell, 1908
R1 p.327 [1]
- S. semipullata Simon, 1909
R1 p.328 [2], P2 p.121 [1]
- S. serripes Mello-Leitão, 1947
B p.150 [1]
- S. sexstriata Roewer, 1960
B p.148 [1]
- S. silvatica Purcell, 1904
R1 p.327 [1]
- S. sordida Dyal, 1935
R1 p.328 [1]

- S. stoliczkai Simon, 1897
R1 p.328 [1]
- S. strandi Spassky, 1941
B p.149 [3]
- S. striatipes (L.Koch, 1872)
R1 p.329 [3], p.328 [*S.nigrolineata*][3],
B p.148 [*S.depressa*][1], P1 p.117 [3]
- S. subadulta Strand, 1911
R1 p.329 [1]
- S. sublata Purcell, 1904
R1 p.327 [1]
- S. subthoracica Strand, 1906
R1 p.327 [1]
- S. suffusa Strand, 1906
R1 p.327 [1]
- S. symmetrica Lawrence, 1938
R1 p.327 [1]
- S. tardigrada Thorell, 1881
R1 p.328 [1]
- S. tenerifensis Wunderlich, 1987
P1 p.118 [1]
- S. tertia Lawrence, 1927
R1 p.327 [1]
- S. testudo Purcell, 1904
R1 p.327 [1]
- S. thoracica (Latreille, 1802)
R1 p.324 [18], P1 p.118 [4], P2 p.121 [9]
- S. triangulifera Purcell, 1904
R1 p.327 [1]
- S. trifoliata Lawrence, 1938
R1 p.327 [1]
- S. uligocetes Valerio, 1981
B p.674 [1], P1 p.118 [1]
- S. univittata Simon, 1882
R1 p.328 [3]
- S. univittata unilineata Thorell, 1887
R1 p.328 [1]
- S. velutina Heineken & Lowe, 1836
R1 p.324 [6], p.325 [*S.v.delicatula*][3],
P1 p.118 [1], P2 p.121 [2]
- S. venusta (Thorell, 1890)
R1 p.328 [3]
- S. vittata Keyserling, 1877
R1 p.330 [3]
- S. zapatana Gertsch & Mulaik, 1940
B p.149 [1]

(133 spp., 4 ssp.)

Family Drymusidae

[In R1 it is considered a subfamily of the Sicariidae.]

Genus *Drymusa* Simon, 1891

- D. armasi Alayón, 1981
B p.674 [1] (F.Scytodidae), P1 p.121 [2]
D. capensis Simon, 1893
R1 p.321 [1]
D. dinora Valerio, 1971
B p.146 [1] (F.Scytodidae)
D. nubila Simon, 1891
R1 p.322 [2]
D. producta Purcell, 1904
R1 p.322 [1]
D. silvicola Purcell, 1904
R1 p.322 [1]
D. simoni Bryant, 1948
B p.146 [2] (F.Scytodidae)
D. spectata Alayón, 1981
B p.674 [1] (F.Scytodidae), P1 p.121 [1]
(8 spp.)

Family Leptonetidae

Genus **Appaleptoneta** Platnick, 1986

- | | | | |
|------------------------|------------------------------|-----------|--|
| <u>A. barrowsi</u> | (Gertsch, 1974) | | |
| B | p.197 [1] [Leptoneta b.], P1 | p.138 [1] | |
| <u>A. coma</u> | (Barrows, 1940) | | |
| B | p.197 [2] [Leptoneta c.], P1 | p.138 [1] | |
| <u>A. credula</u> | (Gertsch, 1974) | | |
| B | p.197 [1] [Leptoneta c.], P1 | p.138 [1] | |
| <u>A. fiskei</u> | (Gertsch, 1974) | | |
| B | p.197 [1] [Leptoneta f.], P1 | p.138 [1] | |
| <u>A. gertschi</u> | (Barrows, 1940) | | |
| B | p.198 [2] [Leptoneta g.], P1 | p.138 [1] | |
| <u>A. jonesi</u> | (Gertsch, 1974) | | |
| B | p.198 [1] [Leptoneta j.], P1 | p.139 [1] | |
| <u>A. silvicultrix</u> | (Crosby & Bishop, 1925) | | |
| R1 | p.313 [1] [Leptoneta s.], P1 | p.139 [1] | |
- (7 spp.)

Genus **Archoleptoneta** Gertsch, 1974

- A. arganoi (Brignoli, 1974)
B p.194 [2]
A. garza Gertsch, 1974
B p.194 [1]

- A. obscura Gertsch, 1974
B p.194 [1]
A. schusteri Gertsch, 1974
B p.194 [1]

{4 spp. }

Genus *Barusia* Kratochvil, 1978

- B. hofferi (Kratochvíl, 1935)
R1 p.313 [2] [Paraleptoneta h.], P2 p.122 [1]
- B. insulana (Kratochvíl, & Miller, 1939)
R1 p.313 [2] [Paraleptoneta i.], P2 p.122 [1]
- B. korculana (Kratochvíl, & Miller, 1939)
R1 p.313 [2] [Paraleptoneta k.], P2 p.122 [1]
- B. laconica (Brignoli, 1974)
B p.194 [3]
- B. maheni (Kratochvíl, 1938)
R1 p.314 [2] [Paraleptoneta m.], P2 p.122 [1]
- (5 spp.)

Genus **Calileptoneta** Platnick, 1986

- C. californica (Banks, 1904)
R1 p.313 [3], B p.194 [1] [Leptoneta sylval,
P1 p.139 [1], P2 p.122 [1]
C. helferi (Gertsch, 1974)
B p.198 [1], P1 p.139 [1]
C. noyoana (Gertsch, 1974)
B p.198 [1], P1 p.139 [1]
C. oasa (Gertsch, 1974)
B p.198 [1], P1 p.139 [1]
C. wapiti (Gertsch, 1974)
B p.198 [1], P1 p.139 [1]
- (5 spp.)

Genus **Cataleptoneta** Denis, 1955

- C. aesculapii (Brignoli, 1968)
B p.195 [3]
C. edentula (Denis, 1955)
B p.195 [2]
C. sbordonii (Brignoli, 1968)
B p.195 [3]
C. sengletii (Brignoli, 1974)
B p.195 [2]

{4 spp.}

Genus *Falcileptoneta* Komatsu, 1970

- F. asuwana* (Nishikawa, 1981)
P1 p.139 [1]
- F. caeca* Yaginuma, 1972
B p.195 [1], P1 p.139 [1]
- F. inabensis* (Nishikawa, 1982)
P1 p.139 [1]
- F. iriei* (Komatsu, 1967)
B p.195 [3], P1 p.139 [2], P2 p.122 [1]
- F. japonica* (Simon, 1893)
R1 p.312 [2], P2 p.122 [1]
- F. kugoana* (Komatsu, 1961)
B p.195 [2]
- F. melanocomata* (Komatsu, 1961)
B p.195 [2]
- F. musculina* (Komatsu, 1961)
B p.195 [2]
- F. okinawaensis* Komatsu, 1972
B p.195 [1], P1 p.139 [1]
- F. speciosa* (Komatsu, 1957)
B p.195 [2]
- F. striata* (Oi, 1952)
B p.195 [4], P1 p.139 [1]
- F. striata fujisana* Yaginuma, 1972
B p.195 [1], P1 p.139 [1]
- F. tofacea* Yaginuma, 1972
B p.195 [1]
- F. tsushimensis* (Yaginuma, 1970)
B p.196 [2], P1 p.139 [1]
- F. uenoi* (Yaginuma, 1963)
B p.196 [2], P1 p.139 [1]
- F. ushihanana* (Komatsu, 1961)
B p.196 [2]
- F. zenjoensis* (Komatsu, 1965)
B p.196 [2]

{16 spp., 1 ssp.}

Genus *Leptoneta* Simon, 1872

- L. abeillei* Simon, 1882
R1 p.311 [3]
- L. alpica* Simon, 1882
R1 p.311 [3]
- L. anocellata* Chen, Zhang & Song, 1986
P1 p.140 [2], P2 p.122 [1]
- L. arquata* Song & Kim, 1991
P2 p.122 [1]
- L. baccettii* Brignoli, 1979
B p.196 [1]

- L. berlandi Machado & Ribera, 1986
P1 p.140 [1]
- L. brunnea Gertsch, 1974
B p.197 [1], P1 p.140 [1]
- L. cavalairensis Dresco, 1987
P1 p.140 [1]
- L. ciaisensis Dresco, 1987
P1 p.140 [1]
- L. comasi Ribera, 1978
B p.196 [1], P2 p.122 [1]
- L. condei Dresco, 1987
P1 p.140 [1]
- L. conimbricensis Machado & Ribera, 1986
P1 p.140 [1]
- L. convexa Simon, 1872
R1 p.311 [4], P2 p.122 [1]
- L. convexa aulotensis Dresco, 1990
P2 p.123 [1]
- L. coreana Paik & Namkung, 1969
B p.197 [1]
- L. corsica Fage, 1943
B p.196 [1], P1 p.140 [1]
- L. crypticola Simon, 1907
R1 p.311 [3], P1 p.140 [1]
- L. crypticola franciscoloi Caporiacco, 1950
B p.196 [4]
- L. crypticola simplex Fage, 1913
R1 p.311 [1]
- L. fagei Simon, 1914 1872 (P1)
R1 p.311 [2], P1 p.140 [2]
- L. fouresi Dresco, 1979
B p.196 [1]
- L. hangzhouensis Chen, Shen & Gao, 1984
P1 p.140 [2], P2 p.123 [1]
- L. hogyegulensis Paik & Namkung, 1969
B p.197 [1]
- L. hongdoensis Paik, 1980
P1 p.140 [1]
- L. huanglongensis Chen, Zhang & Song, 1982
P1 p.140 [3], P2 p.123 [1]
- L. hwanseoensis Namkung, 1987
P1 p.140 [1]
- L. infuscata Simon, 1872
R1 p.312 [3], [L.i.corberensis][2], [L.i.iberica][2],
[L.i.minos][3], P1 p.141 [2], [L.i.minos][1], P2 p.123 [1]
- L. infuscata ovetana Machado, 1939
R1 p.312 [1], B p.196 [1]
- L. insularis Roewer, 1953
B p.196 [3]
- L. jangsanensis Seo, 1989
P2 p.123 [1]

- L. jeanneli Simon, 1907
R1 p.312 [3]
- L. kernensis Simon, 1910
R1 p.312 [2]
- L. lantosquensis Dresco, 1987
P1 p.141 [1]
- L. leucophthalma Simon, 1907
R1 p.312 [2]
- L. lingqiensis Chen, Shen & Gao, 1984
P1 p.141 [2], P2 p.123 [1]
- L. maculosa Song & Xu, 1986
P1 p.141 [2], P2 p.123 [1]
- L. manca Fage, 1913
R1 p.313 [2] [*L. proserpina* m.], P1 p.141 [1]
- L. microdonta Xu & Song, 1983
P1 p.141 [2]
- L. microphthalma Simon, 1872
R1 p.312 [4]
- L. monodactyla Yin, Wang & Wang, 1984
P1 p.141 [2]
- L. namhensis Paik & Seo, 1982
P1 p.141 [1]
- L. olivacea Simon, 1882
R1 p.312 [3], P1 p.141 [2]
- L. paikmyeonggulensis Paik & Seo, 1984
P1 p.141 [1]
- L. paroculus Simon, 1907
R1 p.312 [2], P1 p.141 [1]
- L. patrizii Roewer, 1953
B p.196 [3]
- L. proserpina Simon, 1907
R1 p.313 [3], P1 p.141 [1]
- L. sandra Gertsch, 1974
B p.198 [1], P1 p.141 [1]
- L. secula Namkung, 1987
P1 p.141 [1]
- L. serbariuana Roewer, 1953
B p.196 [2]
- L. simboggulensis Paik, 1971
B p.197 [1]
- L. soryongensis Paik & Namkung, 1969
B p.197 [1]
- L. taeguensis Paik, 1985
P1 p.141 [1]
- L. taramellii Roewer, 1956
B p.196 [2]
- L. trabucensis Simon, 1907
R1 p.313 [3]
- L. trispinosa Yin, Wang & Wang, 1984
P1 p.141 [2]
- L. tunxiensis Song & Xu, 1986
P1 p.141 [2]

- L. unispinosa Yin, Wang & Wang, 1984
 P1 p.141 [2]
L. vittata Fage, 1913
 R1 p.313 [2], P1 p.141 [1]
L. waheulgulensis Namkung, 1991
 P2 p.123 [1]
L. yongdamgulensis Paik & Namkung, 1969
 B p.197 [1]
L. yongyeonensis Seo, 1989
 P2 p.123 [1]

{57 spp., 4 ssp.}

Genus **Leptonetela** Kratochvíl, 1978

- L. andreevi Deltchev, 1985
 P1 p.142 [1]
L. caucasica Dunin, 1990
 P2 p.123 [1]
L. deltshevi (Brignoli, 1979)
 B p.198 [2]
L. kanellisi (Deeleman-Reinhold, 1971)
 B p.198 [3]
L. strinatii (Brignoli, 1976)
 B p.198 [2]

{5 spp.}

Genus **Masirana** Kishida, 1942

- M. akahanei Komatsu, 1963
 B p.198 [2]
M. akiyoshiensis (Oi, 1958)
 B p.199 [3], P1 p.142 [1]
M. akiyoshiensis imperatoria Komatsu, 1974
 B p.199 [1]
M. akiyoshiensis kagekiyoi Komatsu, 1974
 B p.199 [1]
M. akiyoshiensis primocreata Komatsu, 1974
 B p.199 [1]
M. cineracea Komatsu, 1942
 B p.199 [1]
M. kosodeensis Komatsu, 1963
 B p.199 [2]
M. kuramotoi Komatsu, 1974
 B p.199 [1]
M. kyokoae Yaginuma, 1972
 B p.199 [1], P1 p.142 [1]
M. longimana Yaginuma, 1970
 B p.199 [1], P1 p.142 [1]
M. longipalpis Komatsu, 1972
 B p.199 [2], P1 p.142 [1]

- M. nippara Komatsu, 1957
 B p.199 [2], Pl p.142 [1]

{9 spp., 3 ssp.}

Genus **Neoleptoneta** Brignoli, 1972

- N. alabama (Gertsch, 1974)
 B p.199 [2]
N. anopica (Gertsch, 1974)
 B p.199 [2]
N. apachea (Gertsch, 1974)
 B p.199 [2]
N. archeri (Gertsch, 1974)
 B p.199 [2]
N. arkansa (Gertsch, 1974)
 B p.199 [2]
N. blanda (Gertsch, 1974)
 B p.199 [2]
N. bonita (Gertsch, 1974)
 B p.200 [2]
N. caliginosa Brignoli, 1977
 B p.200 [1]
N. capilla (Gertsch, 1971)
 B p.200 [3]
N. chisosea (Gertsch, 1974)
 B p.200 [2]
N. coeca (Chamberlin & Ivie, 1942)
 B p.200 [3]
N. concinna (Gertsch, 1974)
 B p.200 [2]
N. delicata (Gertsch, 1971)
 B p.200 [3]
N. devia (Gertsch, 1974)
 B p.200 [2]
N. furtiva (Gertsch, 1974)
 B p.200 [2]
N. georgia (Gertsch, 1974)
 B p.200 [2]
N. isolata (Gertsch, 1971)
 B p.200 [3]
N. iviei (Gertsch, 1974)
 B p.200 [2]
N. limpida (Gertsch, 1974)
 B p.200 [2]
N. microps (Gertsch, 1974)
 B p.200 [2]
N. modica (Gertsch, 1974)
 B p.200 [2]
N. myopica (Gertsch, 1974)
 B p.200 [2]

- N. novaegalleciae Brignoli, 1979
 B p.201 [1]
N. pecki (Gertsch, 1971)
 B p.201 [3]
N. rainesi (Gertsch, 1971)
 B p.201 [3]
N. reclusa (Gertsch, 1971)
 B p.201 [3]
N. serena (Gertsch, 1974)
 B p.201 [2]
N. uvaldea (Gertsch, 1974)
 B p.201 [2]
N. valverdae (Gertsch, 1974)
 B p.201 [2]

{29 spp.}

Genus Paraleptoneta Fage, 1913

- P. bellesi Ribera & Lopez, 1982
 P1 p.142 [1]
P. spinimana (Simon, 1884)
 R1 p.314 [4], B p.194 [P.fagei][1], [P.parenzani][1],
 [P.pasquini][1], [P.patrizii][1], [Segrea sardiniensis][1],
 [Segrea strinati][1], P2 p.123 [1]

{2 spp.}

Genus Protoleptoneta Deltchev, 1972

- P. beroni Deltchev, 1977
 B p.201 [1]
P. bulgarica Deltchev, 1972
 B p.201 [1]
P. italica (Simon, 1907)
 R1 p.313 [3] [Paraleptoneta i.], P2 p.123 [1]
 {3 spp.}

Genus Sarutana Komatsu, 1957

- S. abensis Kobayashi, 1973
 B p.201 [1]
S. bandoi (Nishikawa, 1986)
 P1 p.142 [1]
S. glabra Komatsu, 1957
 B p.202 [1]
S. kawasawai Komatsu, 1970
 B p.202 [1]
S. silvicola Kobayashi, 1973
 B p.202 [1], P1 p.142 [1]
S. yamauchii (Nishikawa, 1982)
 P1 p.142 [1]

{6 spp.}

Genus **Sulcia** Kratochvíl, 1938

- S. armata Kratochvíl, 1978
 B p.202 [2]
- S. cretica Fage, 1945
 B p.202 [4], P2 p.124 [1]
- S. cretica lindbergi Dresco, 1962
 B p.202 [4]
- S. cretica violacea Brignoli, 1974
 B p.202 [2]
- S. inferna Kratochvíl, 1938
 R1 p.314 [1]
- S. mirabilis (Kratochvíl, 1934)
 R1 p.314 [3]
- S. montenegrina (Kratochvíl, 1938)
 R1 p.314 [2] [Paraleptoneta m.], P2 p.124 [1]
- S. nocturna Kratochvíl, 1938
 R1 p.314 [1]
- S. orientalis (Kulczyński, 1914)
 R1 p.314 [4]
- S. orientalis occulta Kratochvíl, 1938
 R1 p.314 [1]

{7 spp., 3 ssp.}

Geus **Teloleptoneta** Ribera, 1988

- T. synthetica (Machado, 1951)
 B p.201 [1] [Paraleptoneta s.], P1 p.142 [1] [P.s.],
 P2 p.124 [1]

{1 sp.}

Index to Genera and Families

Note: names of families are printed in boldface, and every genus is followed by the number of species and subspecies.

	Number of species and subspecies	Page
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Calileptoneta	5 spp.	137
Cataleptoneta	4 spp.	137
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Drymusidae		136
Ectatosticta	1 sp.	118
Falcileptoneta	16 spp., 1 ssp.	138
Filistata	19 spp., 1 ssp.	121
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Filistatinella	1 sp.	121
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Hickmania	1 sp.	119
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Hypochilus	8 spp.	118
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Spelungula	1 sp.	120
Sulcia	7 spp., 3 ssp.	144
Tarlina	6 spp.	120
Teloleptoneta	1 sp.	144
Thaida	2 spp.	119
Tricalamus	7 spp.	124
Zaitunia	8 spp.	124

Total number of taxa : 8 Families, 43 Genera,
536 Species, 19 Subspecies



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VOLUME 5
PART 4

CAIRO - EGYPT
1997

SERKET

Volume 5

Part 4

December, 1997

Cairo , Egypt

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Volume 3 (1992-1993):
US \$ 35.00 (p.r.), US \$ 45.00 (i.r.)

Correspondence concerning subscription, back issues, publication,
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The Genera of Spiders

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Cairo 11341, Egypt

This work contains all the valid generic names of spiders (Arachnida : Araneida). It depends mainly on Platnick's "*Advances in spider taxonomy 1988-1991*" (1993), in addition to the following arachnological publications : *Arthropoda Selecta* (1992-95), *Bulletin of the British Arachnological Society* (1992-97), *The Journal of Arachnology* (1992-97) & *Korean Arachnology* (1992-95).

There is a list of spider families in addition to the number of genera within each family. This list is followed by a bigger one containing the generic names arranged alphabetically under every family name and followed by the author's name and year of publication. The order of families is also alphabetic. At the end, there is an index of generic names preceded by a list of abbreviations used after the names in the index to facilitate the reference to which family such a genus belongs.

"*The Genera of Spiders*" is a part of "Index Araneorum" which is continuously appearing in *Serket*.

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with number of genera of each family**

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Amphinectidae	17	Hersiliidae	5	Pholcidae	39
Anapidae	32	Heteropodidae	82	Pisauridae	54
Antrodiaetidae	3	Hexathelidae	11	Plectreuridae	2
Anyphaenidae	36	Holarchaeidae	1	Prodidomidae	27
Araneidae	156	Homalonychidae	1	Psechridae	4
Archaeidae	3	Huttoniidae	1	Salticidae	481
Argyronetidae	1	Hypochilidae	2	Scytodidae	1
Atypidae	3	Idiopidae	19	Segestriidae	4
Austrochilidae	3	Lamponidae	1	Selenopidae	4
Barychelidae	32	Leptonetidae	15	Senoculidae	1
Bradystichidae	1	Linyphiidae	489	Sicariidae	2
Caponiidae	8	Liocranidae	44	Stenochilidae	2
Cithaeronidae	2	Liphistiidae	2	Stiphidiidae	8
Clubionidae	26	Lycosidae	96	Symphytognathidae	6
Corinnidae	51	Malkaridae	4	Synotaxidae	12
Cryptothelidae	1	Mecicobothriidae	4	Telemidae	6
Ctenidae	37	Mecysmaucheniidae	7	Tengellidae	5
Ctenizidae	10	Micropholcommatidae	8	Tetrablemmidae	29
Cyatholipidae	12	Microstigmatidae	4	Tetragnathidae	51
Cybaeidae	8	Migidae	9	Theraphosidae	88
Cycloctenidae	5	Mimetidae	12	Theridiidae	62
Cyrtaucheniidae	18	Miturgidae	23	Theridiosomatidae	12
Deinopidae	4	Mysmenidae	22	Thomisidae	160
Desidae	37	Nemesiidae	37	Titanoecidae	5
Dictynidae	47	Neolanidae	1	Trechaleidae	11
Diguetidae	3	Nesticidae	7	Trochanteriidae	6
Dipluridae	20	Nicodamidae	2	Uloboridae	20
Drymusidae	1	Ochyroceratidae	9	Zodariidae	50
Dysderidae	20	Oecobiidae	6	Zoridae	12
Eresidae	10	Oonopidae	54	Zoropsidae	2
Filistatidae	12	Orsolobidae	27		
Gallieniellidae	3	Oxyopidae	9		

Total number of taxa : **106** Families, **3107** Genera

Family **Actinopodidae** 3 genera

Actinopus Perty, 1833
Missulena Walckenaer, 1805
*Plesiolen*a Goloboff & Platnick, 1987

Family **Agelenidae** 43 genera

Agelena Walckenaer, 1805
Agelenella Lehtinen, 1967
Agelenopsis Giebel, 1869
Ahua Forster & Wilton, 1973
Barronopsis Chamberlin & Ivie, 1941
Benoitia Lehtinen, 1967
Calilena Chamberlin & Ivie, 1941
Coreidon Mello-Leitão, 1917
Hadites Keyserling, 1862
Hicanodon Tullgren, 1901
Histopona Thorell, 1869
Hololena Exline, 1938
Huangyuania Song & Li, 1990
Huka Forster & Wilton, 1973
Kidugua Lehtinen, 1967
Lycosoides Lucas, 1846
Mahura Forster & Wilton, 1973
Maimuna Lehtinen, 1967
Malthonica Simon, 1898
Melpomene O.P.-Cambridge, 1898
Mevianops Mello-Leitão, 1941
Mistaria Lehtinen, 1967
Neoramia Forster & Wilton, 1973
Neorepukia Forster & Wilton, 1973
Neotegenaria Roth, 1967
Novalena Chamberlin & Ivie, 1942
Nyssus Walckenaer, 1805
Olorunia Lehtinen, 1967
Oramia Forster, 1964
Oramiella Forster & Wilton, 1973
Orepukia Forster & Wilton, 1973
Paramyro Forster & Wilton, 1973
Philoicides Mello-Leitão, 1944
Porotaka Forster & Wilton, 1973
Pseudophthalmus Joseph, 1882
Pseudotegenaria Caporiacco, 1934
Rualena Chamberlin & Ivie, 1942
Tararua Forster & Wilton, 1973
Tegenaria Latreille, 1804
Textrix Sundevall, 1833
Tikaderia Lehtinen, 1967
Tortolena Chamberlin & Ivie, 1941
Tuapoka Forster & Wilton, 1973

Family **Amaurobiidae** 62 genera

Altellopsis Simon, 1906
Amaurobius C.L.Koch, 1837
Ambohima Griswold, 1990
Anisacate Mello-Leitão, 1941
Arctobius Lehtinen, 1967
Auhunga Forster & Wilton, 1973
Austmusia Gray, 1983
Auximella Strand, 1908
Bakala Davies, 1990
Barrisca Chamberlin & Ivie, 1936
Calacadia Exline, 1960
Callevopsis Tullgren, 1902

Callobius Chamberlin, 1947
Chresiona Simon, 1903
Cinifella Mello-Leitão, 1921
Coelotes Blackwall, 1841
Coras Simon, 1898
Cybaeopsis Strand, 1907
Dardurus Davies, 1976
Emmenomma Simon, 1884
Kulalania Griswold, 1990
Lamaika Griswold, 1990
Livius Roth, 1967
Macrobunus Tullgren, 1901
Malaika Lehtinen, 1967
Maloides Forster & Wilton, 1989
Manjala Davies, 1990
Matundua Lehtinen, 1967
Metaltella Mello-Leitão, 1931
Muritaia Forster & Wilton, 1973
Namaquarachne Griswold, 1990
Neoporteria Mello-Leitão, 1943
*Neuquen*ia Mello-Leitão, 1940
Obatala Lehtinen, 1967
Otira Forster & Wilton, 1973
Pakeha Forster & Wilton, 1973
Paracoelotes Brignoli, 1982
Paravoca Forster & Wilton, 1973
Phyxelida Simon, 1894
Pimus Chamberlin, 1947
Pongolania Griswold, 1990
Pseudauximus Simon, 1902
Retiro Mello-Leitão, 1915
Rhoicinaria Exline, 1950
Rhoicinus Simon, 1898
Rubrius Simon, 1887
Storenosoma Hogg, 1900
Taira Lehtinen, 1967
Tamgrinia Lehtinen, 1967
Themacrys Simon, 1906
Tugana Chamberlin, 1948
Tymbira Mello-Leitão, 1944
Urepus Roth, 1967
Vidole Lehtinen, 1967
Virgilus Roth, 1967
Vytfutia Deeleman-Reinhold, 1986
Wadotes Chamberlin, 1925
Waitetola Forster & Wilton, 1973
Xevioso Lehtinen, 1967
Yacolla Lehtinen, 1967
Yupanquia Lehtinen, 1967
Zanomys Chamberlin, 1948

Family **Ammoxenidae** 2 genera

Ammoxenus Simon, 1893
Rastellus Platnick & Griffin, 1990

Family **Amphinectidae** 17 genera

Akatorea Forster & Wilton, 1973
Amphinecta Simon, 1898
Aorangia Forster & Wilton, 1973
Dunstanoides Forster & Wilton, 1989
Holomamoea Forster & Wilton, 1973
Huara Forster, 1964
Makora Forster & Wilton, 1973
Mamoea Forster & Wilton, 1973

Maniho Marples, 1959
 Marplesia Lehtinen, 1967
 Neororea Forster & Wilton, 1973
 Oparara Forster & Wilton, 1973
 Paramamoea Forster & Wilton, 1973
 Rangitata Forster & Wilton, 1973
 Reinga Forster & Wilton, 1973
 Rorea Forster & Wilton, 1973
 Waterea Forster & Wilton, 1973

Family **Anapidae** 32 genera

Anapis Simon, 1895
 Anapisona Gertsch, 1941
 Caledanapis Platnick & Forster, 1989
 Chasmocephalon O.P.-Cambridge, 1889
 Comaroma Bertkau, 1889
 Conoculus Komatsu, 1940
 Crassanapis Platnick & Forster, 1989
 Crozetulus Hickman, 1939
 Elanapis Platnick & Forster, 1989
 Forsteriola Brignoli, 1981
 Gertschanapis Platnick & Forster, 1990
 Hickmanapis Platnick & Forster, 1989
 Mandanapis Platnick & Forster, 1989
 Maxanapis Platnick & Forster, 1989
 Metanapis Brignoli, 1981
 Minanapis Platnick & Forster, 1989
 Montanapis Platnick & Forster, 1989
 Nortanapis Platnick & Forster, 1989
 Novanapis Platnick & Forster, 1989
 Octanapis Platnick & Forster, 1989
 Paranapis Platnick & Forster, 1989
 Pecanapis Platnick & Forster, 1989
 Pseudanapis Simon, 1905
 Queenslanapis Platnick & Forster, 1989
 Risdonius Hickman, 1939
 Sheranapis Platnick & Forster, 1989
 Sofanapis Platnick & Forster, 1989
 Spinanapis Platnick & Forster, 1989
 Tasmanapis Platnick & Forster, 1989
 Victanapis Platnick & Forster, 1989
 Zangherella Caporiacco, 1949
 Zealanapis Platnick & Forster, 1989

Family **Antrodiaetidae** 3 genera

Aliatypus Smith, 1908
 Antrodiaetus Ausserer, 1871
 Atypoides O.P.-Cambridge, 1883

Family **Anypaenidae** 36 genera

Amaurobioides O.P.-Cambridge, 1883
 Anypaena Sundevall, 1833
 Anypaenoides Berland, 1913
 Aporatea Simon, 1897
 Arachosia O.P.-Cambridge, 1882
 Australaena Berland, 1942
 Axyracrus Simon, 1884
 Aysha Keyserling, 1891
 Coptoprepes Simon, 1884
 Gayenna Nicolet, 1849
 Gayennella Berland, 1913
 Haptisus Simon, 1896
 Hibana Brescovit, 1991

Isigonia Simon, 1896
 Josa Keyserling, 1891
 Liparotoma Simon, 1884
 Macrophytes O.P.-Cambridge, 1893
 Mesilla Simon, 1903
 Mezenina Strand, 1932
 Monapia Simon, 1897
 Olbophthalmus Simon, 1904
 Osoriella Mello-Leitão, 1922
 Oxsoma Nicolet, 1849
 Patrera Simon, 1903
 Pelayo O.P.-Cambridge, 1896
 Quechuella Chamberlin, 1916
 Sillus F.O.P.-Cambridge, 1900
 Tafana Simon, 1904
 Tasata Simon, 1903
 Temnida Simon, 1896
 Terupis Simon, 1904
 Tetromma Keyserling, 1877
 Teudis O.P.-Cambridge, 1896
 Tomopisthes Simon, 1884
 Wulfila O.P.-Cambridge, 1895
 Wulfilopsis Soares & Camargo, 1955

Family **Araneidae** 156 genera

Acacesia Simon, 1895
 Acantharanea Strand, 1929
 Acanthepeira Marx, 1883
 Acroaspis Karsch, 1878
 Acrosomoides Simon, 1887
 Actinacantha Simon, 1864
 Actinosoma Holmberg, 1883
 Aculepeira Chamberlin & Ivie, 1942
 Acusilas Simon, 1895
 Aerea Urquhart, 1891
 Aethruscus Pocock, 1902
 Aethrodiscus Strand, 1913
 Aetrocantha Karsch, 1879
 Afracantha Dahl, 1914
 Agalenatea Archer, 1951
 Agathostichus Simon, 1895
 Alcimosphenus Simon, 1895
 Alpaida O.P.-Cambridge, 1889
 Amazonepeira Levi, 1989
 Anepsion Strand, 1929
 Arachnura Vinson, 1863
 Araneus Clerck, 1757
 Araniella Chamberlin & Ivie, 1942
 Aranoethra Butler, 1873
 Argiope Savigny, 1825
 Artonis Simon, 1895
 Aspidolasius Simon, 1887
 Augusta O.P.-Cambridge, 1877
 Austracantha Dahl, 1914
 Bertrana Keyserling, 1884
 Bunocrania Thorell, 1878
 Caerostris Thorell, 1868
 Cardimia Millo-Leitão, 1940
 Carepalxis L.Koch, 1872
 Celaenia Thorell, 1868
 Cercidia Thorell, 1869
 Chaetacis Simon, 1895
 Chorizopella Lawrence, 1947
 Chorizopes O.P.-Cambridge, 1870
 Cladomelea Simon, 1895

- Cnodalia* Thorell, 1890
Coelossia Simon, 1895
Colaranae Court & Forster, 1988
Collina Urquhart, 1891
Colphepeira Archer, 1941
Cryptaranae Court & Forster, 1988
Cyclosa Menge, 1866
Cyphalonotus Simon, 1895
Cyrtarachne Thorell, 1868
Cyrtophora Simon, 1864
Deione Thorell, 1898
Dolophones Walckenaer, 1837
Dubiepeira Levi, 1991
Edricus O.P.-Cambridge, 1890
Enacrosoma Mello-Leitão, 1932
Encyosaccus Simon, 1895
Epeirella Mello-Leitão, 1941
Epeiroides Keyserling, 1885
Eriophora Simon, 1864
Eriovixia Archer, 1951
Euglyptila Simon, 1909
Eustacesia Caporiacco, 1954
Eustala Simon, 1895
Exechocentrus Simon, 1889
Faradja Grasshoff, 1970
Friula O.P.-Cambridge, 1896
Gasteracantha Sundevall, 1833
Gastroxya Benoit, 1962
Gea C.L.Koch, 1843
Gibbaranea Archer, 1951
Glyptogona Simon, 1884
Gnolus Simon, 1879
Heterognatha Nicolet, 1849
Heurodes Keyserling, 1886
Homalopolys Simon, 1895
Hypognatha Guérin, 1840
Hypsacantha Dahl, 1914
Hypsosinga Ausserer, 1871
Ideocaira Simon, 1903
Isoxya Simon, 1885
Kaira O.P.-Cambridge, 1889
Kilima Grasshoff, 1970
Larinia Simon, 1874
Lariniaria Grasshoff, 1970
Larinioides Caporiacco, 1934
Lipocrea Thorell, 1878
Macracantha Simon, 1864
Madacantha Emerit, 1970
Mahembea Grasshoff, 1970
Mangora O.P.-Cambridge, 1889
Mastophora Holmberg, 1876
Mecynogea Simon, 1903
Megaraneus Lawrence, 1968
Melychiopharis Simon, 1895
Metazygia F.O.P.-Cambridge, 1903
Metepeira F.O.P.-Cambridge, 1903
Micrathena Sundevall, 1833
Micrepeira Schenkel, 1953
Micropoltys Kulczyński, 1911
Milonia Thorell, 1890
Molinaranea Mello-Leitão, 1940
Nanduti Mello-Leitão, 1945
Nemoscolus Simon, 1895
Nemosinga Caporiacco, 1947
Nemospiza Simon, 1903
Neoarchemorus Mascord, 1968
Neogea Levi, 1983
Neoscona Simon, 1864
Novakiella Court & Forster, 1993
Novaranae Court & Forster, 1988
Nuctenea Simon, 1864
Ordgarius Keyserling, 1886
Paralarinia Grasshoff, 1970
Paraplectana Brito Capello, 1867
Paraplectanoides Keyserling, 1886
Pararaneus Caporiacco, 1940
Parawixia F.O.P.-Cambridge, 1904
Pasilobus Simon, 1895
Pherenice Thorell, 1899
Pitharatus Simon, 1895
Poecilarcys Simon, 1895
Poecilopachys Simon, 1895
Polys C.L.Koch, 1843
Pozonia Schenkel, 1953
Prasonica Simon, 1895
Prasonicella Grasshoff, 1971
Pronoides Schenkel, 1937
Pronous Keyserling, 1880
Pseudartonis Simon, 1903
Pseudopsyllo Strand, 1916
Psyllo Thorell, 1899
Pycnacantha Blackwall, 1865
Scoloderus Simon, 1887
Sedasta Simon, 1894
Singa C.L.Koch, 1836
Siwa Grasshoff, 1970
Spilasma Simon, 1895
Spintharidius Simon, 1893
Taczanowskia Keyserling, 1880
Talhybia Thorell, 1898
Testudinaria Taczanowski, 1879
Thelacantha van Hasselt, 1882
Thoracites Thorell, 1898
Thorellina Berg, 1899
Togacantha Dahl, 1914
Umbonata Grasshoff, 1971
Ursa Simon, 1895
Verrucosa McCook, 1888
Wagneriana F.O.P.-Cambridge, 1904
Witica O.P.-Cambridge, 1895
Wixia O.P.-Cambridge, 1882
Xylethrus Simon, 1895
Yaginumia Archer, 1960
Zealaranea Court & Forster, 1988
Zigana Chamberlin & Ivie, 1936
Zilla C.L.Koch, 1834
- Family **Archaeidae** 3 genera

Afrarchaea Forster & Platnick, 1984
Archaea C.L.Koch & Berendt, 1854
Austrarchaea Forster & Platnick, 1984
- Family **Argyronetidae** 1 genus

Argyroneta Latreille, 1804
- Family **Atypidae** 3 genera

Atypus Latreille, 1804
Calommata Lucas, 1837
Sphodros Walckenaer, 1835

Family **Austrochilidae** 3 genera

Austrochilus Gertsch & Zapfe, 1955
Hickmania Gertsch, 1958
Thaïda Karsch, 1880

Family **Barychelidae** 32 genera

Ammonius Thorell, 1899
Atrophothele Pocock, 1903
Barychelus Simon, 1888
Bestrigus Franganillo, 1930
Cosmopelma Simon, 1889
Cyphonisia Simon, 1889
Cyrtogrammomma Pocock, 1895
Dimazion Franganillo, 1926
Diplothele O.P.-Cambridge, 1890
Encyocrypta Simon, 1889
Eubrachycercus Pocock, 1897
Idioctis L.Koch, 1874
Idiommata Ausserer, 1871
Idiophthalma O.P.-Cambridge, 1887
Monodontium Kulczyński, 1908
Paracenobiopelma Feio, 1952
Pisenor Simon, 1888
Plagiobothrus Karsch, 1891
Psalistops Simon, 1889
Rhianodes Raven, 1985
Sason Simon, 1887
Sasonichus Pocock, 1900
Sipalolasma Simon, 1892
Strophaeus Ausserer, 1875
Synothele Simon, 1908
Thalerommata Ausserer, 1875
Tigidia Simon, 1892
Trichopelma Simon, 1888
Trittame L.Koch, 1873
Troglothele Fage, 1929
Zophorame Raven, 1990
Zophoryctes Simon, 1902

Family **Bradystichidae** 1 genus

Bradystichus Simon, 1880

Family **Caponiidae** 8 genera

Bruchnops Mello-Leitão, 1939
Caponia Simon, 1887
Caponina Simon, 1891
Diploglena Purcell, 1904
Nops MacLeay, 1838
Nopsides Chamberlin, 1924
Orthonops Chamberlin, 1924
Tarsonops Chamberlin, 1924

Family **Cithaeronidae** 2 genera

Cithaeron O.P.-Cambridge, 1872
Inthaeron Platnick, 1991

Family **Clubionidae** 26 genera

Adcatomus Karsch, 1880

Alloclubionoides Paik, 1992

Arushina Caporiacco, 1947

Aysenia Tullgren, 1902

Bucliona Benoit, 1977

Carteroniella Strand, 1907

Carteronius Simon, 1896

Cheiracanthium C.L.Koch, 1839

Chiracanthops Mello-Leitão, 1942

Clubiona Latreille, 1804

Clubionina Berland, 1947

Dorymetaecus Rainbow, 1920

Elaver O.P.-Cambridge, 1898

Helebiona Benoit, 1977

Lascona Georgescu, 1989

Macerio Simon, 1897

Matidia Thorell, 1878

Meedo Main, 1987

Neoanagraphis Gertsch & Mulaik, 1936

Nicoletina Mello-Leitão, 1951

Olbus Simon, 1880

Simalio Simon, 1897

Systaria Simon, 1897

Tecution Benoit, 1977

Tixcocoba Gertsch, 1977

Tolophus Thorell, 1891

Family **Corinnidae** 51 genera

Acanthoceto Mello-Leitão, 1944

Aetius O.P.-Cambridge, 1896

Apochinomma Pavesi, 1881

Aristerus Simon, 1909

Asadipus Simon, 1898

Austrachelas Lawrence, 1938

Austrophaea Lawrence, 1952

Brachyphaea Simon, 1895

Castanilla Caporiacco, 1936

Castianeira Keyserling, 1879

Centrothele L.Koch, 1873

Ceto Simon, 1874

Cetonana Mello-Leitão, 1941

Coenoptychus Simon, 1885

Copa Simon, 1885

Corinna C.L.Koch, 1841

Corinnomma Karsch, 1880

Cycalis Thorell, 1877

Diestus Simon, 1898

Graptartia Simon, 1896

Humua Ono, 1987

Lausus Simon, 1898

Lessertina Lawrence, 1942

Mandaneta Strand, 1932

Mazax O.P.-Cambridge, 1898

Medmassa Simon, 1887

Megalostrata Karsch, 1880

Merenius Simon, 1909

Methesis Simon, 1896

Myrmecium Latreille, 1824

Myrmecotypus O.P.-Cambridge, 1894

Oedignatha Thorell, 1881

Paccius Simon, 1898

Poecilopta Simon, 1896

Procopius Thorell, 1899

Pronophaea Simon, 1897

Psellocoptus Simon, 1896

Pseudoceto Mello-Leitão, 1929

Pseudocorinna Simon, 1910
 Sanogasta Mello-Leitão, 1941
 Schiapellia Mello-Leitão, 1938
 Scortecchia Caporiacco, 1936
 Sphecotypus O.P.-Cambridge, 1895
 Sphingius Thorell, 1890
 Stethorrhagus Simon, 1896
 Stratius Simon, 1898
 Supunna Simon, 1896
 Trachelas L.Koch, 1866
 Trachelopachys Simon, 1897
 Utivarachna Kishida, 1940
 Xeropigo O.P.-Cambridge, 1882

Family **Cryptothelidae** 1 genus

Cryptothele L.Koch, 1872

Family **Ctenidae** 37 genera

Acantheis Thorell, 1891
 Acanthoctenus Keyserling, 1876
 Africactenus Hyatt, 1954
 Anahita Karsch, 1879
 Apolania Simon, 1897
 Asthenoctenus Simon, 1897
 Caloctenus Keyserling, 1876
 Celaetycheus Simon, 1897
 Centroctenus Mello-Leitão, 1929
 Ctenopsis Schmidt, 1956
 Ctenus Walckenaer, 1805
 Cupiennius Simon, 1891
 Enoploctenus Simon, 1897
 Gephyroctenus Mello-Leitão, 1936
 Horioctenoides Main, 1954
 Incasoctenus Mello-Leitão, 1942
 Isoctenus Bertkau, 1880
 Itatiaya Mello-Leitão, 1915
 Janusia Gray, 1973
 Leptoctenus L.Koch, 1878
 Montescueia Carcavallo & Martinez, 1961
 Nemoctenus Forster & Wilton, 1973
 Nothroctenus Badcock, 1932
 Oligoctenus Simon, 1887
 Paravulsor Mello-Leitão, 1922
 Phoneutria Perty, 1833
 Phymatoctenus Simon, 1896
 Pseudoctenus Caporiacco, 1949
 Thoriosa Simon, 1910
 Trogloctenus Lessert, 1935
 Trujillina Bryant, 1948
 Tuticanus Simon, 1896
 Viracucha Lehtinen, 1967
 Viridasius Simon, 1889
 Vulsor Simon, 1888
 Wiedenmeyeria. Schenkel, 1953
 Zealotenus Forster & Wilton, 1973

Family **Ctenizidae** 10 genera

Aepycephalus Ausserer, 1871
 Bothriocyrtum Simon, 1891
 Conothele Thorell, 1878
 Cteniza Latreille, 1829
 Cyclocosmia Ausserer, 1871
 Cyrtocarenum Ausserer, 1871

Hebestatis Simon, 1903
 Latouchia Pocock, 1901
 Stasimopus Simon, 1892
 Ummidia Thorell, 1875

Family **Cyatholipidae** 12 genera

Cyatholipus Simon, 1894
 Hanea Forster, 1988
 Ilisoa Griswold, 1987
 Isicabu Griswold, 1987
 Matilda Forster, 1988
 Scharffia Griswold, 1997
 Teemenaarus Davies, 1978
 Tekella Urquhart, 1893
 Tekellatus Wunderlich, 1978
 Tekelloides Forster, 1988
 Toddiana Forster, 1988
 Ulwembua Griswold, 1987

Family **Cybaeidae** 8 genera

Cybaeina Chamberlin & Ivie, 1942
 Cybaeota Chamberlin & Ivie, 1933
 Cybaeozuga Chamberlin & Ivie, 1937
 Cybaeus L.Koch, 1868
 Dolichocybaeus Kishida, 1968
 Heterocybaeus Komatsu, 1968
 Symposia Simon, 1898
 Vagellia Simon, 1899

Family **Cycloctenidae** 5 genera

Anaua Forster, 1970
 Cycloctenus L.Koch, 1878
 Galliena Simon, 1898
 Plectophanes Bryant, 1935
 Toxopsiella Forster, 1964

Family **Cyrtaucheniidae** 18 genera

Acontius Karsch, 1879
 Actinoxia Simon, 1890
 Ancyлотрыпа Simon, 1889
 Aptostichus Simon, 1891
 Astrosoga Chamberlin, 1940
 Bolostromoides Schiapelli & Gerschman, 1945
 Bolostromus Ausserer, 1875
 Cyrtauchenius Thorell, 1869
 Enrico O.P.-Cambridge, 1895
 Entychides Simon, 1888
 Eucteniza Ausserer, 1875
 Fufius Simon, 1888
 Homostola Simon, 1892
 Kiama Main & Mascord, 1969
 Myrmekiaphila Atkinson, 1886
 Nemesoides Chamberlin, 1920
 Promyrmekiaphila Schenkel, 1950
 Rhytidicolus Simon, 1889

Family **Deinopidae** 4 genera

Avella O.P.-Cambridge, 1877
 Avelloopsis Purcell, 1904
 Deinopsis MacLeay, 1839

Menneus Simon, 1876

Family **Desidae** 37 genera

Badumna Thorell, 1890
 Cedicus Simon, 1875
 Cicirra Simon, 1886
 Desis Walckenaer, 1837
 Epimecinus Simon, 1908
 Forsterina Lehtinen, 1967
 Gasparia Marples, 1956
 Gohia Dalmás, 1917
 Goyenia Forster, 1970
 Hapona Forster, 1970
 Helsonia Forster, 1970
 Hulua Forster & Wilton, 1973
 Laestrygones Urquhart, 1893
 Lamina Forster, 1970
 Lathyarca Simon, 1908
 Manawa Forster, 1970
 Mangareia Forster, 1970
 Matachia Dalmás, 1917
 Myro O.P.-Cambridge, 1876
 Naevius Roth, 1967
 Namandia Lehtinen, 1967
 Neomyro Forster & Wilton, 1973
 Notomatachia Forster, 1970
 Nuisiana Forster & Wilton, 1973
 Ommatauxesis Simon, 1903
 Otagoa Forster, 1970
 Panoa Forster, 1970
 Paramatachia Dalmás, 1918
 Paratheuma Bryant, 1940
 Pitonga Davies, 1984
 Porteria Simon, 1904
 Rapua Forster, 1970
 Syrorisa Simon, 1908
 Taurongia Hogg, 1901
 Toxops Hickman, 1940
 Toxopsoides Forster & Wilton, 1973
 Tuakana Forster, 1970

Family **Dictynidae** 47 genera

Aebutina Simon, 1892
 Ajmonia Caporiacco, 1934
 Altella Simon, 1884
 Anaxibia Thorell, 1898
 Arangina Lehtinen, 1967
 Archaeodictyna Caporiacco, 1928
 Arctella Holm, 1945
 Argenna Thorell, 1869
 Argennina Gertsch & Mulaik, 1936
 Atelolathys Simon, 1892
 Banaidja Lehtinen, 1967
 Blabomma Chamberlin & Ivie, 1937
 Brommella Tullgren, 1948
 Callevophthalmus Simon, 1906
 Chaerea Simon, 1884
 Chozomma Simon, 1872
 Cicurina Menge, 1871
 Devade Simon, 1884
 Dictyna Sundevall, 1833
 Dictynomorpha Spassky, 1939
 Emblyna Chamberlin, 1948
 Hackmania Lehtinen, 1967

Helenactyna Benoit, 1977
 Hoplolathys Caporiacco, 1947
 Iviella Lehtinen, 1967
 Lathys Simon, 1884
 Mallos O.P.-Cambridge, 1902
 Marilynia Lehtinen, 1967
 Mashimo Lehtinen, 1967
 Mastigusa Menge, 1854
 Mexitlia Lehtinen, 1967
 Mizaga Simon, 1898
 Nigma Lehtinen, 1967
 Paradictyna Forster, 1970
 Phantyna Chamberlin, 1948
 Qiyunia Song & Xu, 1989
 Rhion O.P.-Cambridge, 1870
 Saltonia Chamberlin & Ivie, 1942
 Shango Lehtinen, 1967
 Sudesna Lehtinen, 1967
 Tahuantina Lehtinen, 1967
 Tandil Mello-Leitão, 1940
 Thallumetus Simon, 1892
 Tivyna Chamberlin, 1948
 Tricholathys Chamberlin & Ivie, 1935
 Viridictyna Forster, 1970
 Yorima Chamberlin & Ivie, 1942

Family **Diguetidae** 3 genera

Diguetia Simon, 1895
 Pertica Simon, 1903
 Segestrioides Keyserling, 1883

Family **Dipluridae** 20 genera

Allothele Tucker, 1920
 Australothele Raven, 1984
 Caledothele Raven, 1991
 Carrai Raven, 1984
 Cethegus Thorell, 1881
 Chilehexops Coyle, 1986
 Diplura C.L.Koch, 1850
 Euagrus Ausserer, 1875
 Hapalothele Lenz, 1886
 Ischnothele Ausserer, 1875
 Lathrothele Benoit, 1965
 Linothele Karsch, 1879
 Masteria L.Koch, 1873
 Microhexura Crosby & Bishop, 1925
 Namirea Raven, 1984
 Phyxioschema Simon, 1889
 Stenygrocercus Simon, 1892
 Striamea Raven, 1981
 Thelechoris Karsch, 1881
 Trechona C.L.Koch, 1850

Family **Drymusidae** 1 genus

Drymusa Simon, 1891

Family **Dysderidae** 20 genera

Dasumia Thorell, 1875
 Dysdera Latreille, 1804
 Dysderocrates Deeleman-Reinhold & Deeleman, 1988
 Folkia Kratochvíl, 1970

Harpactea Bristowe, 1939
 Harpactocrates Simon, 1914
 Holissus Simon, 1882
 Hygrocrates Deeleman-Reinhold, 1988
 Mesostalita Deeleman-Reinhold, 1971
 Minotauria Kulczyński, 1903
 Parachtes Alicata, 1964
 Parastalita Absolon & Kratochvíl, 1932
 Rhode Simon, 1882
 Rhodera Deeleman-Reinhold, 1989
 Speleoharpactea Ribera, 1982
 Stalagtia Kratochvíl, 1970
 Stalita Schiödt, 1847
 Stalitella Absolon & Kratochvíl, 1932
 Stalitochara Simon, 1913
 Tedia Simon, 1882

Family **Eresidae** 10 genera

Adonea Simon, 1873
 Dorceus C.L.Koch, 1846
 Dresserus Simon, 1876
 Eresus Walckenaer, 1805
 Gandanameno Lehtinen, 1967
 Paradonea Lawrence, 1968
 Penestomus Simon, 1902
 Seothyra Purcell, 1903
 Stegodyphus Simon, 1873
 Wajane Lehtinen, 1967

Family **Filistatidae** 12 genera

Andoharano Lehtinen, 1967
 Filistata Latreille, 1810
 Filistatinella Gertsch & Ivie, 1936
 Filistatoides F.O.P.-Cambridge, 1899
 Kukulcania Lehtinen, 1967
 Malalistata Mello-Leitão, 1946
 Microfilistata Zonstein, 1990
 Pikelinia Mello-Leitão, 1946
 Pritha Lehtinen, 1967
 Sahastata Benoît, 1968
 Tricalamus Wang, 1987
 Zaitunia Lehtinen, 1967

Family **Gallieniellidae** 3 genera

Drassodella Hewitt, 1916
 Gallieniella Millot, 1947
 Legendrena Platnick, 1984

Family **Gnaphosidae** 112 genera

Adelphodrassus Rainbow, 1920
 Allodrassus Strand, 1906
 Amusia Tullgren, 1910
 Aneplasa Tucker, 1923
 Anzacia Dalmas, 1919
 Aphantaulax Simon, 1878
 Apodrassodes Vellard, 1924
 Apodrassus Chamberlin, 1916
 Apopyllus Platnick & Shadab, 1984
 Aracus Thorell, 1887
 Asemesthes Simon, 1887
 Asiabadus Roewer, 1961

Battalus Karsch, 1878
 Benoitodes Platnick, 1993
 Berinda Roewer, 1928
 Berlandina Dalmas, 1922
 Cabanadrassus Mello-Leitão, 1941
 Callilepis Westring, 1874
 Camillina Berland, 1919
 Ceryerda Simon, 1909
 Cesonia Simon, 1893
 Cladothela Kishida, 1928
 Coreodrassus Paik, 1984
 Cryptodrassus Miller, 1943
 Diaphractus Purcell, 1907
 Drassodes Westring, 1851
 Drassyllus Chamberlin, 1922
 Echemella Strand, 1906
 Echemographis Caporiacco, 1955
 Echemoides Mello-Leitão, 1938
 Echemus Simon, 1878
 Eilica Keyserling, 1891
 Epicharitus Rainbow, 1916
 Epikurtomma Tucker, 1923
 Fedotovia Charitonov, 1946
 Gertschosa Platnick & Shadab, 1981
 Gnaphosa Latreille, 1804
 Haplodrassus Chamberlin, 1922
 Hemicloea Thorell, 1870
 Hemicloeina Simon, 1893
 Herpyllus Hentz, 1832
 Homoeothela Simon, 1908
 Hypodrassodes Dalmas, 1919
 Intruda Forster, 1979
 Jacaena Thorell, 1897
 Kaitawa Forster, 1979
 Kirmaka Roewer, 1961
 Kishidaia Yaginuma, 1960
 Ladissa Simon, 1907
 Latonigena Simon, 1893
 Leptodrassus Simon, 1878
 Litopyllus Chamberlin, 1922
 Maniana Strand, 1906
 Matua Forster, 1979
 Megamyrmæcion Reuss, 1834
 Melicymnis Simon, 1885
 Mesklia Roewer, 1928
 Micaria Westring, 1851
 Microdrassus Dalmas, 1919
 Microsa Platnick & Shadab, 1977
 Miccythus Thorell, 1897
 Minosia Dalmas, 1921
 Minosiella Dalmas, 1921
 Morenilia Mello-Leitão, 1942
 Moreno Mello-Leitão, 1940
 Nauhea Forster, 1979
 Nodocion Chamberlin, 1922
 Nomisia Dalmas, 1921
 Notiodrassus Bryant, 1935
 Odontodrassus Jézéquel, 1965
 Orodassus Chamberlin, 1922
 Parabonna Mello-Leitão, 1947
 Parasyrisca Schenkel, 1963
 Phaeocedus Simon, 1893
 Poecilochroa Westring, 1874
 Prionosternum Dunn, 1951
 Pseudodrassus Caporiacco, 1935
 Pterochroa Benoit, 1977

Pterotricha Kulczyński, 1903
Pterotrichina Dalmas, 1921
Pyrnus Simon, 1880
Scopoides Platnick, 1989
Scotocesia Caporiacco, 1947
Scotognapha Dalmas, 1920
Scotophaeoides Schenkel, 1963
Scotophaeus Simon, 1893
Sergiolus Simon, 1891
Setaphis Simon, 1893
Shiragaia Paik, 1992
Sillemia Reimoser, 1935
Siruasus Roewer, 1961
Smionia Dalmas, 1920
Sosticus Chamberlin, 1922
Symphanodes Rainbow, 1916
Synaphosus Platnick & Shadab, 1980
Taieria Forster, 1979
Talanites Simon, 1893
Titus O.P.-Cambridge, 1901
Trachycosmus Simon, 1893
Trachytrema Simon, 1909
Trachyzelotes Lohmander, 1944
Trephopoda Tucker, 1923
Trichothyse Tucker, 1923
Upognampa Tucker, 1923
Urozelotes Mello-Leitão, 1938
Vectius Simon, 1897
Xenoplectus Schiapelli & Gerschman, 1958
Xerophaeus Purcell, 1907
Zavattarica Caporiacco, 1941
Zelominor Snazell & Murphy, 1997
Zelotes Gistel, 1848
Zimiromus Banks, 1914

Family **Gradungulidae** 7 genera

Gradungula Forster, 1955
Kaiya Gray, 1987
Macrogradungula Gray, 1987
Pianoa Forster, 1987
Progradungula Forster & Gray, 1979
Spelungula Forster, 1987
Tarlina Gray, 1987

Family **Hahniidae** 25 genera

Alistra Thorell, 1894
Amaloxenops Schiapelli & Gerschman, 1958
Antistea Simon, 1897
Austrohahnia Mello-Leitão, 1942
Calymmaria Chamberlin & Ivie, 1937
Cryphoea Thorell, 1870
Cybaeolus Simon, 1884
Dirksia Chamberlin & Ivie, 1942
Ethobuella Chamberlin & Ivie, 1937
Hahnia C.L.Koch, 1841
Harmiella Brignoli, 1979
Iberina Simon, 1881
Intihuatana Lehtinen, 1967
Kapanga Forster, 1970
Lizarba Roth, 1967
Muizenbergia Hewitt, 1915
Neoantistea Gertsch, 1934
Neoaviola Butler, 1929

Neocryphoea Roth, 1970
Neohahnia Mello-Leitão, 1917
Porioides Forster, 1989
Rinawa Forster, 1970
Scotospilus Simon, 1886
Tuberta Simon, 1884
Willisus Roth, 1981

Family **Halidae** 2 genera

Hala Jocqué, 1994
Tolma Jocqué, 1994

Family **Hersiliidae** 5 genera

Hersilia Savigny, 1825
Hersiliola Thorell, 1870
Murricia Simon, 1882
Tama Simon, 1882
Tamopsis Baehr & Baehr, 1987

Family **Heteropodidae** 82 genera

Adrastis Simon, 1880
Anaptomecus Simon, 1903
Anchognatha Thorell, 1881
Anchonastus Simon, 1898
Arandisa Lawrence, 1938
Barylestis Simon, 1910
Beregama Hirst, 1990
Berlandia Lessert, 1921
Carparachne Lawrence, 1962
Cebrennus Simon, 1880
Cerbalopsis Jézéquel & Junqua, 1966
Cerbalus Simon, 1897
Cercetius Simon, 1902
Chrosioderma Simon, 1897
Clastes Walckenaer, 1837
Damastes Simon, 1880
Decaphora Franganillo, 1931
Defectrix Pertunkevitch, 1925
Delena Walckenaer, 1837
Dermochrosia Mello-Leitão, 1940
Eodelena Hogg, 1902
Eusparassus Simon, 1903
Exopalystes Hogg, 1914
Geminia Thorell, 1897
Heteropoda Latreille, 1804
Hoedillus Simon, 1898
Holconia Thorell, 1877
Isopeda L.Koch, 1875
Isopedella Hirst, 1990
Keilira Hirst, 1989
Leucorchestris Lawrence, 1962
Macrinus Simon, 1887
Megaloremnius Simon, 1903
Micrommata Latreille, 1804
Microrchestris Lawrence, 1962
Neosparassus Hogg, 1903
Nisuetia Simon, 1880
Nonianus Simon, 1885
Olios Walckenaer, 1837
Orchestrella Lawrence, 1965
Origes Simon, 1896
Paenula Simon, 1896

Palystella Lawrence, 1928
 Palystes L.Koch, 1875
 Panaretella Lawrence, 1937
 Panaretidius Simon, 1906
 Panaretus Simon, 1880
 Pandercetes L.Koch, 1875
 Parhedrus Simon, 1887
 Pediana Simon, 1880
 Pleorotus Simon, 1897
 Polybetes Simon, 1896
 Prusias O.P.-Cambridge, 1893
 Prychia L.Koch, 1875
 Pseudomicrommata Järvi, 1914
 Pseudosparianthis Simon, 1887
 Remmius Simon, 1896
 Rhacocnemis Simon, 1897
 Rhitymna Simon, 1896
 Sagellula Strand, 1942
 Sampaiosia Mello-Leitão, 1930
 Sarotesius Pocock, 1898
 Seramba Thorell, 1887
 Sivalicus Dyal, 1957
 Sparianthina Banks, 1929
 Sparianthis Simon, 1880
 Spariolenus Simon, 1880
 Spatala Simon, 1897
 Staianus Simon, 1888
 Stasina Simon, 1877
 Stasinoides Berland, 1922
 Stipax Simon, 1897
 Strandiellum Kolosváry, 1934
 Thelcticopis Karsch, 1884
 Thomasettia Hirst, 1911
 Tibellomma Simon, 1903
 Torania Simon, 1886
 Tychicus Simon, 1880
 Typostola Simon, 1897
 Valonia Piza, 1939
 Vindullus Simon, 1880
 Zachria L.Koch, 1875

Family **Hexathelidae** 11 genera

Atrax O.P.-Cambridge, 1877
 Bymainiella Raven, 1978
 Hadronyche L.Koch, 1873
 Hexathele Ausserer, 1871
 Macrothele Ausserer, 1871
 Mediothele Raven & Platnick, 1978
 Paraembolides Raven, 1980
 Plesiothele Raven, 1978
 Porrhothele Simon, 1892
 Scotinoecus Simon, 1892
 Teranodes Raven, 1985

Family **Holarchaeidae** 1 genus

Holarchaea Forster, 1955

Family **Homalonychidae** 1 genus

Homalonychus Marx, 1891

Family **Huttoniidae** 1 genus

Huttonia O.P.-Cambridge, 1879

Family **Hypochilidae** 2 genera

Ectatosticta Simon, 1892
 Hypochilus Marx, 1888

Family **Idiopidae** 19 genera

Aganippe O.P.-Cambridge, 1877
 Anidiops Pocock, 1897
 Arbanitis L.Koch, 1874
 Blakistonia Hogg, 1902
 Cataxia Rainbow, 1914
 Ctenolophus Purcell, 1904
 Eucyrtops Pocock, 1897
 Galeosoma Purcell, 1903
 Genysa Simon, 1889
 Gorgyrella Purcell, 1902
 Heligmomerus Simon, 1892
 Hiboka Fage, 1922
 Idiops Perty, 1833
 Idiosoma Ausserer, 1871
 Misgolas Karsch, 1878
 Neocteniza Pocock, 1895
 Prothemenops Schwendinger, 1991
 Scalidognathus Karsch, 1891
 Segregara Tucker, 1917

Family **Lamponidae** 1 genus

Lampona Thorell, 1870

Family **Leptonetidae** 15 genera

Appaleptoneta Platnick, 1986
 Archoleptoneta Gertsch, 1974
 Barusia Kratochvil, 1978
 Calileptoneta Platnick, 1986
 Cataleptoneta Denis, 1955
 Falcileptoneta Komatsu, 1970
 Leptoneta Simon, 1872
 Leptonetela Kratochvil, 1978
 Masirana Kishida, 1942
 Neoleptoneta Brignoli, 1972
 Paraleptoneta Fage, 1913
 Protroleptoneta Deltshv, 1972
 Sarutana Komatsu, 1957
 Sulcia Kratochvil, 1938
 Teloleptoneta Ribera, 1988

Family **Linyphiidae** 489 genera

Abacoproeces Simon, 1884
 Aberdaria Holm, 1962
 Acartauchenius Simon, 1884
 Adelonetria Millidge, 1991
 Afromynoglenes Merrett & Russell-Smith, 1996
 Afroneta Holm, 1968
 Agyneta Hull, 1911
 Alaxchelicerca Butler, 1932
 Alioramus Simon, 1926
 Allomengea Strand, 1912
 Allotiso Tanasevitch, 1990
 Anacornia Chamberlin & Ivie, 1933
 Anibontes Chamberlin, 1924

- Annapolis Millidge, 1984
 Anodoration Millidge, 1991
 Anthrobia Tellkamp, 1844
 Antrohyphantes Dumitrescu, 1971
 Antronetes Millidge, 1991
 Aphileta Hull, 1920
 Aprifrontalia Oi, 1960
 Arachosinella Denis, 1958
 Araeocnus Simon, 1884
 Archaraeocnus Tanasevitch, 1987
 Arcterigone Eskov & Marusik, 1993
 Arcuphantes Chamberlin & Ivie, 1943
 Asemonetes Millidge, 1991
 Asemostera Simon, 1898
 Asperthorax Oi, 1960
 Asthenargellus Caporiacco, 1949
 Asthenargus Simon & Fage, 1922
 Atopogyna Millidge, 1984
 Atypena Simon, 1894
 Australolinyphia Wunderlich, 1976
 Bactrogyna Millidge, 1991
 Barycara Millidge, 1991
 Baryphyma Simon, 1884
 Bathyphantes Menge, 1866
 Batueta Locket, 1982
 Beauchenia Usher, 1983
 Birgerius Saaristo, 1973
 Bisetifer Tanasevitch, 1987
 Bishopiana Eskov, 1988
 Bolyphantes C.L.Koch, 1837
 Brachycerasphora Denis, 1962
 Brattia Simon, 1894
 Bursellia Holm, 1962
 Caleurema Millidge, 1991
 Callitrichia Fage, 1936
 Cameroneta Bosmans & Jocqué, 1983
 Canariellum Wunderlich, 1987
 Caracladus Simon, 1884
 Carorita Duffey & Merrett, 1963
 Cassafroneta Blest, 1979
 Catacercus Millidge, 1985
 Catonetria Millidge & Ashmole, 1994
 Caucaspisthes Tanasevitch, 1990
 Cautinella Millidge, 1985
 Caviphantes Oi, 1960
 Centromerita Dahl, 1912
 Centromerus Dahl, 1886
 Centrophantes Miller & Polenec, 1975
 Ceraticelus Simon, 1884
 Ceratinella Emerton, 1882
 Ceratinops Banks, 1905
 Ceratinopsidis Bishop & Crosby, 1930
 Ceratinopsis Emerton, 1882
 Ceratocyba Holm, 1962
 Chaetophyma Millidge, 1991
 Cheniseo Bishop & Crosby, 1935
 Chenjsides Denis, 1962
 Cherserigone Denis, 1954
 Chiangmaia Millidge, 1995
 Cineta Simon, 1884
 Clitistes Simon, 1902
 Clitolyna Simon, 1894
 Cnephalocotes Simon, 1884
 Collinsia O.P.-Cambridge, 1913
 Colonus Chamberlin, 1948
 Comorella Jocqué, 1985
 Concavocephalus Eskov, 1989
 Connithorax Eskov, 1993
 Coreorgonal Bishop & Crosby, 1935
 Cresmatoneta Simon, 1929
 Crispiphantes Tanasevitch, 1992
 Crosbyarachne Charitonov, 1937
 Crosbylonia Eskov, 1988
 Cryptolinyphia Millidge, 1991
 Ctenophysis Millidge, 1985
 Cyphonetria Millidge, 1995
 Dactylopisthes Simon, 1884
 Dactylopisthoides Eskov, 1990
 Deelemania Jocqué & Bosmans, 1983
 Delorrhypis Simon, 1884
 Diastanillus Simon, 1926
 Dicornua Oi, 1960
 Dicymbium Menge, 1868
 Didectoprocne Denis, 1949
 Diechomma Millidge, 1991
 Dietrichia Crosby & Bishop, 1933
 Diplocentria Hull, 1909
 Diplocephaloides Oi, 1960
 Diplocephalus Bertkau, 1883
 Diplophrys Millidge, 1995
 Diploplecta Millidge, 1988
 Diplostyla Emerton, 1882
 Diplothyron Millidge, 1991
 Disembolus Chamberlin & Ivie, 1933
 Dismodicus Simon, 1884
 Doenitzius Oi, 1960
 Dolabritor Millidge, 1991
 Donacochara Simon, 1884
 Drapetisca Menge, 1866
 Drepanotylus Holm, 1945
 Dresconella Denis, 1950
 Dubiaranea Mello-Leitão, 1943
 Dunedinia Millidge, 1988
 Eboria Falconer, 1910
 Eborilaira Eskov, 1989
 Elgonia Holm, 1989
 Emenista Simon, 1894
 Enguterothrix Denis, 1962
 Entelecara Simon, 1884
 Eordea Simon, 1899
 Eperigone Crosby & Bishop, 1928
 Epiceraticelus Crosby & Bishop, 1931
 Epigytholus Tanasevitch, 1995
 Episolder Tanasevitch, 1995
 Epiwubana Millidge, 1991
 Eridantes Crosby & Bishop, 1933
 Erigone Savigny, 1825
 Erigonella Dahl, 1901
 Erigonoploides Eskov, 1989
 Erigonoplus Simon, 1884
 Erigonops Scharff, 1990
 Estrandia Blauvelt, 1936
 Eulaira Chamberlin & Ivie, 1933
 Eurymorion Millidge, 1993
 Evansia O.P.-Cambridge, 1900
 Exechopsis Millidge, 1991
 Exocora Millidge, 1991
 Fageiella Kratochvil, 1934
 Falklandoglenes Usher, 1983
 Fissiscapus Millidge, 1991
 Floricomus Crosby & Bishop, 1925
 Florinda O.P.-Cambridge, 1896

- Flronia Simon, 1887
 Frontella Kulczyński, 1908
 Frontinella F.O.P.-Cambridge, 1902
 Frontinellina van Helsdingen, 1969
 Frontiphantes Wunderlich, 1987
 Gibothorax Eskov, 1989
 Gilvonanus Millidge, 1991
 Glyphesis Simon, 1926
 Gnathonargus Bishop & Crosby, 1935
 Gnathonarium Karsch, 1881
 Gnathonaroides Bishop & Crosby, 1938
 Gonatium Menge, 1868
 Gonatoraphis Millidge, 1991
 Goneatara Bishop & Crosby, 1935
 Gongyliellum Simon, 1884
 Gongylioides Oi, 1960
 Gongylidium Menge, 1868
 Grammonota Emerton, 1882
 Graphomoa Chamberlin, 1924
 Gravipalpus Millidge, 1991
 Gymnocymbium Millidge, 1991
 Habreuresis Millidge, 1991
 Halorates Hull, 1911
 Haplinis Simon, 1894
 Haplomaro Miller, 1970
 Helophora Menge, 1866
 Herbiphantes Tanasevitch, 1992
 Heterolinyphia Wunderlich, 1973
 Heterotrichoncus Wunderlich, 1970
 Hilaira Simon, 1884
 Himalaphantes Tanasevitch, 1992
 Holma Locket, 1974
 Holminaria Eskov, 1991
 Horcotes Crosby & Bishop, 1933
 Hubertella Platnick, 1989
 Hybauchenidium Holm, 1973
 Hybocoptus Simon, 1884
 Hylyphantes Simon, 1884
 Hyperafroneta Blest, 1979
 Hypomma Dahl, 1886
 Hypselistes Simon, 1894
 Hypselocara Millidge, 1991
 Hypsocephalus Millidge, 1978
 Ibadana Locket & Russell-Smith, 1980
 Iberoneta Deeleman-Reinhold, 1984
 Icarrella Brignoli, 1979
 Idionella Banks, 1893
 Incestophantes Tanasevitch, 1992
 Islandiana Braendegaard, 1932
 Itytis Strand, 1932
 Ivielum Eskov, 1988
 Jacksonella Millidge, 1951
 Jalapyphantes Gertsch & Davis, 1946
 Janetschekia Schenkel, 1939
 Johorea Locket, 1982
 Kaestneria Wiehle, 1956
 Kikimora Eskov, 1988
 Knischatiria Wunderlich, 1976
 Koinothrix Jocqué, 1981
 Kolymocyba Eskov, 1989
 Kratochviliella Miller, 1938
 Kuala Locket, 1982
 Labicymbium Millidge, 1991
 Labulla Simon, 1884
 Labullinyphia van Helsdingen, 1985
 Labullula Strand, 1913
 Laetesia Simon, 1908
 Laminacauda Millidge, 1985
 Laperousea Dalmás, 1918
 Lasiargus Kulczyński, 1894
 Latithorax Holm, 1943
 Lepthyphantes Menge, 1866
 Leptorhoptrum Kulczyński, 1894
 Leptothrix Menge, 1869
 Lessertia Smith, 1908
 Lessertinella Denis, 1947
 Liger O.P.-Cambridge, 1896
 Limoneta Bosmans & Jocqué, 1983
 Linyphantes Chamberlin & Ivie, 1942
 Linyphia Latreille, 1804
 Linyphioides Strand, 1909
 Locketidium Jocqué, 1981
 Lomaita Bryant, 1948
 Lophomma Menge, 1868
 Louisfagea Brignoli, 1971
 Lucrinus O.P.-Cambridge, 1904
 Lygarina Simon, 1894
 Machadocara Miller, 1970
 Macrargus Dahl, 1886
 Malkinella Millidge, 1991
 Malkinia Millidge, 1991
 Maorineta Millidge, 1988
 Maro O.P.-Cambridge, 1906
 Martensinus Wunderlich, 1973
 Masikia Millidge, 1984
 Maso Simon, 1844
 Masoncus Chamberlin, 1948
 Masonetta Chamberlin & Ivie, 1939
 Mecopisthes Simon, 1926
 Mecynargoides Eskov, 1988
 Mecynargus Kulczyński, 1894
 Mecynidis Simon, 1894
 Megafroneta Blest, 1979
 Meioneta Hull, 1920
 Mermessus O.P.-Cambridge, 1890
 Mesasigone Tanasevitch, 1989
 Metafroneta Blest, 1979
 Metaleptyphantes Locket, 1968
 Metamynoglenes Blest, 1979
 Metapanamomops Millidge, 1977
 Metopobactrus Simon, 1884
 Micrargus Dahl, 1886
 Microbathyphantes van Helsdingen, 1985
 Microctema Millidge, 1991
 Microctenonyx Dahl, 1886
 Microcyba Holm, 1962
 Microlinyphia Gerhardt, 1928
 Micromaso Tams-Lyche, 1954
 Miconeta Menge, 1869
 Microplanus Millidge, 1991
 Microsphalma Millidge, 1991
 Micryphantes C.L.Koch, 1833
 Millidgea Locket, 1968
 Minicia Thorell, 1875
 Minyriolus Simon, 1884
 Mioxena Simon, 1926
 Mitrager van Helsdingen, 1985
 Moebelia Dahl, 1886
 Monocephalus Smith, 1906
 Monocerellus Tanasevitch, 1983
 Montilaira Chamberlin, 1922

- Moreiraxena Miller, 1970
 Mycula Schikora, 1994
 Myrmecomelix Millidge, 1993
 Mythoplastoides Crosby & Bishop, 1933
 Nanavia Chamberlin & Ivie, 1933
 Nasoona Locket, 1982
 Nematogmus Simon, 1884
 Nenilinium Eskov, 1988
 Nentwigia Millidge, 1995
 Neocautinella Baert, 1990
 Neoeburnella Koçak, 1986
 Neomaso Forster, 1970
 Neonesiotes Millidge, 1991
 Neriene Blackwall, 1833
 Nesioneta Millidge, 1991
 Notholephyphantes Millidge, 1985
 Nothophantes Merrett & Stevens, 1995
 Notiohyphantes Millidge, 1985
 Notiomaso Banks, 1914
 Notioscopus Simon, 1884
 Notiothauma Millidge, 1991
 Novafroneta Blest, 1979
 Novafrontina Millidge, 1991
 Novalaetesia Millidge, 1988
 Oaphantes Chamberlin & Ivie, 1943
 Obrimona Strand, 1934
 Ochronetria Millidge, 1991
 Oculocornia Oligier, 1985
 Oedothorax Bertkau, 1883
 Oia Wunderlich, 1973
 Oilinyphia Ono & Saito, 1989
 Oinia Eskov, 1984
 Onychembolus Millidge, 1985
 Ophrynia Jocqué, 1981
 Oreocyba Holm, 1962
 Oreonetides Strand, 1901
 Oreophantes Eskov, 1984
 Origanates Crosby & Bishop, 1933
 Ostearius Hull, 1911
 Pachydelphus Jocqué & Bosmans, 1983
 Pacifiphantes Eskov & Marusik, 1993
 Palaeohyphantes Millidge, 1984
 Panamomops Simon, 1884
 Paracornicularia Crosby & Bishop, 1931
 Paraeboria Eskov, 1990
 Parafroneta Blest, 1979
 Paraglyphesis Eskov, 1991
 Paragongylidiellum Wunderlich, 1973
 Paraletes Millidge, 1991
 Parameioneta Locket, 1982
 Paranasoona Heimer, 1984
 Parasisis Eskov, 1984
 Paratapinocyba Saito, 1986
 Paro Berland, 1941
 Patagoneta Millidge, 1985
 Pelecopsidis Bishop & Crosby, 1935
 Pelecopsis Simon, 1864
 Pelidida Simon, 1898
 Peponocranium Simon, 1884
 Perimonoides Schenkel, 1963
 Perlongipalpus Eskov & Marusik, 1991
 Perregrinus Tanasevitch, 1992
 Perro Tanasevitch, 1992
 Phanetta Keyserling, 1886
 Phlathothrata Crosby & Bishop, 1933
 Phonophilus Ehrenberg, 1828
 Piesocalus Simon, 1894
 Pimoa Chamberlin & Ivie, 1943
 Pityohyphantes Simon, 1929
 Plaesianillus Simon, 1926
 Plesiophantes Heimer, 1981
 Pocadicnemis Simon, 1884
 Pocobletus Simon, 1894
 Poecilafroneta Blest, 1979
 Poeciloneta Kulczyński, 1894
 Porrhomma Simon, 1884
 Prinerigone Millidge, 1988
 Priperia Simon, 1903
 Priscipalpus Millidge, 1991
 Procerocymbium Eskov, 1989
 Proelauna Jocqué, 1981
 Proislandiana Tanasevitch, 1985
 Promynoglenes Blest, 1979
 Pronasoona Millidge, 1995
 Pronopius Menge, 1869
 Protoerigone Blest, 1979
 Pseudafroneta Blest, 1979
 Pseudocarorita Wunderlich, 1980
 Pseudocyba Tanasevitch, 1984
 Pseudogonatium Strand, 1901
 Pseudohilaira Eskov, 1990
 Pseudomaro Denis, 1966
 Pseudomaso Locket & Russell-Smith, 1980
 Pseudomicrocentria Miller, 1970
 Pseudotyphistes Brignoli, 1972
 Pseudowubana Eskov & Marusik, 1992
 Psilocymbium Millidge, 1991
 Racata Millidge, 1995
 Rhabdogyna Millidge, 1985
 Ringina Tams-Lyche, 1954
 Saaristoa Millidge, 1978
 Saloca Simon, 1926
 Satilatlas Keyserling, 1886
 Savignia Blackwall, 1833
 Schistogyna Millidge, 1991
 Sciastes Bishop & Crosby, 1938
 Scirites Bishop & Crosby, 1938
 Scironis Bishop & Crosby, 1938
 Scolecura Millidge, 1991
 Scolopembolus Bishop & Crosby, 1938
 Scotargus Simon, 1913
 Scotinotylus Simon, 1884
 Scotoneta Simon, 1910
 Scylaceus Bishop & Crosby, 1938
 Scyletria Bishop & Crosby, 1938
 Selenyphantes Gertsch & Davis, 1946
 Semljicola Strand, 1906
 Sibiroyba Eskov & Marusik, 1993
 Silometopoides Eskov, 1990
 Silometopus Simon, 1926
 Simplicistilus Locket, 1968
 Sinoria Bishop & Crosby, 1938
 Sintula Simon, 1884
 Sisicottus Bishop & Crosby, 1938
 Sisicus Bishop & Crosby, 1938
 Sisis Bishop & Crosby, 1938
 Sisyrbes Bishop & Crosby, 1938
 Sitalcas Bishop & Crosby, 1938
 Smermisia Simon, 1894
 Smodix Bishop & Crosby, 1938
 Solenysa Simon, 1894
 Soucron Crosby & Bishop, 1936

Soudinus Crosby & Bishop, 1936
 Souessa Crosby & Bishop, 1936
 Souessoula Crosby & Bishop, 1936
 Sougambus Crosby & Bishop, 1936
 Souidas Crosby & Bishop, 1936
 Soulgas Crosby & Bishop, 1936
 Spanioplanus Millidge, 1991
 Sphecozone O.P.-Cambridge, 1870
 Spirembolus Chamberlin, 1920
 Stemonyphantes Menge, 1866
 Sthelota Simon, 1894
 Stictonanus Millidge, 1991
 Strandella Oi, 1960
 Strongyliceptus Fage, 1936
 Subbekasha Millidge, 1984
 Syedra Simon, 1884
 Symmigma Crosby & Bishop, 1933
 Tachygyna Chamberlin & Ivie, 1939
 Tallusia Lehtinen & Saaristo, 1972
 Tapinocyba Simon, 1884
 Tapinocyboides Wiehle, 1960
 Tapinopa Westring, 1851
 Taranucus Simon, 1884
 Tennesseeellum Petrunkevitch, 1925
 Thainetes Millidge, 1995
 Thaiphantes Millidge, 1995
 Thaleria Tanasevitch, 1984
 Thapsagus Simon, 1894
 Theonina Simon, 1929
 Thyreobaeus Simon, 1888
 Thyreosthenius Simon, 1884
 Tibiaster Tanasevitch, 1987
 Tibioplodes Eskov & Marusik, 1991
 Tibioplus Chamberlin & Ivie, 1947
 Tiso Simon, 1884
 Tmeticides Strand, 1907
 Tmeticus Menge, 1868
 Tomohyphantes Millidge, 1995
 Toschia Caporiacco, 1949
 Totua Keyserling, 1891
 Trachelocampus Simon, 1884
 Trachyneta Holm, 1968
 Traematosisis Bishop & Crosby, 1938
 Trematocephalus Dahl, 1886
 Trichoncoides Denis, 1950
 Trichoncus Simon, 1884
 Trichopterna Kulczyński, 1894
 Triplogyna Millidge, 1991
 Troglolyphantes Joseph, 1882
 Troxochrota Kulczyński, 1894
 Troxochrus Simon, 1884
 Tubercithorax Eskov, 1988
 Tunagyna Chamberlin & Ivie, 1933
 Turbinellina Millidge, 1993
 Turinyphia van Helsdingen, 1982
 Tutaibo Chamberlin, 1916
 Tybaertiella Jocqué, 1979
 Typhistes Simon, 1894
 Typhlonyphia Kratochvil, 1936
 Typhochrestinus Eskov, 1990
 Typhochrestoides Eskov, 1990
 Typhochrestus Simon, 1884
 Uahuka Berland, 1935
 Uapou Berland, 1935
 Ulugurella Jocqué & Scharff, 1986

Ummeliata Strand, 1942
 Valdiviella Millidge, 1985
 Vermontia Millidge, 1984
 Vesicapalpus Millidge, 1991
 Victorium Eskov, 1988
 Wabasso Millidge, 1984
 Walckenaeria Blackwall, 1833
 Wiehlea Braun, 1959
 Wiehlenarius Eskov, 1990
 Wubana Chamberlin, 1919
 Wubanoides Eskov, 1986
 Yakutopus Eskov, 1990
 Zerogone Eskov & Marusik, 1993
 Zilephus Simon, 1902
 Zornella Jackson, 1932
 Zygottus Chamberlin, 1948

Family **Liocranidae** 44 genera

Agraecina Simon, 1932
 Agroeca Westring, 1862
 Andromma Simon, 1893
 Apostenus Westring, 1851
 Argistes Simon, 1897
 Attacobius Mello-Leitão, 1925
 Brachyanillus Simon, 1913
 Cambalida Simon, 1910
 Cerrutia Roewer, 1960
 Chemmis Simon, 1898
 Coryssiphus Simon, 1903
 Cybaeodes Simon, 1878
 Donuea Strand, 1932
 Drassinella Banks, 1904
 Ferrieria Tullgren, 1901
 Hesperocranum Ubick & Platnick, 1991
 Heterochemmis F.O.P.-Cambridge, 1900
 Itatsina Kishida, 1930
 Laudetia Gertsch, 1941
 Liocranum L.Koch, 1866
 Liparochrysis Simon, 1909
 Macedoniella Drensky, 1935
 Mardonia Thorell, 1897
 Mesiotelus Simon, 1897
 Mesobria Simon, 1898
 Messapus Simon, 1898
 Montebello Hogg, 1914
 Orthobula Simon, 1896
 Otacilia Thorell, 1897
 Palaetyra Simon, 1898
 Parachemmis Chickering, 1937
 Paratus Simon, 1898
 Phonotimpus Gertsch & Davis, 1940
 Phrurolithus C.L.Koch, 1839
 Phruonellus Chamberlin, 1921
 Phrurotimpus Chamberlin & Ivie, 1935
 Piabuna Chamberlin & Ivie, 1933
 Rhaeboctesis Simon, 1897
 Scotina Menge, 1873
 Scotinella Banks, 1911
 Sesieutes Simon, 1896
 Teutamus Thorell, 1890
 Thysanina Simon, 1910
 Titiotus Simon, 1897

Family **Liphistiidae** 2 genera

Heptathela Kishida, 1923

Liphistius Schiödt, 1849

Family **Lycosidae** 96 genera

Adelocosa Gertsch, 1973

Agalenocosa Mello-Leitão, 1944

Algidus Simon, 1898

Allocosa Banks, 1900

Alopecosa Simon, 1885

Alopecosella Roewer, 1960

Amblyothele Simon, 1910

Anomalomma Simon, 1890

Anomalosa Roewer, 1960

Arctosa C.L.Koch, 1847

Arctosippa Roewer, 1960

Arctosomma Roewer, 1960

Arctoria Thorell, 1877

Artoriella Roewer, 1960

Artoriellula Roewer, 1960

Aulonia C.L.Koch, 1847

Auloniella Roewer, 1960

Brevilabus Strand, 1908

Bristowiella Saaristo, 1980

Caporiaccosa Roewer, 1960

Chorilycosa Roewer, 1960

Crocodylosa Caporiacco, 1947

Cynosa Caporiacco, 1933

Dalmasicosa Roewer, 1960

Dejerosa Roewer, 1960

Diapontia Keyserling, 1876

Dingosa Roewer, 1955

Dolocosa Roewer, 1960

Donacosa Alderweireldt & Jocqué, 1991

Edenticosa Roewer, 1960

Evippa Simon, 1882

Evippomma Roewer, 1959

Geolycosa Montgomery, 1904

Gladicosa Brady, 1987

Gnatholycosa Mello-Leitão, 1940

Hesperocosa Gertsch & Wallace, 1937

Hippasa Simon, 1885

Hippasosa Roewer, 1960

Hogna Simon, 1885

Hognoides Roewer, 1960

Hyaenosa Caporiacco, 1940

Hygrolycosa Dahl, 1908

Isohogna Roewer, 1960

Loculla Simon, 1909

Lycorma Simon, 1885

Lycosa Latreille, 1804

Lycosella Thorell, 1890

Lycosula Roewer, 1960

Lynxosa Roewer, 1960

Lysania Thorell, 1890

Malimbosa Roewer, 1960

Margonia Hippa & Lehtinen, 1983

Megarctosa Caporiacco, 1948

Melloicosa Roewer, 1960

Melocosa Gertsch, 1937

Molitorosa Roewer, 1960

Mustelicosa Roewer, 1960

Ocyale Savigny, 1825

Orinocosa Chamberlin, 1916

Orthocosa Roewer, 1960

Paratrochosina Roewer, 1960

Pardosa C.L.Koch, 1847

Pardosella Caporiacco, 1939

Passiena Thorell, 1890

Pavocosa Roewer, 1960

Pirata Sundevall, 1833

Piratosia Roewer, 1960

Porrimosa Roewer, 1960

Proevippa Purcell, 1903

Prolycosides Mello-Leitão, 1942

Pseudevippa Simon, 1909

Pterartoria Purcell, 1903

Pterartoriola Roewer, 1959

Satta Lehtinen & Hippa, 1979

Schizocosa Chamberlin, 1904

Shapna Hippa & Lehtinen, 1983

Sosippus Simon, 1888

Syroloma Simon, 1900

Tasmanicosa Roewer, 1959

Trabaeola Roewer, 1960

Trabea Simon, 1876

Trabeops Roewer, 1959

Trebacosa Dondale & Redner, 1981

Tricassa Simon, 1910

Trochosa C.L.Koch, 1847

Trochosippa Roewer, 1960

Trochosula Roewer, 1960

Varacosa Chamberlin & Ivie, 1942

Venator Hogg, 1900

Venonia Thorell, 1894

Vesubia Simon, 1909

Wadicosa Zyuzin, 1985

Xerolycosa Dahl, 1908

Zantheres Thorell, 1887

Zenonina Simon, 1898

Zoica Simon, 1898

Family **Malkaridae** 4 genera

Carathea Moran, 1986

Chilenodes Platnick & Forster, 1987

Malkara Davies, 1980

Sternodes Butler, 1929

Family **Mecicobothriidae** 4 genera

Hexura Simon, 1884

Hexurella Gertsch & Platnick, 1979

Mecicobothrium Holmberg, 1882

Megahexura Kaston, 1972

Family **Mecysmauchenidae** 7 genera

Aotearoa Forster & Platnick, 1984

Chilarchaea Forster & Platnick, 1984

Mecysmauchenoides Forster & Platnick, 1984

Mecysmauchenius Simon, 1884

Mesarchaea Forster & Platnick, 1984

Semysmauchenius Forster & Platnick, 1984

Zearchaea Wilton, 1946

Family **Micropholcommatidae** 8 genera

Eterosonycha Butler, 1932

Micropholcomma Crosby & Bishop, 1927

Olgania Hickman, 1979
 Parapua Forster, 1959
 Pua Forster, 1959
 Teutoniella Brignoli, 1981
 Textricella Hickman, 1945
 Tricellina Forster & Platnick, 1989

Family **Microstigmatidae** 4 genera

Micromygale Platnick & Forster, 1982
 Microstigmata Strand, 1932
 Ministigmata Raven & Platnick, 1981
 Pseudonemesia Caporiacco, 1955

Family **Migidae** 9 genera

Calathotarsus Simon, 1903
 Heteromigas Hogg, 1902
 Mallecomigas Goloboff & Platnick, 1987
 Micromesomma Pocock, 1895
 Migas L.Koch, 1873
 Moggridgea O.P.-Cambridge, 1875
 Paramigas Pocock, 1895
 Poecilomigas Simon, 1903
 Thyropoeus Pocock, 1895

Family **Mimetidae** 12 genera

Arkys Walckenaer, 1837
 Arocha Simon, 1893
 Arochoides Mello-Leitão, 1835
 Australomimetes Heimer, 1986
 Ero C.L.Koch, 1837
 Gelanor Thorell, 1870
 Kratochvilia Strand, 1934
 Melaenosia Simon, 1906
 Mimetes Hentz, 1832
 Oarces Simon, 1879
 Phobetinus Simon, 1895
 Reo Brignoli, 1979

Family **Miturgidae** 23 genera

Camptostichomma Karsch, 1891
 Devendra Lehtinen, 1967
 Diaprogapta Simon, 1909
 Eutichurus Simon, 1896
 Hebrithale Berland, 1938
 Machadonia Lehtinen, 1967
 Miturga Thorell, 1870
 Pacificana Hogg, 1904
 Palicanus Thorell, 1897
 Parapostenus Lessert, 1923
 Paratyle Simon, 1896
 Phanotea Simon, 1896
 Philisca Simon, 1884
 Prochora Simon, 1885
 Radulphius Keyserling, 1891
 Raecius Simon, 1892
 Strotarchus Simon, 1888
 Syrisca Simon, 1885
 Syspira Simon, 1895
 Teminius Keyserling, 1887
 Uduba Simon, 1880
 Uliodon L.Koch, 1873
 Zorodictyna Strand, 1907

Family **Mysmenidae** 22 genera

Acrobleps Hickman, 1979
 Anjouanella Baert, 1986
 Brucharachne Mello-Leitão, 1925
 Calodipoena Gertsch & Davis, 1936
 Calomyspoena Baert & Maelfait, 1983
 Cepheia Simon, 1894
 Iardinis Simon, 1899
 Isela Griswold, 1985
 Itapua Baert, 1984
 Kekenboschiella Baert, 1982
 Kilifina Baert & Murphy, 1992
 Leviola Miller, 1970
 Maymena Gertsch, 1960
 Microdipoena Banks, 1895
 Mysmena Simon, 1894
 Mysmenella Brignoli, 1980
 Mysmenopsis Simon, 1897
 Phricotelus Simon, 1895
 Synaphris Simon, 1894
 Tamasesia Marples, 1955
 Taphiassa Simon, 1880
 Trogloneta Simon, 1922

Family **Nemesiidae** 37 genera

Acanthogonatus Karsch, 1880
 Aname L.Koch, 1873
 Atmetochilus Simon, 1887
 Brachythele Ausserer, 1871
 Calisoga Chamberlin, 1937
 Chaco Tullgren, 1905
 Damarchus Thorell, 1892
 Diplotelopsis Tullgren, 1905
 Entypesa Simon, 1902
 Hermacha Simon, 1889
 Hermachura Mello-Leitão, 1923
 Ixamatus Simon, 1887
 Kwonkan Main, 1983
 Lepthercus Purcell, 1902
 Merredinia Main, 1983
 Mexentypesa Raven, 1987
 Mygaloides Nicolet, 1849
 Namea Raven, 1984
 Nemesia Savigny, 1825
 Neodiplothele Mello-Leitão, 1917
 Pionothele Purcell, 1902
 Prorachias Mello-Leitão, 1924
 Pselligmus Simon, 1892
 Pseudoteyle Main, 1985
 Pycnothele Chamberlin, 1917
 Rachias Simon, 1892
 Raveniola Zonstein, 1987
 Spelocteniza Gertsch, 1982
 Spiroctenus Simon, 1889
 Stanwellia Rainbow & Pulleine, 1918
 Stenoterommata Holmberg, 1881
 Teyl Main, 1975
 Teylodes Main, 1985
 Troglodiplura Main, 1969
 Xamiatus Raven, 1981
 Xenonemesia Goloboff, 1989
 Yilgarnia Main, 1986

Family **Neolanidae** 1 genus

Neolana Forster & Wilton, 1973

Family **Nesticidae** 7 genera

Carpathonesticus Lehtinen & Saaristo, 1980

Cyclocarcina Komatsu, 1942

Eidmannella Roewer, 1935

Gaucelmus Keyserling, 1884

Nesticella Lehtinen & Saaristo, 1980

Nesticus Thorell, 1869

Typhlonesticus Kulczyński, 1914

Family **Nicodamidae** 2 genera

Megadictyna Dahl, 1906

Nicodamus Simon, 1887

Family **Ochyroceratidae** 9 genera

Althepus Torell, 1898

Dundocera Machado, 1951

Lundacera Machado, 1951

Merizocera Fage, 1912

Ochyrocera Simon, 1891

Psilodermes Simon, 1892

Simonocera Brignoli, 1979

Speocera Berland, 1914

Theotima Simon, 1892

Family **Oecobiidae** 6 genera

Oecobius Lucas, 1846

Paroecobius Lamoral, 1981

Platoecobius Chamberlin & Ivie, 1935

Uroctea Dufour, 1820

Urocteana Roewer, 1961

Uroecobius Kullmann & Zimmernann, 1976

Family **Oonopidae** 54 genera

Anophthalmoonops Benoit, 1976

Aprusia Simon, 1893

Australoonops Hewitt, 1915

Blanioonops Simon & Fage, 1922

Brignolia Dumitrescu & Georgescu, 1983

Caecoonops Benoit, 1964

Calculus Purcell, 1910

Camptoscaphiella Caporiacco, 1934

Decuana Dumitrescu & Georgescu, 1987

Dysderina Simon, 1891

Dysderoides Fage, 1946

Epectris Simon, 1893

Eusimonia Dumitrescu & Georgescu, 1987

Gamasomorpha Karsch, 1881

Grymeus Harvey, 1987

Heteroonops Dalmas, 1916

Hypnoonops Benoit, 1977

Hytanis Simon, 1892

Ischnothyreus Simon, 1892

Kapitia Forster, 1956

Kijabe Berland, 1914

Lionneta Benoit, 1979

Lucetia Dumitrescu & Georgescu, 1983

Marsupopaea Cooke, 1972

Myrmecoscaphiella Mello-Leitão, 1926

Myrmopopaea Reimoser, 1933

Neoxyphinus Birabén, 1953

Nephrochirus Simon, 1910

Oonopinus Simon, 1892

Oonopoides Bryant, 1940

Oonops Templeton, 1835

Opopaea Simon, 1891

Orchestina Simon, 1882

Pelcinus Simon, 1891

Pescennina Simon, 1903

Plectoptilus Simon, 1905

Prodysderina Dumitrescu & Georgescu, 1987

Pseudoscaphiella Simon, 1970

Pseudotriaeris Brignoli, 1974

Scaphiella Simon, 1891

Silhouettella Benoit, 1979

Stenoonops Simon, 1981

Sulsula Simon, 1882

Tapinesthis Simon, 1914

Telchius Simon, 1892

Termitoonops Benoit, 1964

Triaeris Simon, 1891

Wanops Chamberlin & Ivie, 1938

Xestaspis Simon, 1884

Xiombarg Brignoli, 1979

Xyccarph Brignoli, 1978

Xyphinus Simon, 1893

Yumates Chamberlin, 1924

Zyngoonops Benoit, 1977

Family **Orsolobidae** 27 genera

Afrilobus Griswold & Platnick, 1987

Anopsolobus Forster & Platnick, 1985

Ascuta Forster, 1956

Australobus Forster & Platnick, 1985

Azanalobus Griswold & Platnick, 1987

Bealeyia Forster & Platnick, 1985

Chileolobus Forster & Platnick, 1985

Cornifalx Hickman, 1979

Dugdalea Forster & Platnick, 1985

Duripelta Forster, 1956

Falklandia Forster & Platnick, 1985

Hickmanolobus Forster & Platnick, 1985

Mallecolobus Forster & Platnick, 1985

Maoriata Forster & Platnick, 1985

Orongia Forster & Platnick, 1985

Orsolobus Simon, 1893

Osornolobus Forster & Platnick, 1985

Paralobus Forster & Platnick, 1985

Pounamuella Forster & Platnick, 1985

Subantarctia Forster, 1955

Tangata Forster & Platnick, 1985

Tasmanoonops Hickman, 1930

Tautukua Forster & Platnick, 1985

Turretia Forster & Platnick, 1985

Waiporia Forster & Platnick, 1985

Waipoua Forster & Platnick, 1985

Wiltonia Forster & Platnick, 1985

Family **Oxyopidae** 9 genera

Hamataliwa Keyserling, 1887

Hostus Simon, 1898

Megullia Thorell, 1897
 Oxyopes Latreille, 1804
 Peucetia Thorell, 1869
 Pseudohostus Rainbow, 1915
 Schaenioscelis Simon, 1898
 Tapinillus Simon, 1898
 Tapponia Simon, 1885

Family Palpimanidae 14 genera

Anisaedus Simon, 1893
 Badia Roewer, 1961
 Boagrius Simon, 1893
 Chedima Simon, 1873
 Diaphorocellus Simon, 1893
 Fernandezina Birabén, 1951
 Hybosida Simon, 1897
 Ikuma Lawrence, 1938
 Otiothops MacLeay, 1839
 Palpimanus Dufour, 1820
 Sarascelis Simon, 1897
 Scelidocteus Simon, 1907
 Scelidomachus Pocock, 1899
 Steriphopus Simon, 1887

Family Pararchaeidae 1 genus

Pararchaea Forster, 1955

Family Paratropididae 4 genera

Anisaspis Simon, 1891
 Anisaspoides F.O.P.-Cambridge, 1896
 Melloina Brignoli, 1985
 Paratropis Simon, 1889

Family Philodromidae 30 genera

Apollophanes O.P.-Cambridge, 1898
 Bacillocnemis Mello-Leitão, 1938
 Berlandiella Mello-Leitão, 1929
 Catuna Mello-Leitão, 1940
 Cleocnemis Simon, 1886
 Ebo Keyserling, 1884
 Fageia Mello-Leitão, 1929
 Gephyrellula Strand, 1932
 Gephyrina Simon, 1895
 Gephyrota Strand, 1932
 Hirriusa Strand, 1932
 Metacleocnemis Mello-Leitão, 1929
 Pagiopalus Simon, 1900
 Paracleocnemis Schiapelli & Gerschman, 1942
 Paratibellus Simon, 1932
 Petricus Simon, 1886
 Philodromops Mello-Leitão, 1943
 Philodromus Walckenaer, 1826
 Procleocnemis Mello-Leitão, 1929
 Proernus Simon, 1900
 Psellonus Simon, 1897
 Pseudopsellonus Balogh, 1936
 Quemedice Mello-Leitão, 1942
 Senoculifer Balogh, 1936
 Suemus Simon, 1895
 Thanatus C.L.Koch, 1837
 Tibellinus Simon, 1910
 Tibellus Simon, 1875

Tibitanus Simon, 1907
 Vacchellia Caporiacco, 1935

Family Pholcidae 39 genera

Anopsicus Chamberlin & Ivie, 1938
 Artema Walckenaer, 1837
 Belisana Thorell, 1898
 Blechroscelis Simon, 1893
 Bryantina Brignoli, 1985
 Calpnita Simon, 1892
 Ceratopholcus Spassky, 1934
 Coryssocnemis Simon, 1893
 Crossopriza Simon, 1893
 Gertschiola Brignoli, 1981
 Hedypsilus Simon, 1893
 Holocneminus Berland, 1942
 Holocnemus Simon, 1875
 Hoplopholcus Kulczyński, 1908
 Ibotyporanga Mello-Leitão, 1944
 Leptopholcus Simon, 1893
 Litoporus Simon, 1893
 Mecolaesthus Simon, 1893
 Metagonia Simon, 1893
 Micromerys Bradley, 1877
 Micropholcus Deeleman-Reinhold & Prinsen, 1987
 Modisimus Simon, 1893
 Myrmidonella Berland, 1919
 Mystes Bristowe, 1938
 Ninetis Simon, 1890
 Panjange Deeleman-Reinhold & Deeleman, 1983
 Paramicromerys Millot, 1946
 Pholciella Roewer, 1960
 Pholcoides Roewer, 1960
 Pholcophora Banks, 1896
 Pholcus Walckenaer, 1805
 Physocyclus Simon, 1893
 Psilochorus Simon, 1893
 Smeringopina Kraus, 1957
 Smeringopus Simon, 1890
 Spermophora Hentz, 1841
 Systemita Simon, 1893
 Trichocyclus Simon, 1908
 Uthina Simon, 1893

Family Pisauridae 54 genera

Afropisaura Blandin, 1976
 Aglaoctenus Tullgren, 1905
 Ancyloletes Bertkau, 1880
 Anoteropsis L.Koch, 1878
 Archipirata Simon, 1898
 Architis Simon, 1898
 Campostichommides Strand, 1911
 Caripetella Strand, 1926
 Charminus Thorell, 1899
 Chiasmopes Pavesi, 1883
 Cispinilus Roewer, 1955
 Cispinus Simon, 1898
 Cladycnis Simon, 1898
 Conakrya Schmidt, 1956
 Dendrolycosa Doleschall, 1859
 Dolomedes Latreille, 1804
 Eucamptopus Pocock, 1900
 Euprostenops Pocock, 1897

Euprosthénopsis Blandin, 1974
 Eurychoera Thorell, 1897
 Hesydrimorpha Strand, 1911
 Hygropoda Thorell, 1895
 Hypsithylla Simon, 1903
 Ilipula Simon, 1903
 Inola Davies, 1982
 Maypaciús Simon, 1898
 Megadolomedes Davies & Raven, 1980
 Nilus O.P.-Cambridge, 1876
 Nukuhiva Berland, 1935
 Papakula Strand, 1911
 Paracladycnis Blandin, 1979
 Perenethis L.Koch, 1878
 Phalaecops Roewer, 1955
 Pisaura Simon, 1885
 Pisaurellus Roewer, 1961
 Pisaurina Simon, 1898
 Polyboea Thorell, 1895
 Ransonia Blandin, 1979
 Rothus Simon, 1898
 Shinobius Yaginuma, 1991
 Staberius Simon, 1898
 Stoliczka O.P.-Cambridge, 1885
 Tallonia Simon, 1889
 Tapinothele Simon, 1898
 Tapinothelella Strand, 1909
 Tapinothelops Roewer, 1955
 Tetragonophthalma Karsch, 1878
 Thalassiopsis Roewer, 1955
 Thalassius Simon, 1885
 Thaumasia Perty, 1833
 Tinus F.O.P.-Cambridge, 1901
 Voraptipus Roewer, 1955
 Vuattouxia Blandin, 1979
 Walrencea Blandin, 1979

Family **Plectreuridae** 2 genera

Kibramoa Chamberlin, 1924
 Plectreurys Simon, 1893

Family **Prodidomidae** 27 genera

Achalaicola Mello-Leitão, 1943
 Anagraphis Simon, 1893
 Anagrina Berland, 1920
 Austrodomus Lawrence, 1947
 Caudalia Alayón, 1980
 Cryptoeirithus Rainbow, 1915
 Eleleis Simon, 1893
 Encoptarthria Main, 1954
 Honunius Simon, 1908
 Hyltonia Birabén, 1954
 Katumbea Cooke, 1964
 Lygromma Simon, 1892
 Lygrommatoides Strand, 1918
 Molycria Simon, 1887
 Myandra Simon, 1887
 Neozimiris Simon, 1903
 Oltaclaea Mello-Leitão, 1940
 Plutonodomus Cooke, 1964
 Prodida Dalmas, 1918
 Prodidomus Hentz, 1847
 Purcelliana Cooke, 1964
 Theuma Simon, 1893

Theumella Strand, 1906
 Tivodrassus Chamberlin & Ivie, 1936
 Tricongius Simon, 1892
 Zimirina Dalmas, 1918
 Zimiris Simon, 1882

Family **Psechridae** 4 genera

Fecenia Simon, 1887
 Haurokoa Forster & Wilton, 1973
 Poaka Forster & Wilton, 1973
 Psechrus Thorell, 1878

Family **Salticidae** 481 genera

Abracadabrella Żabka, 1991
 Acragas Simon, 1900
 Aculeobreda Caporiacco, 1955
 Admestina Peckham & Peckham, 1888
 Admesturius Galiano, 1988
 Adoxotoma Simon, 1909
 Aelurillus Simon, 1884
 Afrobeata Caporiacco, 1941
 Agassa Simon, 1901
 Agelista Simon, 1900
 Agobardus Keyserling, 1885
 Agorius Thorell, 1877
 Aillutticus Galiano, 1987
 Akela Peckham & Peckham, 1896
 Albionella Chickering, 1946
 Alcmene C.L.Koch, 1846
 Alfenus Simon, 1902
 Allococalodes Wanless, 1982
 Allodecta Bryant, 1950
 Amphidraus Simon, 1900
 Amycus C.L.Koch, 1846
 Anarrhotus Simon, 1902
 Anasaitis Bryant, 1950
 Anaurus Simon, 1900
 Anicius Chamberlin, 1925
 Anokopsis Bauab & Soares, 1980
 Antillattus Bryant, 1943
 Apamamia Roewer, 1944
 Aphirape C.L.Koch, 1850
 Arachnomura Mello-Leitão, 1917
 Arachnotermes Mello-Leitão, 1928
 Araegeus Simon, 1901
 Arasia Simon, 1901
 Artabrus Simon, 1902
 Aruana Strand, 1911
 Asaphobelis Simon, 1902
 Asaracus C.L.Koch, 1846
 Ascyltus Karsch, 1878
 Asemonea O.P.-Cambridge, 1869
 Ashtabula Peckham & Peckham, 1894
 Astia L.Koch, 1879
 Atelurius Simon, 1901
 Athamas O.P.-Cambridge, 1877
 Atomosphyrus Simon, 1902
 Attulus Simon, 1889
 Augustaea Szombathy, 1915
 Avarua Marples, 1955
 Avitus Peckham & Peckham, 1896
 Bacelarella Berland & Millot, 1941
 Bagheera Peckham & Peckham, 1896

- Ballognatha Caporiacco, 1935
 Ballus C.L.Koch, 1850
 Balmaceda Peckham & Peckham, 1894
 Banksetosa Chickering, 1946
 Baryphas Simon, 1902
 Bathippus Thorell, 1892
 Bavia Simon, 1877
 Baviola Simon, 1897
 Beata Peckham & Peckham, 1895
 Belippo Simon, 1909
 Belliena Simon, 1902
 Bellota Peckham & Peckham, 1892
 Bianor Peckham & Peckham, 1885
 Bindax Thorell, 1892
 Blaisea Simon, 1902
 Bocus Peckham & Peckham, 1892
 Bokokius Roewer, 1942
 Brancus Simon, 1902
 Breda Peckham & Peckham, 1894
 Bredana Gertsch, 1936
 Brettus Thorell, 1895
 Bristowia Reimoser, 1934
 Bryantella Chickering, 1946
 Bulolia Žabka, 1996
 Bythocrotus Simon, 1903
 Canama Simon, 1903
 Capidava Simon, 1902
 Carabella Chickering, 1946
 Caribattus Bryant, 1950
 Carrhotus Thorell, 1891
 Ceglusa Thorell, 1895
 Ceriomura Simon, 1901
 Cerionesta Simon, 1901
 Chalcolecta Simon, 1884
 Chalcoscirtus Bertkau, 1880
 Chalcotropis Simon, 1902
 Chapoda Peckham & Peckham, 1896
 Charippus Thorell, 1895
 Cheliceroides Žabka, 1985
 Cheliferoides F.O.P.-Cambridge, 1901
 Chinoscopus Simon, 1901
 Chira Peckham & Peckham, 1896
 Chirothecia Taczanowski, 1878
 Chloridusa Simon, 1902
 Chrysilla Thorell, 1887
 Clynotis Simon, 1901
 Clynotoides Mello-Leitão, 1944
 Cobanus F.O.P.-Cambridge, 1900
 Cocalodes Pocock, 1897
 Cocalus C.L.Koch, 1846
 Coccorchestes Thorell, 1881
 Colaxes Simon, 1900
 Colyttus Thorell, 1891
 Commoris Simon, 1902
 Compsodecta Simon, 1903
 Consingis Simon, 1900
 Copocrossa Simon, 1901
 Corambis Simon, 1901
 Corcovetella Galiano, 1975
 Coryphasia Simon, 1902
 Corythalia C.L.Koch, 1850
 Cosmophasis Simon, 1901
 Cotinusa Simon, 1900
 Curubis Simon, 1902
 Cylistella Simon, 1901
 Cyllostania Simon, 1902
 Cynapes Simon, 1900
 Cyrba Simon, 1876
 Cytaca Keyserling, 1882
 Damoetas Peckham & Peckham, 1886
 Darwinneon Cutler, 1971
 Dasycyptus Simon, 1902
 Davidina Brignoli, 1985
 Deloripa Simon, 1901
 Dendryphantes C.L.Koch, 1837
 Depreissia Lessert, 1942
 Descanso Peckham & Peckham, 1892
 Dexippus Thorell, 1891
 Diagondas Simon, 1902
 Dinattus Bryant, 1943
 Diolenius Thorell, 1869
 Diplocanthopoda Abraham, 1925
 Dolichoneon Caporiacco, 1935
 Donaldius Chickering, 1946
 Donoessus Simon, 1902
 Dryphias Simon, 1901
 Echeclus Thorell, 1890
 Echinussa Simon, 1901
 Efate Berland, 1938
 Emathis Simon, 1899
 Empanda Simon, 1903
 Encolpius Simon, 1900
 Encymachus Simon, 1902
 Enoplomischus Giltay, 1931
 Epeus Peckham & Peckham, 1885
 Epidelaxia Simon, 1902
 Epocilla Thorell, 1887
 Erasinus Simon, 1899
 Ergane L.Koch, 1881
 Erica Peckham & Peckham, 1892
 Eris C.L.Koch, 1846
 Euophrys C.L.Koch, 1834
 Eupoa Žabka, 1985
 Euryattus Thorell, 1881
 Eustiromastix Simon, 1902
 Evarcha Simon, 1902
 Featheroides Peng, Yin, Xie & Kim, 1994
 Festucula Simon, 1901
 Flacillula Strand, 1932
 Fluda Peckham & Peckham, 1892
 Freya C.L.Koch, 1850
 Frigga C.L.Koch, 1850
 Fritzia O.P.-Cambridge, 1879
 Fuentes Peckham & Peckham, 1894
 Furculattus Balogh, 1980
 Gangus Simon, 1902
 Gedeia Simon, 1902
 Gelotia Thorell, 1890
 Giuiria Strand, 1906
 Goleba Wanless, 1980
 Goleta Peckham & Peckham, 1894
 Gorgasella Chickering, 1946
 Gypogyna Simon, 1900
 Habrocestum Simon, 1876
 Habronattus F.O.P.-Cambridge, 1901
 Haplopsecas Caporiacco, 1955
 Heratemita Strand, 1932
 Harmochirus Simon, 1885
 Hasarina Schenkel, 1963
 Hasarius Simon, 1871

- Helicius Żabka, 1981
 Heliophanillus Prószyński, 1989
 Heliophanus C.L.Koch, 1833
 Helpis Simon, 1901
 Helvetia Peckham & Peckham, 1894
 Hentzia Marx, 1883
 Hermotimus Simon, 1903
 Hispo Simon, 1886
 Hisukattus Galiano, 1987
 Holcolaetis Simon, 1886
 Holoplatys Simon, 1885
 Homalattus White, 1841
 Hurius Simon, 1901
 Hyciotia Strand, 1911
 Hyetussa Simon, 1902
 Hyllus C.L.Koch, 1846
 Hypaeus Simon, 1900
 Hypoblemum Peckham & Peckham, 1886
 Icius Simon, 1876
 Idastrandia Strand, 1929
 Ilargus Simon, 1901
 Iona Peckham & Peckham, 1886
 Irura Peckham & Peckham, 1901
 Itata Peckham & Peckham, 1894
 Jacksonoides Wanless, 1988
 Jaluiticola Roewer, 1944
 Jollas Simon, 1901
 Jotus L.Koch, 1881
 Kinhia Żabka, 1985
 Klamathia Peckham & Peckham, 1903
 Lagnus L.Koch, 1879
 Langerra Żabka, 1985
 Langona Simon, 1901
 Lapsias Simon, 1900
 Laufeia Simon, 1888
 Lauharulla Keyserling, 1883
 Lechia Żabka, 1985
 Lepidemathis Simon, 1903
 Leptathamas Balogh, 1980
 Leptorchestes Thorell, 1870
 Letoia Simon, 1900
 Ligdus Thorell, 1895
 Ligonipes Karsch, 1878
 Ligurra Simon, 1903
 Longareus Simon, 1903
 Lophostica Simon, 1902
 Lurio Simon, 1901
 Luxuria Wesołowska, 1989
 Lycidas Karsch, 1878
 Lyssomanes Hentz, 1845
 Lystrocteisa Simon, 1884
 Mabellina Chickering, 1946
 Macopaeus Simon, 1900
 Maenola Simon, 1900
 Maeota Simon, 1901
 Maeotella Bryant, 1950
 Maevia C.L.Koch, 1846
 Mago O.P.-Cambridge, 1882
 Magyarus Żabka, 1985
 Maileus Peckham & Peckham, 1907
 Malloneta Simon, 1902
 Maltecora Simon, 1909
 Mantisatta Warburton, 1900
 Mantius Thorell, 1891
 Maratus Karsch, 1878
 Marchena Peckham & Peckham, 1909
 Marengo Peckham & Peckham, 1892
 Margaromma Keyserling, 1882
 Marma Simon, 1902
 Marpissa C.L.Koch, 1846
 Martella Peckham & Peckham, 1892
 Massagris Simon, 1900
 Mburuvicha Scioscia, 1993
 Meata Żabka, 1985
 Meleon Wanless, 1984
 Menemerus Simon, 1868
 Metacyrba F.O.P.-Cambridge, 1901
 Metaphidippus F.O.P.-Cambridge, 1901
 Mexcala Peckham & Peckham, 1902
 Micalula Strand, 1932
 Microhasarius Simon, 1902
 Mintonia Wanless, 1984
 Mirandia Badcock, 1932
 Mogrus Simon, 1882
 Monaga Chickering, 1946
 Mopiopia Simon, 1902
 Mopsolodes Żabka, 1991
 Mopsus Karsch, 1878
 Muziris Simon, 1901
 Myrmarachne MacLeay, 1839
 Nagaina Peckham & Peckham, 1896
 Nannenus Simon, 1902
 Natta Karsch, 1879
 Naubolus Simon, 1901
 Neaetha Simon, 1884
 Nebridia Simon, 1902
 Neobrettus Wanless, 1984
 Neon Simon, 1876
 Neonella Gertsch, 1936
 Nicylla Thorell, 1890
 Noegus Simon, 1900
 Nungia Żabka, 1985
 Nycerella Galiano, 1982
 Ocnotelus Simon, 1902
 Ocrisiona Simon, 1901
 Ogdenia Peckham & Peckham, 1908
 Omoedus Thorell, 1881
 Oningis Simon, 1901
 Onomastus Simon, 1900
 Opisthoncana Strand, 1913
 Opisthuncus L.Koch, 1880
 Orsima Simon, 1901
 Orthrus Simon, 1900
 Orvilleus Chickering, 1946
 Osericta Simon, 1901
 Pachyballus Simon, 1900
 Pachynomastus Caporiacco, 1947
 Pachypoessa Simon, 1902
 Padilla Peckham & Peckham, 1894
 Palpelius Simon, 1903
 Panachraesta Simon, 1900
 Pancorius Simon, 1902
 Pandisus Simon, 1900
 Panysinus Simon, 1901
 Paradamoetas Peckham & Peckham, 1885
 Paradecta Bryant, 1950
 Paradescanso Vellard, 1924
 Parafluda Chickering, 1946
 Paraharmochirus Szombathy, 1915
 Paraheliophanus Clark & Benoit, 1977

- Parajotus Peckham & Peckham, 1903
 Paraneaetha Denis, 1947
 Paraplexippus Franganillo, 1930
 Parasaitis Bryant, 1950
 Parathiodina Bryant, 1943
 Parkella Chickering, 1946
 Parnaenus Peckham & Peckham, 1896
 Peckhamia Simon, 1901
 Pelegrina Franganillo, 1930
 Pellenes Simon, 1876
 Pellolessertia Strand, 1929
 Penionomus Simon, 1903
 Pensacola Peckham & Peckham, 1885
 Pensacolops Bauab, 1983
 Peplometus Simon, 1900
 Phaeacius Simon, 1900
 Phantias F.O.P.-Cambridge, 1901
 Pharacocerus Simon, 1902
 Phaulostylus Simon, 1902
 Phausina Simon, 1901
 Phiale C.L.Koch, 1846
 Phidippus C.L.Koch, 1846
 Philaeus Thorell, 1869
 Phintella Strand, 1906
 Phlegra Simon, 1876
 Phyaces Simon, 1902
 Pilia Simon, 1902
 Piranthus Thorell, 1895
 Platycryptus Hill, 1979
 Platypsecas Caporiacco, 1955
 Plexippoides Prószyński, 1984
 Plexippus C.L.Koch, 1846
 Pochyta Simon, 1901
 Poecilorchestes Simon, 1901
 Poessa Simon, 1902
 Polemus Simon, 1902
 Porius Thorell, 1892
 Portia Karsch, 1878
 Poultonella Peckham & Peckham, 1909
 Pristobaeus Simon, 1902
 Proctonemesia Bauab & Soares, 1978
 Prostheclina Keyserling, 1882
 Proszynskiana Logunov, 1996
 Psecas C.L.Koch, 1850
 Pselcis Simon, 1903
 Pseudamphidraus Caporiacco, 1947
 Pseudamycus Simon, 1885
 Pseudattulus Caporiacco, 1947
 Pseudemathis Simon, 1902
 Pseudicius Simon, 1885
 Pseudocorythalia Caporiacco, 1938
 Pseudofluda Mello-Leitão, 1928
 Pseudoheliophanus Schenkel, 1963
 Pseudomaevia Rainbow, 1920
 Pseudopartona Caporiacco, 1954
 Pseudoplexippus Caporiacco, 1947
 Pseudosynagelides Żabka, 1991
 Ptocasius Simon, 1885
 Pystira Simon, 1901
 Quekettia Peckham & Peckham, 1902
 Rarahu Berland, 1929
 Rhene Thorell, 1869
 Rhetenor Simon, 1902
 Rhombonotus L.Koch, 1879
 Rhyphelia Simon, 1902
 Roeweriella Kratochvíl, 1932
 Rogmocrypta Simon, 1900
 Romitia Caporiacco, 1947
 Rudra Peckham & Peckham, 1885
 Sadies Wanless, 1984
 Saitidops Simon, 1901
 Saitis Simon, 1876
 Saitissus Roewer, 1938
 Salpesia Simon, 1901
 Salticus Latreille, 1804
 Sandalodes Keyserling, 1883
 Saraina Wanless & Clark, 1975
 Sarinda Peckham & Peckham, 1892
 Sarindoides Mello-Leitão, 1922
 Sassacus Peckham & Peckham, 1895
 Scartes Menge, 1879
 Schenkelia Lessert, 1927
 Scopocira Simon, 1900
 Scoturius Simon, 1901
 Sebastira Simon, 1901
 Selimus Peckham & Peckham, 1901
 Semiopyla Simon, 1901
 Semnolius Simon, 1902
 Semora Peckham & Peckham, 1892
 Semorina Simon, 1901
 Servaea Simon, 1888
 Sidusa Peckham & Peckham, 1895
 Sigytes Simon, 1902
 Siler Simon, 1888
 Siloca Simon, 1902
 Simaetha Thorell, 1881
 Simaethula Simon, 1902
 Simonurius Galiano, 1988
 Simprulla Simon, 1901
 Sitticus Simon, 1901
 Sobasina Simon, 1897
 Sondra Wanless, 1988
 Sonoita Peckham & Peckham, 1903
 Spartaeus Thorell, 1891
 Spilargis Simon, 1902
 Stagetillus Simon, 1885
 Stenaelurillus Simon, 1885
 Stenodeza Simon, 1900
 Stergusa Simon, 1888
 Stertinius Simon, 1890
 Stichius Thorell, 1890
 Stoidis Simon, 1901
 Stridulattus Petrunkevitch, 1926
 Sumampattus Galiano, 1983
 Synageles Simon, 1876
 Synagelides Strand, 1906
 Synemosyna Hentz, 1846
 Tacuna Peckham & Peckham, 1901
 Taivala Peckham & Peckham, 1907
 Talavera Peckham & Peckham, 1909
 Tamigalesus Żabka, 1988
 Tanna Berland, 1938
 Tanybelus Simon, 1902
 Tara Peckham & Peckham, 1885
 Taraxella Wanless, 1984
 Tariona Simon, 1902
 Tarne Simon, 1885
 Tarodes Pocock, 1899
 Tasa Wesołowska, 1981
 Tatari Berland, 1938
 Taula Wanless, 1988

Telamonia Thorell, 1887
 Thammaca Simon, 1901
 Thianella Strand, 1907
 Thiania C.L.Koch, 1846
 Thianitara Simon, 1903
 Thiodina Simon, 1900
 Thiratoscirtus Simon, 1886
 Thorelliola Strand, 1942
 Thyene Simon, 1885
 Thyenillus Simon, 1909
 Thyenula Simon, 1902
 Titanattus Peckham & Peckham, 1885
 Toloella Chickering, 1946
 Tomocyra Simon, 1900
 Trite Simon, 1885
 Tularosa Peckham & Peckham, 1902
 Tullgrenella Mello-Leitão, 1941
 Tulpus Peckham & Peckham, 1896
 Tusitala Peckham & Peckham, 1902
 Tutelina Simon, 1901
 Tylogonus Simon, 1902
 Uluella Chickering, 1946
 Urobullus Simon, 1902
 Uxuma Simon, 1902
 Vailima Peckham & Peckham, 1907
 Vatovia Caporiacco, 1940
 Veissella Wanless, 1984
 Viciria Thorell, 1877
 Vinnius Simon, 1902
 Viroqua Peckham & Peckham, 1901
 Wallaba Mello-Leitão, 1940
 Wedoquella Galiano, 1984
 Yaginumaella Prószyński, 1979
 Yaginumanis Wanless, 1984
 Yepoella Galiano, 1970
 Yllenus Simon, 1868
 Zenodorus Peckham & Peckham, 1886
 Zeuxippus Thorell, 1891
 Zuniga Peckham & Peckham, 1892
 Zygoballus Peckham & Peckham, 1885

Family **Scytodidae** 1 genus

Scytodes Latreille, 1804

Family **Segestriidae** 4 genera

Ariadna Savigny, 1825
 Gippsicola Hogg, 1900
 Periegops Simon, 1893
 Segestria Latreille, 1804

Family **Selenopidae** 4 genera

Anyphops Benoit, 1968
 Hovops Benoit, 1968
 Orops Benoit, 1968
 Selenops Latreille, 1819

Family **Senoculidae** 1 genus

Senoculus Taczanowski, 1872

Family **Sicariidae** 2 genera

Loxosceles Heineken & Lowe, 1835
 Sicarius Walckenaer, 1847

Family **Stenochilidae** 2 genera

Colopea Simon, 1893
 Stenochilus O.P.-Cambridge, 1870

Family **Stiphidiidae** 8 genera

Baiami Lehtinen, 1967
 Cambridgea L.Koch, 1872
 Corasoides Butler, 1929
 Ischalea L.Koch, 1872
 Nanocambridgea Forster & Wilton, 1973
 Procambridgea Forster & Wilton, 1973
 Stiphidion Simon, 1902
 Tjurunga Lehtinen, 1967

Family **Symphytognathidae** 6 genera

Anapistula Gertsch, 1941
 Anapogonia Simon, 1905
 Curimagua Forster & Platnick, 1977
 Globignatha Balogh & Loksa, 1968
 Patu Marples, 1951
 Symphytognatha Hickman, 1931

Family **Synotaxidae** 12 genera

Chileotaxus Platnick, 1990
 Mangua Forster, 1990
 Meringa Forster, 1990
 Nomaua Forster, 1990
 Pahora Forster, 1990
 Pahoroides Forster, 1990
 Paratupua Platnick, 1990
 Physoglenes Simon, 1904
 Runga Forster, 1990
 Synotaxus Simon, 1894
 Tupua Platnick, 1990
 Wairua Forster, 1990

Family **Telemidae** 6 genera

Apneumonella Fage, 1921
 Cangoderces Harington, 1951
 Jocquella Baert, 1980
 Seychellia Saaristo, 1978
 Telema Simon, 1882
 Usofila Keyserling, 1891

Family **Tengellidae** 5 genera

Calamistrula Dahl, 1901
 Lauricius Simon, 1888
 Liocranoides Keyserling, 1881
 Tengella Dahl, 1901
 Zorocrates Simon, 1888

Family **Tetrablemmidae** 29 genera

Ablemma Roewer, 1963

Afroblemma Lehtinen, 1981
 Anansia Lehtinen, 1981
 Borneomma Deeelman-Reinhold, 1980
 Brignoliella Shear, 1978
 Caraimatta Lehtinen, 1981
 Chavia Lehtinen, 1981
 Choiolemma Bourne, 1980
 Cuangoblemma Brignoli, 1974
 Fallablemma Shear, 1978
 Gunasekara Lehtinen, 1981
 Hexablemma Berland, 1920
 Indicoblemma Bourne, 1980
 Lamania Lehtinen, 1981
 Maijana Lehtinen, 1981
 Mariblemma Lehtinen, 1981
 Matta Crosby, 1934
 Micromatta Lehtinen, 1981
 Monoblemma Gertsch, 1941
 Paculla Simon, 1887
 Pahanga Shear, 1979
 Perania Thorell, 1893
 Rhinoblemma Lehtinen, 1981
 Sabahya Deeelman-Reinhold, 1980
 Shearella Lehtinen, 1981
 Singalanga Lehtinen, 1981
 Singaporemma Shear, 1978
 Sulaimania Lehtinen, 1981
 Tetrablemma O.P.-Cambridge, 1873

Family **Tetragnathidae** 51 genera

Agriognatha O.P.-Cambridge, 1896
 Antillognatha Bryant, 1945
 Atelidea Simon, 1894
 Atimiosa Simon, 1894
 Azilia Keyserling, 1882
 Chrysometa Simon, 1894
 Clitaetra Simon, 1889
 Cyrtognatha Keyserling, 1882
 Deliochus Simon, 1894
 Diphya Nicolet, 1849
 Dolichognatha O.P.-Cambridge, 1869
 Doryonychus Simon, 1900
 Dyschiriognatha Simon, 1893
 Eryciniolia Strand, 1912
 Glenognatha Simon, 1887
 Herennia Thorell, 1877
 Hispanognatha Bryant, 1945
 Homalomete Simon, 1897
 Leucauge White, 1841
 Mecynomete Simon, 1894
 Menosira Chikuni, 1955
 Mesida Kulczyński, 1911
 Meta C.L.Koch, 1836
 Metabus O.P.-Cambridge, 1899
 Metargyra F.O.P.-Cambridge, 1903
 Metellina Chamberlin & Ivie, 1941
 Metimorpha Strand, 1906
 Metleucauge Levi, 1980
 Mimicosa Petrunkevitch, 1925
 Mitoscelis Thorell, 1890
 Nanomete Simon, 1908
 Napomete Benoit, 1977
 Neoprolochus Reimoser, 1927
 Nephila Leach, 1815
 Nephilengys L.Koch, 1871

Opadometa Archer, 1951
 Orsinome Thorell, 1890
 Pachygnatha Sundevall, 1823
 Parameta Simon, 1895
 Parazilia Lessert, 1938
 Perilla Thorell, 1895
 Phonognatha Simon, 1894
 Pickardinella Archer, 1951
 Prionolaema Simon, 1894
 Sancus Tullgren, 1910
 Schenkeliella Strand, 1934
 Singafrotypa Benoit, 1962
 Tetragnatha Latreille, 1804
 Timonoe Thorell, 1898
 Tylorida Simon, 1894
 Zygiella F.O.P.-Cambridge, 1902

Family **Theraphosidae** 88 genera

Acanthopelma F.O.P.-Cambridge, 1897
 Acanthoscurria Ausserer, 1871
 Annandaliella Hirst, 1909
 Anoploscelus Pocock, 1897
 Aphonopelma Pocock, 1901
 Avicularia Lamarck, 1818
 Batesiella Pocock, 1903
 Brachionopus Pocock, 1897
 Ceratogyrus Pocock, 1897
 Ceropelma Mello-Leitão, 1923
 Chaetopelma Ausserer, 1871
 Chilobrachys Karsch, 1891
 Citharacanthus Pocock, 1900
 Citharischius Pocock, 1900
 Citharognathus Pocock, 1895
 Coelogenium Purcell, 1902
 Coremiocnemis Simon, 1892
 Crassiscrus Reichling & West, 1996
 Cratorrhagus Simon, 1892
 Crypsidromus Ausserer, 1871
 Cyclosternum Ausserer, 1871
 Cyriocosmus Simon, 1903
 Cyriopagopus Simon, 1887
 Cyrtopholis Simon, 1892
 Drytopelma Simon, 1889
 Encyocrates Simon, 1892
 Ephebopus Simon, 1892
 Euathlus Ausserer, 1875
 Eucratoscelus Pocock, 1898
 Eumenophorus Pocock, 1897
 Eupalaestrus Pocock, 1901
 Euphrictus Hirst, 1908
 Grammostola Simon, 1892
 Hapalopus Ausserer, 1875
 Hapalotremus Simon, 1903
 Haploclastus Simon, 1892
 Haplopelma Simon, 1892
 Harpactira Ausserer, 1871
 Harpactirella Purcell, 1902
 Hemirrhagus Simon, 1903
 Heteroscodra Pocock, 1899
 Heterothele Karsch, 1879
 Holothele Karsch, 1879
 Homoeomma Ausserer, 1871
 Hysterochrates Simon, 1892
 Ischnocolus Ausserer, 1871
 Lampropelma Simon, 1892

Lasiadora C.L.Koch, 1850
Loxomphalia Simon, 1888
Loxoptygus Simon, 1903
Lyrognathus Pocock, 1895
Megaphobema Pocock, 1901
Monocentropus Pocock, 1897
Myostola Simon, 1903
Nesiergus Simon, 1903
Oligoxystre Vellard, 1924
Ornithoctonus Pocock, 1892
Orphnaecus Simon, 1892
Ozopactus Simon, 1889
Pachistopelma Pocock, 1901
Pachypelma Karsch, 1880
Pamphobeteus Pocock, 1901
Paraphysa Simon, 1892
Phlogiellus Pocock, 1897
Phoneyusa Karsch, 1884
Phormictopus Pocock, 1901
Phormingochilus Pocock, 1895
Phrixotrichus Simon, 1888
Plesiophrictus Pocock, 1899
Poecilotheria Simon, 1885
Psalmopoeus Pocock, 1898
Pseudotheraphosa Tinter, 1991
Pterinochilus Pocock, 1897
Schizopelma F.O.P.-Cambridge, 1897
Selenocosmia Ausserer, 1871
Selenogyrus Pocock, 1897
Selenotholus Hogg, 1902
Selenotypus Pocock, 1895
Sericopelma Ausserer, 1875
Spelopelma Gertsch, 1982
Sphaerobothria Karsch, 1879
Stichoplastus Simon, 1889
Stromatopelma Karsch, 1881
Tapinauchenius Ausserer, 1871
Theraphosa Thorell, 1870
Thrigmopoeus Pocock, 1899
Xenesthis Simon, 1891
Yamia Kishida, 1920

Family **Theridiidae** 62 genera

Achaearanea Strand, 1929
Adansonia Saville-Kent, 1897
Anatea Berland, 1927
Anelosimus Simon, 1891
Argyroides Simon, 1864
Audifia Keyserling, 1884
Cabello Levi, 1964
Carniella Thaler & Steinberger, 1988
Cephalobares O.P.-Cambridge, 1871
Cerocida Simon, 1894
Chrosiothes Simon, 1894
Chrysso O.P.-Cambridge, 1882
Coleosoma O.P.-Cambridge, 1882
Coscinida Simon, 1894
Craspedisia Simon, 1894
Crustulina Menge, 1868
Cyllognatha L.Koch, 1872
Dipoena Thorell, 1869
Dipoenata Wunderlich, 1988
Dipoenura Simon, 1908
Echinotheridion Levi, 1963
Enoplognatha Pavesi, 1880

Episinus Walckenaer, 1809
Euryopis Menge, 1868
Gmogala Keyserling, 1889
Gnophomytis Simon, 1894
Guaraniella Baert, 1984
Hadrotarsus Thorell, 1881
Helvibis Keyserling, 1884
Helvidia Thorell, 1890
Hetschkia Keyserling, 1886
Histagonia Simon, 1894
Icona Forster, 1955
Lasaeola Simon, 1881
Latrodectus Walckenaer, 1805
Marianana Georgescu, 1989
Molione Thorell, 1892
Nesticodes Archer, 1950
Paidiscura Archer, 1950
Paratheridula Levi, 1957
Pholcomma Thorell, 1869
Phoroncidia Westwood, 1835
Proboscidula Miller, 1970
Propostira Simon, 1894
Robertus O.P.-Cambridge, 1879
Rugathodes Archer, 1950
Spheropistha Yaginuma, 1957
Spintharus Hentz, 1850
Steatoda Sundevall, 1833
Stemmops O.P.-Cambridge, 1894
Styposis Simon, 1894
Tekellina Levi, 1957
Theonoe Simon, 1881
Theridion Walckenaer, 1805
Theridula Emerton, 1882
Thwaitesia O.P.-Cambridge, 1881
Thymoites Keyserling, 1884
Tidarren Chamberlin & Ivie, 1934
Tomoxena Simon, 1894
Wirada Keyserling, 1886
Yoroa Baert, 1984
Zercidium Benoit, 1977

Family **Theridiosomatidae** 12 genera

Allototua Bryant, 1945
Baalzebub Coddington, 1986
Chthonos Coddington, 1986
Epeirotypus O.P.-Cambridge, 1894
Epiileutes Coddington, 1986
Haliger Mello-Leitão, 1943
Naatlo Coddington, 1986
Ogulnius O.P.-Cambridge, 1882
Parogulnius Archer, 1953
Plato Coddington, 1986
Theridiosoma O.P.-Cambridge, 1879
Wendilgarda Keyserling, 1886

Family **Thomisidae** 160 genera

Acentroscelus Simon, 1886
Acracanthosoma Mello-Leitão, 1917
Alcimochthes Simon, 1885
Amyciaea Simon, 1885
Angaeus Thorell, 1881
Aphantochilus O.P.-Cambridge, 1870
Apyretina Strand, 1932
Ascurisoma Strand, 1928

- Avelis Simon, 1895
 Bassaniana Strand, 1928
 Bassanioides Pocock, 1903
 Boliscodes Simon, 1909
 Boliscus Thorell, 1891
 Bomis L.Koch, 1873
 Bonapruncinia Benoit, 1977
 Borboropactus Simon, 1884
 Browningella Mello-Leitão, 1948
 Bucranium O.P.-Cambridge, 1881
 Camaricus Thorell, 1887
 Carcinarachne Schmidt, 1956
 Cebrennius Simon, 1887
 Cerarachne Keyserling, 1880
 Cetratus Kulczyński, 1911
 Coenypha Simon, 1895
 Coriarachne Thorell, 1869
 Corynethrix L.Koch, 1876
 Cupa Strand, 1906
 Cymbacha L.Koch, 1874
 Cymbachina Bryant, 1933
 Cynathea Simon, 1895
 Cyriogonus Simon, 1886
 Deltoclitia Simon, 1877
 Demogenes Simon, 1895
 Diaea Thorell, 1870
 Dietopsa Strand, 1932
 Dimizonops Pocock, 1903
 Diplotychus Simon, 1903
 Domatha Simon, 1895
 Ebrechtella Dahl, 1907
 Emplesiogonus Simon, 1903
 Epicadinus Simon, 1895
 Epicadus Simon, 1895
 Epidius Thorell, 1877
 Erissoides Mello-Leitão, 1929
 Erissus Simon, 1895
 Felsina Simon, 1895
 Firmicus Simon, 1895
 Geraesta Simon, 1888
 Gnoerichia Dahl, 1907
 Haedanula Caporiacco, 1941
 Haplotmarus Simon, 1909
 Hedana L.Koch, 1874
 Herbessus Simon, 1903
 Heriaesynaema Caporiacco, 1939
 Heriaeus Simon, 1875
 Heterogriffus Platnick, 1976
 Hewittia Lessert, 1928
 Hexommulocymus Caporiacco, 1955
 Holopelus Simon, 1886
 Iphoctesis Simon, 1903
 Isala L.Koch, 1876
 Isaloides F.O.P.-Cambridge, 1900
 Lampertia Strand, 1907
 Latifrons Kulczyński, 1911
 Loxobates Thorell, 1877
 Loxoporetas Kulczyński, 1911
 Lycopus Thorell, 1895
 Lysiteles Simon, 1895
 Majellula Strand, 1932
 Martus Mello-Leitão, 1943
 Massuria Thorell, 1887
 Mecaphesa Simon, 1900
 Megapyge Caporiacco, 1947
 Misumena Latreille, 1804
 Misumenoides F.O.P.-Cambridge, 1900
 Misumenops F.O.P.-Cambridge, 1900
 Monaeses Thorell, 1869
 Musaeus Thorell, 1890
 Mystaria Simon, 1895
 Narcaeus Thorell, 1890
 Nyctimus Thorell, 1877
 Ocyllus Thorell, 1887
 Onocolus Simon, 1895
 Ostanes Simon, 1895
 Oxytate L.Koch, 1878
 Ozyptila Simon, 1864
 Pactates Simon, 1895
 Pagida Simon, 1895
 Parabomis Kulczyński, 1901
 Paramystaria Lessert, 1919
 Parasmodix Jézéquel, 1966
 Parastephanops F.O.P.-Cambridge, 1900
 Parastrophius Simon, 1903
 Parasynema F.O.P.-Cambridge, 1900
 Pasias Simon, 1895
 Pasiasula Roewer, 1942
 Peritraeus Simon, 1895
 Phaenopoma Simon, 1895
 Pharta Thorell, 1891
 Pherecydes O.P.-Cambridge, 1883
 Philodamia Thorell, 1894
 Philogaeus Simon, 1895
 Phireza Simon, 1886
 Phrynarachne Thorell, 1870
 Physoplatys Simon, 1895
 Pistius Simon, 1875
 Plancinus Simon, 1886
 Plastonimus Simon, 1903
 Platyarachne Keyserling, 1880
 Platypyresthesia Simon, 1903
 Platythomisus Doleschall, 1859
 Poecilothomisus Simon, 1895
 Porropis L.Koch, 1876
 Pothaeus Thorell, 1895
 Prepotelus Simon, 1897
 Pseudamyciaea Simon, 1905
 Pseudoporrhopis Simon, 1886
 Pycnaxis Simon, 1895
 Pyresthesia Butler, 1879
 Reinickella Dahl, 1907
 Rhaebobates Thorell, 1881
 Runcinia Simon, 1875
 Saccodomus Rainbow, 1900
 Scoticus Simon, 1895
 Sidymella Strand, 1942
 Simorcus Simon, 1895
 Smodicinus Simon, 1895
 Soelteria Dahl, 1907
 Stephanopsis O.P.-Cambridge, 1869
 Stephanopoides Keyserling, 1880
 Stiphropella Lawrence, 1952
 Stiphropus Gerstäcker, 1873
 Strigoplus Simon, 1885
 Strophius Keyserling, 1880
 Sylligma Simon, 1895
 Synaemops Mello-Leitão, 1929
 Synalus Simon, 1895
 Synema Simon, 1864
 Synstrophius Mello-Leitão, 1925

Tagulinus Simon, 1902
 Tagulis Simon, 1895
 Takachihoa Ono, 1985
 Talaus Simon, 1886
 Tarrocanus Simon, 1895
 Tharpyna L.Koch, 1874
 Tharrhalea L.Koch, 1875
 Thomisops Karsch, 1879
 Thomisus Walckenaer, 1905
 Titidiops Mello-Leitão, 1929
 Titidius Simon, 1895
 Tmarus Simon, 1875
 Tobias Simon, 1895
 Trichopagis Simon, 1886
 Ulocymus Simon, 1886
 Uraarachne Keyserling, 1880
 Wechselia Dahl, 1907
 Xysticus C.L.Koch, 1835
 Zametopias Thorell, 1890
 Zametopina Strand, 1909
 Zygometis Simon, 1901

Family **Titanoecidae** 5 genera

Anuvinda Lehtinen, 1967
 Goeldia Keyserling, 1891
 Nurscia Simon, 1874
 Pandava Lehtinen, 1967
 Titanoeca Thorell, 1869

Family **Trechaleidae** 11 genera

Demelodos Mello-Leitão, 1943
 Dossenus Simon, 1898
 Dyrines Simon, 1903
 Dyrinoides Badcock, 1932
 Enna O.P.-Cambridge, 1897
 Hesydrus Simon, 1898
 Paradossenus F.O.P.-Cambridge, 1903
 Sisenna Simon, 1898
 Syntrechalea F.O.P.-Cambridge, 1902
 Trechalea Thorell, 1870
 Xingusiella Mello-Leitão, 1940

Family **Trochanteriidae** 6 genera

Corimaethes Simon, 1908
 Doliomalus Simon, 1897
 Plator Simon, 1880
 Platyoides O.P.-Cambridge, 1890
 Rebilus Simon, 1880
 Trochanteria Karsch, 1878

Family **Uloboridae** 20 genera

Ariston O.P.-Cambridge, 1896
 Aştavakra Lehtinen, 1967
 Conitåber Opell, 1982
 Daramulunia Lehtinen, 1967
 Hyptiotes Walckenaer, 1837
 Lubinella Opell, 1984
 Miagrammopes O.P.-Cambridge, 1869
 Octonoba Opell, 1979
 Orinomana Strand, 1934
 Petrunkevitchia Mello-Leitão, 1915
 Philoponella Mello-Leitão, 1917

Polenecia Lehtinen, 1967
 Ponella Opell, 1979
 Purumitra Lehtinen, 1967
 Siratoba Opell, 1979
 Sybota Simon, 1892
 Tangaroa Lehtinen, 1967
 Uloborus Latreille, 1806
 Waitkera Opell, 1979
 Zosis Walckenaer, 1841

Family **Zodariidae** 50 genera

Acanthinozodium Denis, 1952
 Akyttara Jocqué, 1987
 Antillorena Jocqué, 1991
 Asceua Thorell, 1887
 Aschema Jocqué, 1991
 Asteron Jocqué, 1991
 Caesetius Simon, 1893
 Capheris Simon, 1893
 Chariobas Simon, 1893
 Cicynethus Simon, 1910
 Cybaeodamus Mello-Leitão, 1938
 Cydrela Thorell, 1873
 Cyrioctea Simon, 1889
 Diores Simon, 1893
 Dusmadiores Jocqué, 1987
 Forsterella Jocqué, 1991
 Habronestes L.Koch, 1872
 Heradida Simon, 1893
 Hermippus Simon, 1893
 Hetaerica Rainbow, 1916
 Ishania Chamberlin, 1925
 Lachesana Strand, 1932
 Leprolochus Simon, 1892
 Lutica Marx, 1891
 Macedoniola Strand, 1932
 Madrela Jocqué, 1991
 Mallinella Strand, 1906
 Mallinus Simon, 1893
 Mastidiores Jocqué, 1987
 Microdiores Jocqué, 1987
 Nanahua Badcock, 1932
 Neostorena Rainbow, 1914
 Nostera Jocqué, 1991
 Palaestina O.P.-Cambridge, 1872
 Palfuria Simon, 1910
 Pax Levy, 1990
 Platnickia Jocqué, 1991
 Psammoduon Jocqué, 1991
 Psammorygma Jocqué, 1991
 Ranops Jocqué, 1991
 Selamia Simon, 1873
 Storena Walckenaer, 1805
 Storenomorpha Simon, 1884
 Storsia Jocqué, 1991
 Suffasia Jocqué, 1991
 Tenedos O.P.-Cambridge, 1897
 Thaumastochilus Simon, 1897
 Tristichops Taczanowski, 1874
 Trygetus Simon, 1882
 Zodarion Walckenaer, 1847

Family **Zoridae** 12 genera

Argoctenus L.Koch, 1878

Diallomus Simon, 1897
 Hestimodema Simon, 1909
 Neoctenus Simon, 1897
 Odo Keyserling, 1887
 Odomasta Simon, 1909
 Simonus Ritsema, 1881
 Thasyraea L.Koch, 1878
 Tunabo Chamberlin, 1916

Voraptus Simon, 1897
 Zora C.L.Koch, 1847
 Zoroides Berland, 1924

Family **Zoropsidae** 2 genera

Takeoa Lehtinen 1967
 Zoropsis Simon, 1878

Families Abbreviations

AC Actinopodidae	DG Diguettidae	MC Mecicobothriidae	SA Salticidae
AD Antrodiaetidae	DI Dictynidae	MF Micropholcommatidae	SC Scytodidae
AG Agelenidae	DP Dipluridae	MG Migidae	SF Stiphidiidae
AI Amphinectidae	DR Drymusidae	MK Mecysmaucheniidae	SG Segestriidae
AK Archaeidae	DS Desidae	ML Malkaridae	SI Sicariidae
AM Amaurobiidae	DY Dysderidae	MM Mimetidae	SL Selenopidae
AN Anyphaenidae	ER Eresidae	MS Microstigmatidae	SN Senoculidae
AP Anapidae	FI Filistatidae	MT Miturgidae	ST Stenochilidae
AR Araneidae	GL Gallieniellidae	MY Mysmenidae	SX Synotaxidae
AT Atypidae	GN Gnaphosidae	NE Neolanidae	SY Symphytognathidae
AU Austrochilidae	GR Gradungulidae	NI Nicodamidae	TB Tetrablemmidae
AX Ammoxenidae	HA Halidae	NM Nemesiidae	TC Trechaleidae
AY Argyronetidae	HH Hahniidae	NS Nesticidae	TD Theridiidae
BA Barychelidae	HN Homalonychidae	OC Ochyroceratidae	TE Telemidae
BR Bradystichidae	HO Holarchaeidae	OE Oecobiidae	TG Tetragnathidae
CA Caponiidae	HP Hypochilidae	OO Oonopidae	TH Thomisidae
CB Cybaeidae	HR Hersiliidae	OR Orsolobidae	TN Tengellidae
CC Cycloctenidae	HT Heteropodidae	OX Oxyopidae	TP Theraphosidae
CI Cithaeronidae	HU Huttoniidae	PA Paratropididae	TR Trochanteriidae
CL Clubionidae	HX Hexathelidae	PC Pholcidae	TS Theridiosomatidae
CO Corinnidae	ID Idiopidae	PD Philodromidae	TT Titanoecidae
CR Cryptothelidae	LA Lamponidae	PI Pisauridae	UL Uloboridae
CT Ctenidae	LC Liocranidae	PO Prodidomidae	ZD Zodariidae
CU Cyrtaucheniidae	LE Leptonetidae	PP Palpimanidae	ZO Zoridae
CY Cyatholipidae	LI Linyphiidae	PR Pararchaeidae	ZP Zoropsidae
CZ Ctenizidae	LP Liphistiidae	PS Psechridae	
DE Deinopidae	LY Lycosidae	PT Plectreuridae	

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Gypogyna SA	Hermachura NM	Homalopolys AR	Ilargus SA
Habreuresis LI	Hermippus ZD	Homoeomma TP	Ilipula PI
Habrocestum SA	Hermotimus SA	Homoeothele GN	Ilisoa CY
Habronattus SA	Herpyllus GN	Homostola CU	Incasoctenus CT
Habronestes ZD	Hersilia HR	Honunius PO	Incestophantes LI
Hackmania DI	Hersiliola HR	Hoplalathys DI	Indicoblema TB
Hadites AG	Hesperocosa LY	Hoplopholcus PC	Inola PI
Hadronyche HX	Hesperocranum LC	Horcotes LI	Inthaeron CI
Hadrotarsus TD	Hestimodema ZO	Horioctenoides CT	Intihuatana HH
Haedanula TH	Hesydrimorpha PI	Hostus OX	Intruda GN
Hahnia HH	Hesydrus TC	Hovops SL	Iona SA
Hala HA	Hetaerica ZD	Huangyuania AG	Iphoctesis TH
Haliger TS	Heterochemmis LC	Huara AI	Irura SA
Halorates LI	Heterocybaeus CB	Hubertella LI	Isala TH
Hamataliwa OX	Heterognatha AR	Huka AG	Isaloides TH
Hanea CY	Heterogriffus TH	Hulua DS	Ischalea SF
Hapalopus TP	Heterolinyphia LI	Humua CO	Ischnocolus TP
Hapalothele DP	Heteromigas MG	Hurius SA	Ischnothele DP
Hapalotremus TP	Heteroonops OO	Huttonia HU	Ischnothyreus OO
Haplinis LI	Heteropoda HT	Hyaenosa LY	Isela MY
Haploclastus TP	Heteroscodra TP	Hybauchenidium LI	Ishania ZD
Haplodrassus GN	Heterothele TP	Hybocoptus LI	Isicabu CY
Haplomaro LI	Heterotrichoncus LI	Hybosida PP	Isigonia AN
Haplopelma TP	Hetschkia TD	Hyciotea SA	Islandiana LI
Haplopsecas SA	Heurodes AR	Hyetussa SA	Isoctenus CT
Haplotmarus TH	Hewittia TH	Hygrocrates DY	Isoghona LY
Hapona DS	Hexablemma TB	Hygrolycosa LY	Isopeda HT
Haptisus AN	Hexathele HX	Hygropoda PI	Isopedella HT

- Isoxya AR
 Itapua MY
 Itata SA
 Itatiaya CT
 Itatsina LC
 Itytis LI
 Iviella DI
 Ivielum LI
 Ixamatus NM

 Jacaena GN
 Jacksonella LI
 Jacksonoides SA
 Jalapyphantes LI
 Jaluiticola SA
 Janetschekia LI
 Janusia CT
 Jocquella TE
 Johorea LI
 Jollas SA
 Josa AN
 Jotus SA

 Kaestneria LI
 Kaira AR
 Kaitawa GN
 Kaiya GR
 Kapanga HH
 Kapitia OO
 Katumbea PO
 Keilira HT
 Kekenboschiella MY
 Kiama CU
 Kibramoa PT
 Kidugua AG
 Kijabe OO
 Kikimora LI
 Kilifina MY
 Kilima AR
 Kinhia SA
 Kirmaka GN
 Kishidaia GN
 Klamathia SA
 Knischatiria LI
 Koinothrix LI
 Kolymocyba LI
 Kratochvilia MM
 Kratochviliella LI
 Kuala LI
 Kukulcania FI
 Kulalania AM
 Kwonkan NM

 Labicymbium LI
 Labulla LI
 Labullinyphia LI
 Labullula LI
 Lachesana ZD
 Ladissa GN
 Laestrygones DS
 Laetesia LI
 Lagnus SA
 Lamaika AM
 Lamania TB
 Lamina DS
 Laminacauda LI

 Lampertia TH
 Lampona LA
 Lampropelma TP
 Langerra SA
 Langona SA
 Laperousea LI
 Lapsias SA
 Larinia AR
 Lariniaria AR
 Larinioides AR
 Lasaeola TD
 Lascona CL
 Lasiargus LI
 Lasiodora TP
 Lathrothele DP
 Lathyarca DS
 Lathys DI
 Latifrons TH
 Latithorax LI
 Latonigena GN
 Latouchia CZ
 Latroectus TD
 Laudetia LC
 Laufeia SA
 Lauharulla SA
 Lauricius TN
 Lausus CO
 Lechia SA
 Legendrena GL
 Lepidemathis SA
 Leprolochus ZD
 Leptathamas SA
 Lepthercus NM
 Lepthyphantes LI
 Leptoctenus CT
 Leptodrassus GN
 Leptoneta LE
 Leptonetela LE
 Leptopholcus PC
 Leptorchestes SA
 Leptorhoptrum LI
 Leptothrix LI
 Lessertia LI
 Lessertina CO
 Lessertinella LI
 Letoia SA
 Leucauge TG
 Leucorchestris HT
 Leviola MY
 Ligdus SA
 Liger LI
 Ligonipes SA
 Ligurra SA
 Limoneta LI
 Linothele DP
 Linyphantes LI
 Linyphia LI
 Linyphioides LI
 Liocranoides TN
 Liocranum LC
 Lionnetta OO
 Liparochrysis LC
 Liparotoma AN
 Liphistius LP
 Lipocrea AR
 Litoporus PC

 Litopyllus GN
 Livius AM
 Lizarba HH
 Locketidium LI
 Loculla LY
 Lomaita LI
 Longarenus SA
 Lophomma LI
 Lophostica SA
 Louisfagea LI
 Loxobates TH
 Loxomphalia TP
 Loxoporetes TH
 Loxoptygus TP
 Loxosceles SI
 Lubinella UL
 Lucetia OO
 Lucrinus LI
 Lundacera OC
 Lurio SA
 Lutica ZD
 Luxuria SA
 Lycidas SA
 Lycopus TH
 Lycorma LY
 Lycosa LY
 Lycosella LY
 Lycosoides AG
 Lycosula LY
 Lygarina LI
 Lygromma PO
 Lygrommatoides PO
 Lynxosa LY
 Lyrognathus TP
 Lysania LY
 Lysiteles TH
 Lyssomanes SA
 Lystrocteisa SA

 Mabellina SA
 Macedoniella LC
 Macedoniola ZD
 Macerio CL
 Machadocara LI
 Machadonia MT
 Macopaeus SA
 Macracantha AR
 Macrargus LI
 Macrinus HT
 Macrobonus AM
 Macrogradungula GR
 Macrophytes AN
 Macrothele HX
 Madacantha AR
 Madreia ZD
 Maenola SA
 Maeota SA
 Maeotella SA
 Maevia SA
 Mago SA
 Magyarus SA
 Mahembea AR
 Mahura AG
 Majjana TB
 Maileus SA
 Maimuna AG

 Majellula TH
 Makora AI
 Malaika AM
 Malalistata FI
 Malimbosa LY
 Malkara ML
 Malkinella LI
 Malkinia LI
 Mallecolobus OR
 Mallecomigas MG
 Mallinella ZD
 Mallinus ZD
 Malloneta SA
 Mallos DI
 Maloides AM
 Maltecora SA
 Malthonica AG
 Mamoea AI
 Manawa DS
 Mandanapis AP
 Mandaneta CO
 Mangareia DS
 Mangora AR
 Mangua SX
 Maniana GN
 Maniho AI
 Manjala AM
 Mantisatta SA
 Mantius SA
 Maoriata OR
 Maorineta LI
 Maratus SA
 Marchena SA
 Mardonica LC
 Marengo SA
 Margaromma SA
 Margonia LY
 Marianana TD
 Mariblemma TB
 Marilynia DI
 Marma SA
 Maro LI
 Marpissa SA
 Marplesia AI
 Marsupopaea OO
 Martella SA
 Martensinus LI
 Martus TH
 Mashimo DI
 Masikia LI
 Masirana LE
 Maso LI
 Masoncus LI
 Masonetta LI
 Massagris SA
 Massuria TH
 Masteria DP
 Mastidiores ZD
 Mastigusa DI
 Mastophora AR
 Matachia DS
 Matidia CL
 Matilda CY
 Matta TB
 Matua GN
 Matundua AM

Maxanapis AP	Metamynoglenes LI	Misgolas ID	Namaquarachne AM
Maymena MY	Metanapis AP	Missulena AC	Namea NM
Maypacijs PI	Metapanamomops LI	Mistaria AG	Namirea DP
Mazax CO	Metaphidippus SA	Misumena TH	Nanahua ZD
Mburuvicha SA	Metargyra TG	Misumenoides TH	Nanavia LI
Meata SA	Metazygia AR	Misumenops TH	Nanduti AR
Mecaphesa TH	Metellina TG	Mitoscelis TG	Nannenus SA
Mecicobothrium MC	Metepira AR	Mitrager LI	Nanocambridgea SF
Mecolaesthus PC	Methesis CO	Miturga MT	Nanometa TG
Mecopisthes LI	Metimorpha TG	Mizaga DI	Napometa TG
Mecynargoides LI	Metleucauge TG	Modisimus PC	Narcaeus TH
Mecynargus LI	Metopobactrus LI	Moebelia LI	Nasoonia LI
Mecynidis LI	Mevianops AG	Moggridgea MG	Natta SA
Mecynogea AR	Mexcala SA	Mogrus SA	Naubolus SA
Mecynometa TG	Mexentypesa NM	Molinaranea AR	Nauhea GN
Mecysmauchenoides MK	Mexitlia DI	Molione TD	Neaetha SA
Mecysmauchenius MK	Mezenina AN	Molitorosa LY	Nebidia SA
Mediothele HX	Miagrammopes UL	Molycrisa PO	Nematogmus LI
Medmassa CO	Micalula SA	Monaeses TH	Nemesia NM
Meedo CL	Micaria GN	Monaga SA	Nemesoides CU
Megadictyna NI	Micrargus LI	Monapia AN	Nemoctenus CT
Megadolomedes PI	Micrathena AR	Monoblemma TB	Nemoscolus AR
Megafroneta LI	Micrepeira AR	Monocentropus TP	Nemosinga AR
Megaheura MC	Microbathyphantes LI	Monocephalus LI	Nemospiza AR
Megaloremnius HT	Microtema LI	Monocerellus LI	Neilinium LI
Megalostrata CO	Microctenonyx LI	Monodontium BA	Nentwigia LI
Megamyrmaekion GN	Microcyba LI	Montanapis AP	Neoanagraphis CL
Megaphobema TP	Microdiores ZD	Montebello LC	Neoantistea HH
Megapyge TH	Microdipoena MY	Montescueia CT	Neoarchemorus AR
Megaraneus AR	Microdrassus GN	Montilaira LI	Neoaviola HH
Megarcota LY	Microfilistata FI	Mopiopia SA	Neobrettus SA
Megullia OX	Microhasarius SA	Mopsolodes SA	Neocautinella LI
Meioneta LI	Microhexura DP	Mopsus SA	Neocryphoea HH
Melaenosia MM	Microlinyphia LI	Moreiraxena LI	Neocteniza ID
Meleon SA	Micromaso LI	Morenilia GN	Neoctenus ZO
Melicymnis GN	Micromatta TB	Moreno GN	Neodiplothele NM
Melloicosa LY	Micromerys PC	Muizenbergia HH	Neoburnella LI
Melloina PA	Micromesomma MG	Muritaia AM	Neogea AR
Melocosa LY	Micrommata HT	Murricia HR	Neohahnia HH
Melpomene AG	Micromygale MS	Musaeus TH	Neolana NE
Melychiopharis AR	Microneta LI	Mustelica LY	Neoleptoneta LE
Menemerus SA	Micropholcomma MF	Muziris SA	Neomaso LI
Menneus DE	Micropholcus PC	Myandra PO	Neomyro DS
Menosira TG	Microplanus LI	Mycula LI	Neon SA
Merenius CO	Micropoltys AR	Mygaloides NM	Neonella SA
Meringa SX	Microrchestris HT	Myostola TP	Neonesiotes LI
Merizocera OC	Microsa GN	Myrmarachne SA	Neoporteria AM
Mermessus LI	Microsphalma LI	Myrmecium CO	Neoprolodus TG
Merredinia NM	Microstigmata MS	Myrmecomelix LI	Neoramia AG
Mesarchaea MK	Micryphantes LI	Myrmecoscapheila OO	Neorepukia AG
Mesasigone LI	Micythus GN	Myrmecotypus CO	Neororea AI
Mesida TG	Migas MG	Myrmekiaphila CU	Neoscona AR
Mesilla AN	Millidgea LI	Myrmidonella PC	Neosparassus HT
Mesiotelus LC	Milonia AR	Myrmopopaea OO	Neostorena ZD
Meskia GN	Mimetus MM	Myro DS	Neotegenaria AG
Mesobria LC	Mimicosa TG	Mysmena MY	Neoxyphinus OO
Mesostalita DY	Minanapis AP	Mysmenella MY	Neozimiris PO
Messapus LC	Minicia LI	Mysmenopsis MY	Nephila TG
Meta TG	Ministigmata MS	Mystaria TH	Nephilengys TG
Metabus TG	Minosia GN	Mystes PC	Nephrochirus OO
Metacleonemius PD	Minosiella GN	Mythoplastoides LI	Neriene LI
Metacyrba SA	Minotauria DY		Nesiergus TP
Metafroneta LI	Mintonia SA	Naatlo TS	Nesioneta LI
Metagonia PC	Minyriolus LI	Naevius DS	Nesticella NS
Metaleptyphantes LI	Mioxena LI	Nagaina SA	Nesticodes TD
Metaltella AM	Mirandia SA	Namandia DS	Nesticus NS

Neuquenina AM	Olbophthalmus AN	Pachyballus SA	Paralarinia AR
Nicodamus NI	Olbus CL	Pachydelphus LI	Paraleptoneta LE
Nicoletina CL	Olgania MF	Pachygnatha TG	Paraletes LI
Nicylla SA	Oligoctenus CT	Pachyonomastus SA	Paralobus OR
Nigma DI	Oligoxystre TP	Pachypelma TP	Paramamoea AI
Nilus PI	Olios HT	Pachypoessa SA	Paramatachia DS
Ninetis PC	Olorunia AG	Pacificana MT	Parameioneta LI
Nisueta HT	Oltacloea PO	Pacifiphantes LI	Parameta TG
Nodocion GN	Ommatauxesis DS	Pactates TH	Paramicromerys PC
Noegus SA	Omoedus SA	Paculla TB	Paramigas MG
Nomaua SX	Oningis SA	Padilla SA	Paramyro AG
Nomisia GN	Onocolus TH	Paenula HT	Paramystaria TH
Nonianus HT	Onomastus SA	Pagida TH	Paranapis AP
Nops CA	Onychembolus LI	Pagiopalus PD	Paranasoona LI
Nopsides CA	Oonopinus OO	Pahanga TB	Paraneaetha SA
Nortanapis AP	Oonopoides OO	Pahora SX	Paraphysa TP
Nostera ZD	Oonops OO	Pahoroides SX	Paraplectana AR
Notholephyphantes LI	Opadometa TG	Paidiscura TD	Paraplectanoides AR
Nothophantes LI	Oparara AI	Pakeha AM	Paraplexippus SA
Nothroctenus CT	Ophrynia LI	Palaeohyphantes LI	Parapostenus MT
Notiodrassus GN	Opisthancana SA	Palaestina ZD	Parapua MF
Notiohyphantes LI	Opisthonus SA	Palaetyra LC	Pararaneus AR
Notiomaso LI	Opopaea OO	Palfuria ZD	Pararchaea PR
Notioscopus LI	Oramia AG	Palicanus MT	Parasaitis SA
Notiothauma LI	Oramiella AG	Palpelius SA	Parasisis LI
Notomatachia DS	Orchestina OO	Palpimanus PP	Parasmodix TH
Novafroneta LI	Orchestrella HT	Palystella HT	Parastalita DY
Novafrontina LI	Ordgarius AR	Palystes HT	Parastephanops TH
Novakiella AR	Oreocyba LI	Pamphobeteus TP	Parastrophius TH
Novalaetesia LI	Oreonetides LI	Panachraesta SA	Parasynema TH
Novalena AG	Oreophantes LI	Panamomops LI	Parasyrisca GN
Novanapis AP	Orepukia AG	Panaretella HT	Paratapinocyba LI
Novaranea AR	Origanates LI	Panaretidius HT	Paratheridula TD
Nuctenea AR	Origes HT	Panaretus HT	Paratheuma DS
Nuisiana DS	Orinocosa LY	Pancorius SA	Parathiodina SA
Nukuhiva PI	Orinomana UL	Pandava TT	Paratibellus PD
Nungia SA	Ornithoctonus TP	Pandercetes HT	Paratrochosina LY
Nurscia TT	Orodrassus GN	Pandisus SA	Paratropis PA
Nycerella SA	Orongia OR	Panjange PC	Paratupua SX
Nyctimus TH	Orops SL	Panoa DS	Paratus LC
Nyssus AG	Orphaecus TP	Panysinus SA	Paratyle MT
	Orsima SA	Papakula PI	Paravoca AM
Oaphantes LI	Orsinome TG	Parabomis TH	Paravulsor CT
Oarces MM	Orsolobus OR	Parabonna GN	Parawixia AR
Obatala AM	Orthobula LC	Paracnobiopelma BA	Parazilia TG
Obrimona LI	Orthocosa LY	Parachemmis LC	Pardosa LY
Ochronetria LI	Orthonops CA	Parachtes DY	Pardosella LY
Ochyrocera OC	Orthrus SA	Paracladycnis PI	Parhedrus HT
Ocnotelus SA	Orvilleus SA	Paracleocnemis PD	Parkella SA
Ocrisiona SA	Osericta SA	Paracoelotes AM	Parnaenus SA
Octanapis AP	Osoriella AN	Paracornicularia LI	Paro LI
Octonoba UL	Osornolobus OR	Paradamoetas SA	Paroecobius OE
Oculocornia LI	Ostanes TH	Paradecta SA	Parogulnius TS
Ocyale LY	Ostearius LI	Paradescanso SA	Pasias TH
Ocyllus TH	Otacia LC	Paradictyna DI	Pasiasula TH
Odo ZO	Otagoa DS	Paradonea ER	Pasilobus AR
Odomasta ZO	Otiathops PP	Paradossenus TC	Passiena LY
Odontodrassus GN	Otira AM	Paraeboria LI	Patagoneta LI
Oecobius OE	Oxyopes OX	Paraembolides HX	Patrera AN
Oedignatha CO	Oxysoma AN	Parafluda SA	Patu SY
Oedothorax LI	Oxytate TH	Parafironeta LI	Pavocosa LY
Ogdenia SA	Ozopactus TP	Paraglyphesis LI	Pax ZD
Ogulnius TS	Ozyptila TH	Paragongylidiellum LI	Pecanapis AP
Oia LI		Paraharmochirus SA	Peckhamia SA
Oilinyphia LI	Paccius CO	Paraheliophanus SA	Pediana HT
Oinia LI	Pachistopelma TP	Parajotus SA	Pelayo AN

- Pelecopsidis LI
 Pelecopsis LI
 Pelegrina SA
 Pelicinus OO
 Pelidida LI
 Pellenes SA
 Pellolessertia SA
 Penestomus ER
 Penionomus SA
 Pensacola SA
 Pensacolops SA
 Peplometus SA
 Peponocranium LI
 Perania TB
 Perenethis PI
 Periegops SG
 Perilla TG
 Perimonoides LI
 Peritraeus TH
 Perlongipalpus LI
 Perregrinus LI
 Perro LI
 Pertica DG
 Pescennina OO
 Petricus PD
 Petrunkévitchia UL
 Peucetia OX
 Phaenius SA
 Phaenopoma TH
 Phaeocedus GN
 Phalaops PI
 Phanetta LI
 Phantias SA
 Phanotea MT
 Phantyna DI
 Pharacocerus SA
 Pharta TH
 Phaulostylus SA
 Phausina SA
 Pherecydes TH
 Pherenice AR
 Phiale SA
 Phidippus SA
 Philaeus SA
 Philisca MT
 Philodamia TH
 Philodromops PD
 Philodromus PD
 Philogaeus TH
 Philoicides AG
 Philoponella UL
 Phintella SA
 Phireza TH
 Phlattrothra LI
 Phlegra SA
 Phlogiellus TP
 Phobetinus MM
 Pholciella PC
 Pholcoides PC
 Pholcomma TD
 Pholcophora PC
 Pholcus PC
 Phoneutria CT
 Phoneyusa TP
 Phonognatha TG
 Phonophilus LI
 Phonotimpus LC
 Phormictopus TP
 Phormingochilus TP
 Phoroncidia TD
 Phricotelus MY
 Phrixotrichus TP
 Phrurolithus LC
 Phruronellus LC
 Phrurotimpus LC
 Phrynarachne TH
 Phyaces SA
 Phymatoctenus CT
 Physocyclus PC
 Physoglenes SX
 Physoplatys TH
 Phyxelida AM
 Phyxioschema DP
 Piabuna LC
 Pianoa GR
 Pickardinella TG
 Piesocalus LI
 Pikelinia FI
 Pilia SA
 Pimosa LI
 Pimus AM
 Pionothela NM
 Piranthus SA
 Pirata LY
 Piratosa LY
 Pisaura PI
 Pisarellus PI
 Pisaurina PI
 Pisenor BA
 Pistius TH
 Pitharatus AR
 Pitonga DS
 Pityohyphantes LI
 Plaesianillus LI
 Plagiobothrus BA
 Plancinus TH
 Plastonimus TH
 Platnickia ZD
 Plato TS
 Plator TR
 Plateocobius OE
 Platyarachne TH
 Platycryptus SA
 Platyoides TR
 Platypsecas SA
 Platypyresthesia TH
 Platythomisus TH
 Plectophanes CC
 Plectoptilus OO
 Plectreurus PT
 Pleorotus HT
 Plesiolena AC
 Plesiophantes LI
 Plesiophrictus TP
 Plesiothela HX
 Plexippoides SA
 Plexippus SA
 Plutonodorus PO
 Poaka PS
 Pocadicnemis LI
 Pochyta SA
 Pocobletus LI
 Poecilafoneta LI
 Poecilarcys AR
 Poecilopta CO
 Poecilochroa GN
 Poecilomigas MG
 Poeciloneta LI
 Poecilopachys AR
 Poecilorchestes SA
 Poecilotheria TP
 Poecilothomisus TH
 Poessa SA
 Polemus SA
 Polenecia UL
 Poltys AR
 Polybetes HT
 Polyboea PI
 Ponella UL
 Pongolania AM
 Porioides HH
 Porius SA
 Porotaka AG
 Porrhomma LI
 Porrhothela HX
 Porrimosa LY
 Porropis TH
 Porteria DS
 Portia SA
 Pothaeus TH
 Poultonella SA
 Pounamuella OR
 Pozonia AR
 Prasonica AR
 Prasonicella AR
 Prepotelus TH
 Prinerigone LI
 Prionolaema TG
 Prionosternum GN
 Priperia LI
 Priscipalpus LI
 Pristobaeus SA
 Priitha FI
 Proboscidula TD
 Procambidgea SF
 Procerocymbium LI
 Prochora MT
 Procleocnemis PD
 Procopius CO
 Proctonemesia SA
 Prodida PO
 Prodidomus PO
 Prodisderina OO
 Proelauna LI
 Proernus PD
 Proevippa LY
 Progradungula GR
 Proislandiana LI
 Prolycosides LY
 Promynoglenes LI
 Promyrmekiaphila CU
 Pronasoona LI
 Pronoides AR
 Pronophaea CO
 Pronopius LI
 Pronous AR
 Propostira TD
 Prorachias NM
 Prostheclina SA
 Proszynskiana SA
 Prothemenops ID
 Protoerigone LI
 Protopleptoneta LE
 Prusias HT
 Prychia HT
 Psalistops BA
 Psalmopoeus TP
 Psammoduon ZD
 Psammorygma ZD
 Psecas SA
 Psechrus PS
 Pselcis SA
 Pselligmus NM
 Psellocoptus CO
 Psellonus PD
 Pseudafroneta LI
 Pseudamphidraus SA
 Pseudamyciaea TH
 Pseudamycus SA
 Pseudanapis AP
 Pseudartonis AR
 Pseudattulus SA
 Pseudauximus AM
 Pseudemathis SA
 Pseudevippa LY
 Pseudicius SA
 Pseudocarorita LI
 Pseudoceto CO
 Pseudocorinna CO
 Pseudocorythalia SA
 Pseudoctenus CT
 Pseudocyba LI
 Pseudodrassus GN
 Pseudofluda SA
 Pseudogonatium LI
 Pseudoheliophanus SA
 Pseudohilaira LI
 Pseudohostus OX
 Pseudomaevia SA
 Pseudomaro LI
 Pseudomaso LI
 Pseudomicrocentria LI
 Pseudomicrommata HT
 Pseudonemesia MS
 Pseudopartona SA
 Pseudophthalmus AG
 Pseudoplexippus SA
 Pseudoporrhopis TH
 Pseudopsellonus PD
 Pseudopsyllo AR
 Pseudoscaphiella OO
 Pseudosparianthis HT
 Pseudosynagelides SA
 Pseudotegenaria AG
 Pseudoteyl NM
 Pseudothraphosa TP
 Pseudotriaeris OO
 Pseudotyphistes LI
 Pseudowubana LI
 Psilochorus PC
 Psilocymbium LI
 Psiloderces OC
 Psyllo AR
 Pterartoria LY

Pterartoriola LY	Rubrius AM	Scotophaeoides GN	Singaporemma TB
Pterinochilus TP	Rudra SA	Scotophaeus GN	Sinoria LI
Pterochroa GN	Rugathodes TD	Scotospilus HH	Sintula LI
Pterotricha GN	Runcinia TH	Scoturius SA	Sipalolasma BA
Pterotrichina GN	Runga SX	Scyletria LI	Siratoba UL
Ptocasius SA		Scylaceus LI	Siruasus GN
Pua MF	Saaristoa LI	Scytodes SC	Sisenna TC
Purcelliana PO	Sabahya TB	Sebastira SA	Sisicottus LI
Purumitra UL	Saccodomus TH	Sedasta AR	Sisicus LI
Pycnacantha AR	Sadies SA	Segestria SG	Sisis LI
Pycnaxis TH	Sagellula HT	Segestrioides DG	Sisyrbes LI
Pycnothele NM	Sahastata FI	Segregara ID	Sitalcas LI
Pyresthesis TH	Saitidops SA	Selamia ZD	Sitticus SA
Pyrnus GN	Saitis SA	Selenocosmia TP	Sivalicus HT
Pystira SA	Saitissus SA	Selenogyrys TP	Siwa AR
	Saloca LI	Selenops SL	Smeringopina PC
Qiyunia DI	Salpesia SA	Selenotholus TP	Smeringopus PC
Quechuella AN	Salticus SA	Selenotypus TP	Smermisia LI
Queenslanapis AP	Saltonia DI	Selenyphantes LI	Smionia GN
Quekettia SA	Sampaiosis HT	Selimus SA	Smodicinus TH
Quemedice PD	Sancus TG	Semiopyla SA	Smodix LI
	Sandalodes SA	Semljicola LI	Sobasina SA
Racata LI	Sanogasta CO	Semnolius SA	Soelteria TH
Rachias NM	Saraina SA	Semora SA	Sofanapis AP
Radulphius MT	Sarascelis PP	Semorina SA	Solenysa LI
Raecius MT	Sarinda SA	Semysmauchenius MK	Sondra SA
Rangitata AI	Sarindoides SA	Senoculifer PD	Sonoita SA
Ranops ZD	Sarotesius HT	Senoculus SN	Sosippus LY
Ransonia PI	Sarutana LE	Seothyra ER	Sosticus GN
Rapua DS	Sason BA	Seramba HT	Soucron LI
Rarahu SA	Sasonichus BA	Sergiolus GN	Soudinus LI
Rastellus AX	Sassacus SA	Sericopelma TP	Souessa LI
Raveniola NM	Satilatlas LI	Servaea SA	Souessoula LI
Rebilus TR	Satta LY	Sesieutes LC	Sougambus LI
Reinga AI	Savignia LI	Setaphis GN	Souidas LI
Reinickella TH	Scalidognathus ID	Seychellia TE	Soulgas LI
Remmius HT	Scaphiella OO	Shango DI	Spanioplanus LI
Reo MM	Scartes SA	Shapna LY	Sparianthina HT
Retiro AM	Scelidocteus PP	Shearella TB	Sparianthis HT
Rhabdogyna LI	Scelidomachus PP	Sheranapis AP	Spariolenus HT
Rhacocnemis HT	Schaeniuscelis OX	Shinobius PI	Spartaeus SA
Rhaebobates TH	Scharffia CY	Shiragaia GN	Spatala HT
Rhaeboctesis LC	Schenkella SA	Sibirocyba LI	Speleoharpactea DY
Rhene SA	Schenkeliella TG	Sicarius SI	Spelocteniza NM
Rhetenor SA	Schiapellia CO	Sidusa SA	Spelopelma TP
Rhianodes BA	Schistogyna LI	Sidymella TH	Spelungula GR
Rhinoblemma TB	Schizocosa LY	Sigytes SA	Speocera OC
Rhion DI	Schizopelma TP	Siler SA	Spermophora PC
Rhitymna HT	Sciastes LI	Silhouettella OO	Sphaerobothria TP
Rhode DY	Scirites LI	Sillemia GN	Sphecotypus CO
Rhoderia DY	Scironis LI	Sillus AN	Sphecozone LI
Rhoicinaria AM	Scolecura LI	Siloca SA	Spheropistha TD
Rhoicinus AM	Scoloderus AR	Silometopoides LI	Sphingus CO
Rhombonotus SA	Scolopembolus LI	Silometopus LI	Sphodros AT
Rhyphelia SA	Scopocira SA	Simaetha SA	Spilargis SA
Rhytidicolus CU	Scopoides GN	Simaethula SA	Spilasma AR
Rinawa HH	Scopticus TH	Simalio CL	Spinanapis AP
Ringina LI	Scortecchia CO	Simonicerca OC	Spintharidius AR
Risdonius AP	Scotargus LI	Simonurius SA	Spintharus TD
Robertus TD	Scotina LC	Simonus ZO	Spirembolus LI
Roeweriella SA	Scotinella LC	Simorcus TH	Spiroctenus NM
Rogmocrypta SA	Scotinocerus HX	Simplicistilus LI	Staberius PI
Romitia SA	Scotinotylus LI	Simprulla SA	Stagetillus SA
Rorea AI	Scotocesonina GN	Singa AR	Staianus HT
Rothus PI	Scotognapha GN	Singafrotypa TG	Stalagtia DY
Rualena AG	Scotoneta LI	Singalangia TB	Stalita DY

Stalitella DY	Synaemops TH	Tarrocanus TH	Theraphosa TP
Stalitochara DY	Synageles SA	Tarsonops CA	Theridion TD
Stanwellia NM	Synagelides SA	Tasa SA	Theridiosoma TS
Stasimopus CZ	Synalus TH	Tasata AN	Theridula TD
Stasina HT	Synaphosus GN	Tasmanapis AP	Theuma PO
Stasinoides HT	Synaphris MY	Tasmanicosa LY	Theumella PO
Steatoda TD	Synema TH	Tasmanoonops OR	Thianella SA
Stegodyphus ER	Synemosyna SA	Tatari SA	Thiania SA
Stemmops TD	Synotaxus SX	Tauala SA	Thianitara SA
Stemonyphantes LI	Synothele BA	Taurongia DS	Thiodina SA
Stenaelurillus SA	Synstrophius TH	Tautukua OR	Thiratoscirtus SA
Stenochilus ST	Syntrechalea TC	Tecution CL	Thomasettia HT
Stenodeza SA	Syrisca MT	Tedia DY	Thomisops TH
Stenoonops OO	Syroloma LY	Teemenaarus CU	Thomisus TH
Stenoterommata NM	Syroris DS	Tegenaria AG	Thoracites AR
Stenygrocerus DP	Syspira MT	Tekella CY	Thorellina AR
Stephanopis TH	Systaria CL	Tekellatus CY	Thorelliola SA
Stephanopoides TH	Systemita PC	Tekellina TD	Thoriosa CT
Stergusa SA		Tekelloides CY	Thrigmopoeus TP
Steriphopus PP	Tachygyna LI	Telamonia SA	Thwaitesia TD
Sternodes ML	Tacuna SA	Telchius OO	Thyene SA
Sertinius SA	Taczanowskia AR	Telema TE	Thyenillus SA
Stethorrhagus CO	Tafana AN	Teloleptoneta LE	Thyenula SA
Sthelota LI	Tagulinus TH	Teminius MT	Thymoites TD
Stichius SA	Tagulis TH	Temnida AN	Thyreobaeus LI
Stichoplastus TP	Tahuantina DI	Tenedos ZD	Thyreosthenius LI
Stictonanus LI	Taieria GN	Tengella TN	Thyropoeus MG
Stipax HT	Taira AM	Tennesseeillum LI	Thysanina LC
Stiphidion SF	Taivala SA	Teranodes HX	Tibellinus PD
Stiphropella TH	Takachihoa TH	Termitoonops OO	Tibellomma HT
Stiphropus TH	Takeoa ZP	Terupis AN	Tibellus PD
Stoidis SA	Talanites GN	Testudinaria AR	Tibiaster LI
Stoliczka PI	Talaus TH	Tetrablemma TB	Tibioploides LI
Storena ZD	Talavera SA	Tetragnatha TG	Tibioplus LI
Storenomorpha ZD	Tallonia PI	Tetragonophthalma PI	Tibitanus PD
Storenosoma AM	Tallusia LI	Tetromma AN	Tidarren TD
Storosa ZD	Talhybia AR	Teudis AN	Tigidia BA
Strandella LI	Tama HR	Teutamus LC	Tikaderia AG
Strandiellum HT	Tamasesia MY	Teutoniella MF	Timonoe TG
Stratius CO	Tamgrinia AM	Textricella MF	Tinus PI
Striamea DP	Tamigalesus SA	Textrix AG	Tiso LI
Stridulattus SA	Tamopsis HR	Teyl NM	Titanattus SA
Strigoplus TH	Tandil DI	Teyloides NM	Titanoeca TT
Stromatopelma TP	Tangaroa UL	Thaida AU	Titidiops TH
Strongyliceps LI	Tangata OR	Thainetes LI	Titidius TH
Strophaeus BA	Tanna SA	Thaiphantes LI	Titiotus LC
Strophius TH	Tanybelus SA	Thalassiopsis PI	Titus GN
Strotarchus MT	Taphiassa MY	Thalassius PI	Tivodrassus PO
Styposis TD	Tapinauchenius TP	Thaleria LI	Tivyna DI
Subantarctica OR	Tapinesthis OO	Thalerommata BA	Tixcocoba CL
Subbekasha LI	Tapinillus OX	Thallumetus DI	Tjurunga SF
Sudesna DI	Tapinocyba LI	Thammaca SA	Tmarus TH
Suemus PD	Tapinocyboides LI	Thanatus PD	Tmeticides LI
Suffasia ZD	Tapinopa LI	Thapsagus LI	Tmeticus LI
Sulaimania TB	Tapinothele PI	Tharpyrna TH	Tobias TH
Sulcia LE	Tapinothelella PI	Tharrhalea TH	Toddiana CY
Sulsula OO	Tapinothelops PI	Thasyraea ZO	Togacantha AR
Sumampattus SA	Tapponia OX	Thaumasia PI	Tolma HA
Supunna CO	Tara SA	Thaumastochilus ZD	Toloella SA
Sybota UL	Taranucnus LI	Thelacantha AR	Tolophus CL
Syedra LI	Tararua AG	Thelcticopis HT	Tomocyrba SA
Sylligma TH	Taraxella SA	Thelchoris DP	Tomohyphantes LI
Symmigma LI	Tariona SA	Themacrys AM	Tomopisthes AN
Symphanodes GN	Tarlina GR	Theonina LI	Tomoxena TD
Symphytognatha SY	Tarne SA	Theonoe TD	Torania HT
Symposia CB	Tarodes SA	Theotima OC	Tortolena AG

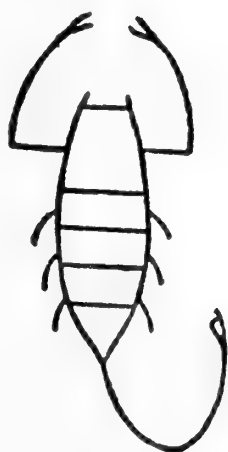
Toschia LI	Tupua SX	Vesubia LY	Xylethrus AR
Totua LI	Turbinellina LI	Viciria SA	Xyphinus OO
Toxops DS	Turinyphia LI	Victanapis AP	Xysticus TH
Toxopsiella CC	Turretia OR	Victorium LI	
Toxopsoides DS	Tusitala SA	Vidole AM	Yacolla AM
Trabaeola LY	Tutaibo LI	Vindullus HT	Yaginemaella SA
Trabea LY	Tutelina SA	Vinnius SA	Yaginumanis SA
Trabeops LY	Tuticanus CT	Viracucha CT	Yaginunia AR
Trachelas CO	Tybaertiella LI	Virgilus AM	Yakutopus LI
Trachelocamptus LI	Tychicus HT	Viridasius CT	Yamia TP
Trachelopachys CO	Tylogonus SA	Viridictyna DI	Yepoella SA
Trachycosmus GN	Tylorida TG	Viroqua SA	Yilgarnia NM
Trachyneta LI	Tymbira AM	Voraptipus PI	Yllenus SA
Trachytrema GN	Typhistes LI	Voraptus ZO	Yorima DI
Trachyzelotes GN	Typhlonesticus NS	Vuattouxia PI	Yoroa TD
Traematosisis LI	Typhlonyphia LI	Vulsor CT	Yumates OO
Trebacosa LY	Typhochrestinus LI	Vytfutia AM	Yupanquia AM
Trechalea TC	Typhochrestoides LI		
Trechona DP	Typhochrestus LI	Wabasso LI	Zachria HT
Trematocephalus LI	Typostola HT	Wadicoso LY	Zaitunia FI
Trephopoda GN		Wadotes AM	Zametopias TH
Triaeris OO	Uahuka LI	Wagneriana AR	Zametopina TH
Tricalamus FI	Uapou LI	Waiporia OR	Zangherella AP
Tricassa LY	Uduba MT	Waipoua OR	Zanomys AM
Tricellina MF	Ulidon MT	Wairua SX	Zantheres LY
Trichocyclus PC	Uloborus UL	Waitetola AM	Zavattarica GN
Tricholathys DI	Ulocymus TH	Waitkera UL	Zealanapis AP
Trichoncoides LI	Uluella SA	Wajane ER	Zealaranea AR
Trichoncus LI	Ulugurella LI	Walckenaeria LI	Zealotenus CT
Trichopagis TH	Ulwembua CY	Wallaba SA	Zearchaea MK
Trichopelma BA	Umbonata AR	Walrencea PI	Zelominor GN
Trichopterna LI	Ummeliata LI	Wanops OO	Zelotes GN
Trichothyse GN	Ummidia CZ	Waterea AI	Zenodorus SA
Tricongius PO	Upognampa GN	Wechsalia TH	Zenonina LY
Triplogyna LI	Uraarachne TH	Wedoquella SA	Zercidium TD
Tristichops ZD	Urepus AM	Wendilgarda TS	Zerogone LI
Trite SA	Uroballus SA	Wiedenmeyeria CT	Zeuxippus SA
Trittame BA	Uroctea OE	Wiehlea LI	Zigana AR
Trochanteria TR	Urocteana OE	Wiehlenarius LI	Zilephus LI
Trochosa LY	Uroecobius OE	Willisus HH	Zilla AR
Trochosippa LY	Urozelotes GN	Wiltonia OR	Zimirina PO
Trochosula LY	Ursa AR	Wirada TD	Zimiris PO
Trogloctenus CT	Usofila TE	Witica AR	Zimiromus GN
Troglodiplura NM	Uthina PC	Wixia AR	Zodarion ZD
Troglohyphantes LI	Utivarachna CO	Wubana LI	Zoica LY
Trogloneta MY	Uxuma SA	Wubanoides LI	Zophorame BA
Troglothele BA		Wulfila AN	Zophoryctes BA
Troxochrota LI	Vacchellia PD	Wulfilopsis AN	Zora ZO
Troxochrus LI	Vagellia CB		Zornella LI
Trujillina CT	Vailima SA	Xamiatus NM	Zorocrates TN
Trygetus ZD	Valdiviella LI	Xenesthis TP	Zorodictyna MT
Tuakana DS	Valonia HT	Xenonemesia NM	Zoroides ZO
Tuapoka AG	Varacosa LY	Xenoplectus GN	Zoropsis ZP
Tubercithorax LI	Vatovia SA	Xerolycosa LY	Zosis UL
Tuberta HH	Vectius GN	Xerophaeus GN	Zuniga SA
Tugana AM	Veissella SA	Xeropigo CO	Zygiella TG
Tularosa SA	Venator LY	Xestaspis OO	Zygoballus SA
Tullgrenella SA	Venonia LY	Xevioso AM	Zygomētis TH
Tulpus SA	Vermontia LI	Xingusiella TC	Zygottus LI
Tunabo ZO	Verrucosa AR	Xiombarg OO	Zyngoonops OO
Tunagyna LI	Vesicapalpus LI	Xyccarph OO	

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Volume 6

Part 1

December, 1998

Cairo, Egypt

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Subscription for volume 6 (1998-1999) :

US \$ 25.00 (personal rate)

US \$ 35.00 (institutional rate)

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Volume 1 (1987-1990), Volume 2 (1990-1992),

Volume 4 (1994-1996), Volume 5 (1996-1997):

US \$ 25.00 (p.r.) per volume

US \$ 35.00 (i.r.) per volume

Volume 3 (1992-1993):

US \$ 35.00 (p.r.), US \$ 45.00 (i.r.)

Correspondence concerning subscription, back issues, publication,
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The editor:

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Serket (1998) vol. 6(1): 1-37.

Arachnida of Egypt. I. Order Solpugida

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Introduction

In 1825, four species of Solpugida were recorded, for the first time, from Egypt by Audouin. Two species of them were recorded again by O.Pickard-Cambridge (1870) from Sinai. Simon (1880) described two new genera (*Blossia* and *Barrus*) for two new species from a place near Alexandria, to add two other families (Daesiidae and Karschiidae) to the records from Egypt. The same author (Simon, 1899) recorded another species from Wadi Natron. Tullgren (1909) added a third species to the records of Sinai, with another species from Tourah (near Cairo). Roewer (1934) in his historical work on the world's Solpugida, added six genera and fifteen species to the records of Egypt (including a new genus and 5 species new to science). In 1939, Whittick (Roewer, 1941) described a new species of *Galeodes* from El-Fayum. In 1953, Lawrence confirmed the recording of two species of *Rhagodes* from Cairo and Siwa, depending upon specimens in the collections of the British Museum (Natural History). Turk (1960) described two new species of *Galeodes* from Qena, their types were preserved in the collections of the Hebrew University of Jerusalem. Benoit (1964) recorded three species from two other localities in Egypt.

In this study, 27 species are recorded, classified within 12 genera in 5 families. Eleven species are endemic, only in Egypt.

The diagnoses of the order and families are those of Muma (1982). The key to families is adopted from El-Hennawy (1990). The keys to genera and species are adopted from the diagnoses of Roewer (1934 & 1941) in addition to Turk (1960).

Abbreviations: D = description, *f* = female, *m* = male, N = note,
Ref = references, {T} = type

Order Solpugida

Diagnosis of Order Solpugida :

Solpugids, often referred to as solifugids, are predatory arachnids. The order is distinguished from other arachnid groups by the massive, forwardly projecting, chelate chelicerae. The divided prosoma bears a pair of anteriorly located simple eyes on a headlike propeltidium. The elongate, leglike pedipalpi are provided with tarsal adhesive organs. The legs have seven segments; legs 2, 3, and 4 have divided trochanters, and leg 4 has two to five racket organs or malleoli located ventrally on the coxae and trochanters. There is no sternum. The opisthosoma or abdomen has 11 segments. There are three pairs of ventral tracheal stigmata, one between the coxae of legs 2 and 3, and one each on the posterior margins of the third and fourth sternites. There is one unpaired ventral stigma on the posterior margin of the fifth sternite (Figs. 1,2,3).

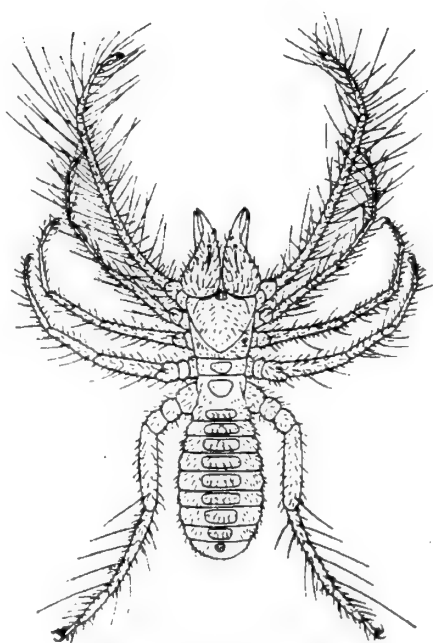
The identification of solpugid species depends mainly on the dentition of chelicerae (Fig. 4), legs' spination (Fig. 5), kinds of bristles and ctenidia (Figs. 5,6,7).

The male and female genital openings are on the second abdominal sternite, protected by two plates or opercula. The female chelicerae are strongly dentate. The male chelicerae are dentate but often modified, the immovable or fixed finger provided with a variously located and developed flagellum.

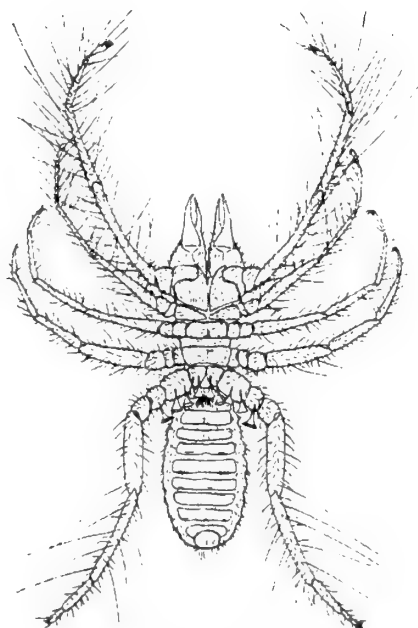
The long-legged male solpugids run rapidly over the surface of the ground, but females, especially when gravid or pregnant, are somewhat slower. All studied species are oviparous, laying masses of oval to round eggs in subterranean burrows. The eggs take several hours to as long as

4 weeks to hatch into helpless incomplete solpugids, known as embryos, first larvae, or postembryos.

Solpugids are mainly confined to arid areas and deserts of the tropical and subtropical regions of the world. When found in humid areas, they are usually restricted to well-drained sands or soils which provide xeric conditions in humid or mesic areas. Although they have been recorded from Tierra del Fuego in South America, South Africa, Canada, and the Gobi Desert in Asia, there are no known representatives from Australia or New Zealand. There are 12 uniquely geographically distributed families and over 900 species.



Solifugae; dorsal aspect. Species *Galeodes arabs*.
(After Savory, 1977, fig.99 p.234)



Solifugae; ventral aspect. Species *Galeodes arabs*.
(After Savory, 1977, fig.104 p.240)

Fig. 1. *Galeodes arabs* dorsal and ventral view (After Savory, 1977)

Key to Solpugid Families

1. Anus : ventrally located

Family RHAGODIDAE

Tarsal segmentation : 1-1-1-1

Heavy-bodied; short-legged; small to large (10-60 mm)

Leg 1 : tarsi : with a pretarsus + 2 claws,

metatarsi : with a dense ventral clothing of short spinelike setae

Male cheliceral flagellum : paraxially immovable; composed of 2 flattened, curled, setae that form a nearly complete, slightly curved, truncate, hornlike tube on the mesial surface

Female genital opercula : not differentiated from other abdominal sternites and not specifically variable

Distribution : northeastern Africa, southwestern Asia, and Near East
[26 genera, 91 species]

-. Anus : terminally located ...2

2. Tarsal segmentation : 1-4-4-(6-7) **Family SOLPUGIDAE**

Long-legged; small to large (8-60 mm)

Leg 1 : tarsi : without claws

Male cheliceral flagellum : paraxially immovable; mesodorsal to dorsal, whiplike structure separated from the fixed cheliceral finger by a suture

Female genital opercula : indistinctly differentiated from other abdominal sternites, and although they are some-times variable from one genus to another, they are not specifically so

Distribution : predominantly in Africa
[23 genera, >200 species]

-. Tarsal segmentation : 1-1-1-1 to 1-2-2-4 ...3

3. Tarsal claws of legs 2 to 4 : setaceous **Family GALEODIDAE**

Tarsal segmentation : 1-2-2-3

Long-legged; small to large (12-70 mm)

Leg 1 : tarsi : without claws or with 1 or 2 claws

Male cheliceral flagellum : paraxially movable; a single, capitate (terminally enlarged) seta located on the mesial surface

Female genital opercula : not differentiated from other abdominal sternites and not specifically variable

Distribution : northern Africa, and Asia
[4 genera, 180 species]

-. Tarsal claws of legs 2 to 4 : smooth ...4

4. Leg 1 : tarsi : without claws **Family DAESIIDAE**

Tarsal segmentation : 1-1-1-1 to 1-2-2-4

Long-legged; tiny to moderate-sized (6-23 mm)

Male cheliceral flagellum : paraxially movable, ovate to irregular membranous structure attached to the mesial surface by a disk

Female genital opercula : not differentiated from other abdominal sternites and not specifically variable

Propeltidium : exterior lobes : fused

Distribution : Africa, southern Europe, Near East, and South America
[7 subfamilies, 34 genera, 182 species]

-. Leg 1 : tarsi : with 1 or 2 claws

Tarsal segmentation : 1-1-1-1

Small to moderate-sized (8-26 mm); long-legged

Male cheliceral flagellum : paraxially immovable; fanlike to coiled, whiplike seta located on the mesial surface, with associated modified setae and a dorsal cheliceral horn

Female genital opercula : differentiated from other abdominal sternites and specifically variable

Propeltidium : exterior lobes : posteriorly fused

Distribution : Asia and Near East to southeastern Europe and northwestern Africa
[5 genera, 41 species]

Family KARSCHIIDAE

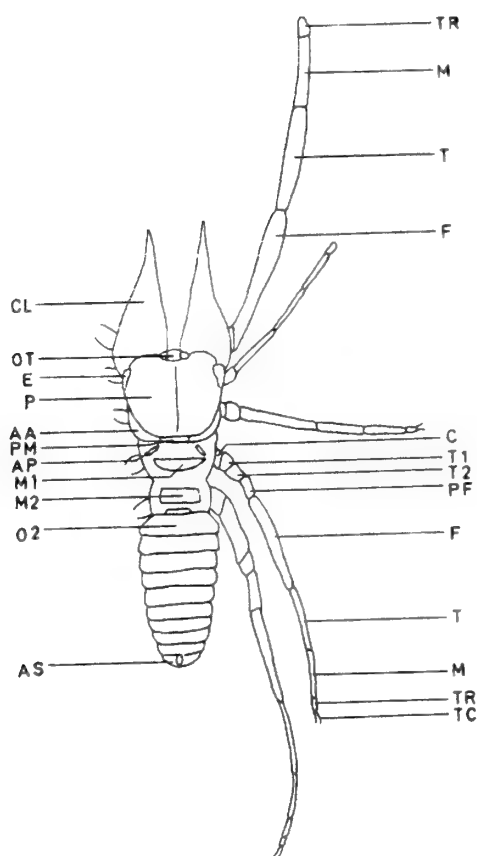


Fig. 3: *Blossia spinicornis* Lawr. ventral view. C = coxa; CT = ctenidium; GS = genital sternite; MA = malleolus; 10S = first opisthosomal sternite; 1PS-3PS = first, second and third post-genital sternites; T1 and T2 = first and second trochanters. (After Wharton, 1981. fig.2 p.8)

Fig. 2: *Blossia* sp., dorsal view. AA = arcus anterior; AP = arcus posterior; AS = anal segment; C = coxa; CL = chelicera; E = exterior lobe of prosoma; F = femur; M = metatarsus; M1 =

mesopeltidium; M2 = metapeltidium; O2 = second opisthosomal tergite; OT = ocular tubercle; P = propeltidium; PF = prefemur; PM = *plagula mediana*; T = tibia; T1 and T2 = first and second trochanters; TC = tarsal claw; TR = tarsus. (After Wharton, 1981. fig.1 p.7)

Fig. 4 *Galeodes araneoides*, female,
left chelicera, prolaterally
HZ = central tooth VZ = fore tooth
WZ = cheek-teeth ZZ = intermediate
tooth. (After Roewer, 1934. fig.47
p.52)

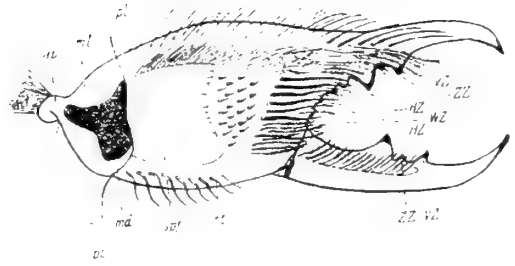


Fig. 5 *Rhagodima* sp., metatarsus and
tarsus III : b = bristles d1,2 = spines
db = spinous-bristles sh = sense hair.
(After Roewer, 1934. fig.102 p.122)

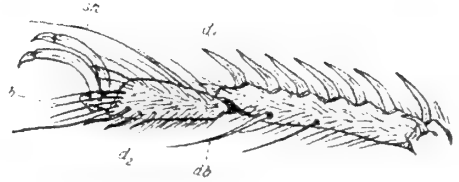


Fig. 6 Plantar-bristles of male tarsus 4 and opisthosomal ctenidia of Galeodidae in
their 4 typical forms : a = needle-like (type 1) b = spindle-shaped (type 2)
c = acutely clubbed (type 3) d = obtusely clavate (type 4).
(After Roewer, 1934. fig.316 p.511)

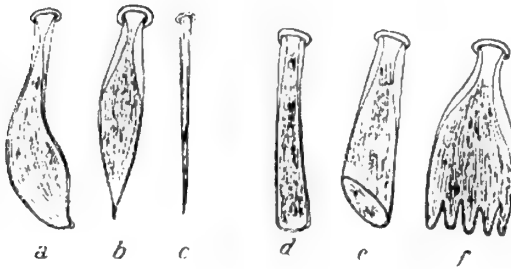


Fig. 7 Ctenidia of opisthosomal sternites :
a = clavate b = spindle-shaped c = needle-like d = staff shaped
e = obtusely clavate f = leaf-like. (After Roewer, 1934. fig.110 p.128)

Family Daesiidae

Diagnosis of the family :

Tiny to moderate-sized (6-23 mm), long-legged solpugids with a terminal anus. The exterior lobes of the propeltidium are fused. The tarsus of leg 1 lacks claws. The tarsal segmentation of legs 1 to 4 varies from 1-1-1-1 to 1-2-2-4, and the tarsal claws of legs 2 to 4 are smooth. The male flagellum is a paraxially movable, ovate to irregular, membranous structure attached to the mesial surface by a disk. The female genital opercula are not differentiated from other abdominal sternites and are not interspecifically variable.

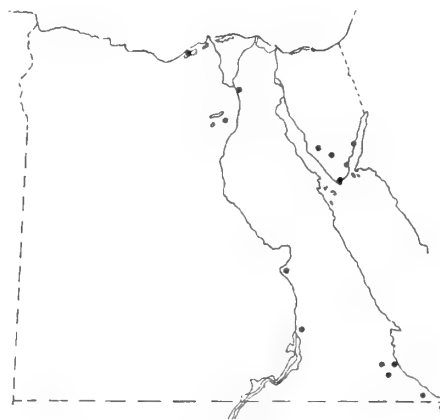
Although the reddish-brown, violet, purple, and black distinctive markings of most genera and species indicate that this family might be predominantly diurnal, *Blossiola* and *Biton* are known to be nocturnal. The biology and ecology of this family are unknown.

The family is distributed throughout Africa, southern Europe, the Near East, and South America. C.F. Roewer recognized 6 subfamilies, 26 genera, 114 species, and several subspecies. These figures now stand at 7 subfamilies, 34 genera, 182 species, and numerous subspecies. Recent workers have repeatedly criticized Roewer's generic diagnostic characters as unusable or invalid, and a generic revision is needed.

Among Roewer's larger genera are : *Hemiblossia*, *Blossiola*, *Gluviopsis*, and *Biton*.

Five species of 4 genera are recorded from Egypt.

Distribution map I.



Distribution Map of Daesiidae in Egypt

Map I.

Key to Genera :

1. Tarsi of leg 4: 4-segmented; tarsi of legs 2 and 3: 2-segmented ***Biton***
 -. Tarsi of leg 4: 1 or 2-segmented; tarsi of legs 2 and 3: 1-segmented ...2
2. Tarsi of leg 4: 1-segmented ***Gnosippus***
 -. Tarsi of leg 4: 2-segmented ...3
3. Tarsi of legs 2 and 3 with 2.2.2.2 ventral spines ***Blossia***

-. Tarsi of legs 2 and 3 with 1.2.2.2 ventral spines

Blossiola

Genus *Biton* Karsch, 1880

C.L.Koch, 1842 p.355 (sub *Gluvia* part.); Karsch, 1880 p.234 (sub *Daesia* and *Biton*); Simon, 1879 p.144 (sub *Datames*), 1882 p.252 and 253 (sub *Biton*); 1885 p.42 and 46 (sub *Biton*); Pocock, 1896 p.185, 1897 p.394 and 1898 p.522 (sub *Biton*); Purcell, 1899 p. 391-393 (sub *Daesia*); Kraepelin, 1899 p.227 and 1901 p.88 (sub *Daesia*).

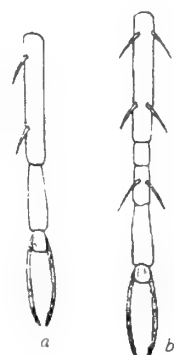
Genotype: *Biton ehrenbergi* Karsch, 1880

Diagnosis: Daesiinae, with tarsus 2 and 3 ventrally with 1.1/0 spines and tarsus 4 ventrally with 2.2/0/2/0 spines (Fig. 8 a,b). Opisthosoma ventrally at female always without, at male with or without ctenidia. Metatarsus 2-4 ventrally always with 1.2 spinous-bristles.

38 species from Africa, Asia and South-east Europe.

Fig. 8 *Biton*, ventral spination. a = tarsus 2 and 3.

b = tarsus 4. (After Roewer, 1934. fig.274 b,i p.387)



Key to males :

Common characters: Chelicerae: movable finger with 1 front, 1 or 2 small intermediate and 1 big main tooth; edge of immovable finger forms a basin behind its tip, in which the fore tooth of the movable finger almost settles (Fig. 9 b1,c); immovable finger without fore teeth; the first tooth of the immovable finger is the main tooth or the short intermediate tooth which lies before its base (Fig. 9 b1,c).

1. Chelicerae: immovable finger without intermediate teeth, with a main tooth and with 4 prolateral and 4 lateral cheek-teeth; set of teeth and flagellum (Fig. 9 b1,b2); Pedipalps ventrally on metatarsus with 1.2.2.2 true spines and tarsus with 1 prolateral spine; Opisthosoma ventrally without ctenidia; Colouration pale yellow; Propeltidium in front and both sides light brown, legs yellow, other parts brown, pedipalps on apical femur, whole tibia, metatarsus and tarsus strongly browned, often almost black; Body length 14-18 mm *B. ehrenbergi*

-. Chelicerae: immovable finger with 1 small intermediate tooth before the main tooth; set of teeth and flagellum (Fig. 9 c); Pedipalps only with bristles, tarsus without spines; Opisthosoma ventrally without ctenidia; Colouration rusty yellow, opisthosomal tergites with 3 narrow yellow

longitudinal stripes; Legs strongly brown; Pedipalps from the femur till its end quite brown; Body length 13 mm *B. bellulus*

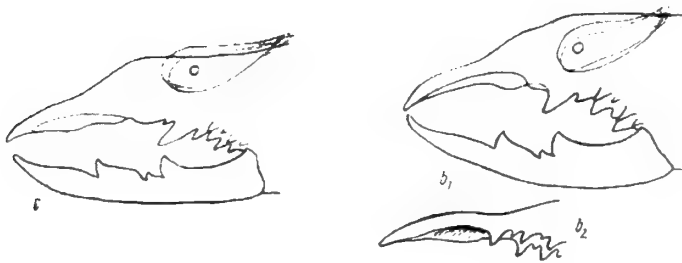


Fig. 9 Prolateral view of male right chelicera.
b1 = *Biton ehrenbergi* (b2 = tip of immovable finger oblique-lateral) c = *Biton bellulus*.
(After Roewer, 1934. fig.275 b1-2,c p.389)

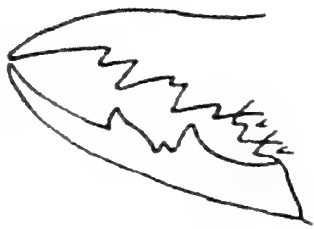


Fig. 10 *Biton ehrenbergi*, female, right chelicera, prolaterally. (After Roewer, 1934. fig.278 i p.398)

Key to females :

Common characters: Pedipalps without true spines on metatarsus, this at most with more or less stand out spinous-bristles occupied; The first fore tooth of the cheliceral immovable finger distinctly smaller than the second fore tooth.

- 1. Chelicerae: quite uniformly rusty yellow, not with dark brown longitudinal stripes; its dentition (Fig. 10); Pedipalps on tibia, metatarsus and tarsus uniformly brown, distally darker; Opisthosomal tergites with more or less distinct, brown median stripe; Body length 18 mm *B. ehrenbergi*
- . Chelicerae dorsally with 2 dark brown longitudinal stripes (often with a third lateral one); Opisthosomal tergites with 3 brownish longitudinal stripes; Body length 15 mm *B. bellulus*

Biton bellulus (Pocock, 1902) (Fig. 9 c)

Synonyms: <i>Daesia b.</i> Pocock,1902	[Roewer,1934 p.402]
World Distribution: Egypt.	Endemic Species
Local Distribution: Egypt: Wadi Sinkat ?.	

Ref: 1. *B.b.* Roewer,1934 pp.391,401,402 fig.275 c *Dmf* Wadi Sinkat?

Biton ehrenbergi Karsch, 1880 {T} (Figs. 9 b1,b2, 10)

Synonyms: *Daesia e.* Kraepelin, 1901 [Roewer, 1934 p.402]

World Distribution: Cyprus, Greece, Egypt, Ethiopia, Palestine, Saudi Arabia, Somalia, Sudan, Tunisia.

Local Distribution: Egypt: Cairo, El-Fayum, Luxor, Elephantine (Assuan).

- Ref: 1. *B.e.* Roewer, 1934 pp.390,391,400,402 fig.275 b1-b2 *Dmf*
Elephantine, Luxor, Cairo, El-Fayum
2. *B.e.* Roewer, 1941 p.140 N Palestine, Somalia
3. *B.e.* Benoit, 1964 pp.96-97 N Cairo, Elephantine (Assuan),
El-Fayum, Luxor
4. *B.e.* Delle Cave & Simonetta, 1971 pp.44-45 N

Genus *Blossia* Simon, 1880

Simon, 1880 p.399; Kraepelin, 1901 p.101 (part).

Genotype: *Blossia spinosa* Simon, 1880

Diagnosis: Blossiinae, with metatarsus 2 and 3 dorsally with a longitudinal row of 1.1.1 spines, tarsus 2 and 3 ventrally with 2.2.2.2 spines and tarsus 4 ventrally with 2.2.2/2.2 spines.

3 species from north and east Africa.

Blossia spinosa Simon, 1880 {T} (Fig. 11 a1,a2)

World Distribution: Algeria, Egypt, Palestine, Sudan.

Local Distribution: Egypt: west of Alexandria, Upper Egypt ?.

- Ref: 1. *B.s.* Simon, 1880 p.400 *Dm* near Mex, between the sea and the western horn of lake Mariout (10-14 km west of Alexandria)
2. *B.s.* Roewer, 1934 pp.371-372 fig.265 a1-a2 *Dmf* Upper Egypt? (Sudan)
3. *B.s.* Levy & Shulov, 1964 p.106 N Palestine

Habitat : It was found near the Mediterranean sea shore.

Diagnosis: Pedipalps: only metatarsus ventrally with 2.2.2 spines and tibia with cylindrical-bristles; Opisthosoma: stigmal sternite with 2-3:2-3 ctenidia (Fig. 11 a2); Colouration: rusty yellow, only metatarsus and tarsus of pedipalps dark brown; Male: Cheliceral dentition and flagellum (Fig. 11 a1); Body length: 12 mm; Female: Chelicerae: immovable finger with 3 equal in size cheek-teeth; Body length: 12-14 mm.

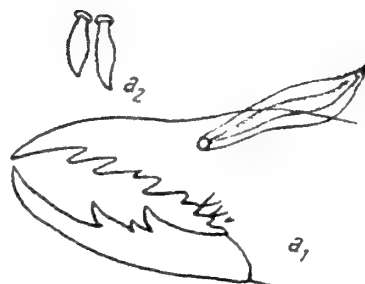


Fig. 11 *Blossia spinosa* male
a1 = right chelicera, prolaterally.
a2 = 2 opisthosomal ctenidia.

(After Roewer, 1934. fig.265 a1-2 p.371)

Genus *Blossiola* Roewer, 1934

Purcell, 1902 pp.213,214,...; Kraepelin, 1908 p.275-278 and 1914 p.128; Hewitt, 1919 p.56; Lawrence, 1928 p.267 (all sub *Blossia*).

Genotype: *Blossia unguicornis* Purcell, 1902

Diagnosis: Blossiinae, with metatarsus 2 and 3 dorsally with a longitudinal row of 1.1.1 spines, tarsus 2 and 3 ventrally with 1.2.2.2 spines and tarsus 4 ventrally with 2.2.2/2.2 spines.

30 species from north Africa, Sudan, east and south Africa, and rarely from Arabia.

Blossiola aegyptiaca Roewer, 1934 (Fig. 12)

World Distribution: Egypt, Palestine.

Local Distribution: Egypt: Upper Egypt.

Ref: 1. *B.a.* Roewer, 1934 pp.366,370 fig.264 a Df Upper Egypt (in Desert)

2. *B.a.* Levy & Shulov, 1964 p.106 N Palestine

Habitat : It was found in desert.

Diagnosis: Female only; Chelicerae: Immobile finger with 4 lateral and 4 prolateral cheek-teeth (Fig. 12); Pedipalps: tibia and metatarsus ventrally with 2.2.2.2.2 spines; Opisthosoma: without ctenidia; Colouration: body and appendages uniformly rusty yellow; Body length: 10 mm.

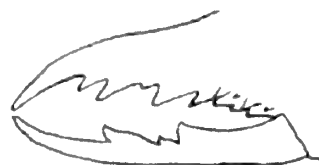


Fig. 12 *Blossiola aegyptiaca* female right chelicera, prolaterally.
(After Roewer, 1934. fig.264 a p.367)

Genus *Gnosippus* Karsch, 1880

Karsch, 1880 p.461; Kraepelin, 1899 p.231 and 1901 p.100

Genotype: *Gnosippus klunzingeri* Karsch, 1880

Diagnosis: Gnosippinae, with metatarsus 2 and 3 dorsally with a longitudinal row of 5 spines, tarsus 2 and 3 ventrally with 1.2.2.2.2 spines and tarsus 4 ventrally with 2.2.2.2.2.2 spines; Cheliceral dentition: typical Daesiidae in females (Fig. 13 a3) and greatly differentiated in males of different species; Pedipalps: metatarsus and tarsus ventrally with spines and numerous cylindrical-bristles; Opisthosomal sternites: partly with ctenidia.

3 species from Lower Egypt, Syria and Arabia.

Gnosippus klunzingeri Karsch, 1880 {T} (Figs. 13, 14)

World Distribution: Egypt.

Endemic Species

Local Distribution: Egypt: Cairo.

Ref: 1. *G.k.* Roewer, 1934 pp.355-356 fig.258 a1-a4, 259 a1-a2 *Dmf*
Lower Egypt (Cairo)

Diagnosis: Male: Chelicerae: immovable finger's tip with a dorsal spine; the intermediate tooth of the movable finger very big; dentition and flagellum (Fig. 13 a1, a2); Pedipalp with spines and spinous-bristles (Fig. 14 a1), metatarsus with numerous cylindrical-bristles; Opisthosoma: stigmal sternite with 6:6 ctenidia in the form of (Fig. 13 a4); Colouration: dirty rusty yellow, malleoli one-coloured; Body length 15 mm.

Fig. 13

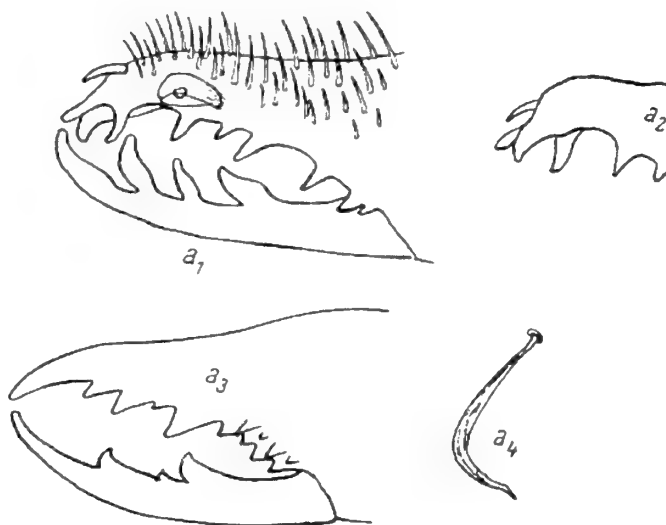
Gnosippus klunzingeri

a1 = male, right chelicera with flagellum, prolaterally.

a2 = male left chelicera, tip of immovable finger, laterally.

a3 = female, right chelicera, prolaterally. a4 = male

ctenidium. (After Roewer, 1934, fig.258 a1-4 p.354)



Female: Chelicerae: immovable finger with 4 prolateral cheek-teeth (Fig. 13 a3); Pedipalp: metatarsus and tarsus prolaterally with spines (Fig. 14 a2), metatarsus ventrally with long cylindrical-bristles; Opisthosomal sternites without ctenidia; Colouration: as in male; Body length 15-18 mm.

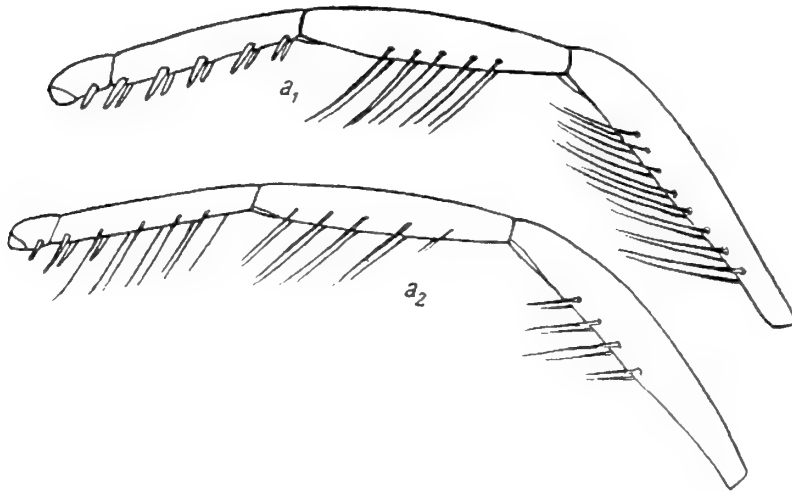


Fig. 14 *Gnosippus klunzingeri*

a1 = male, right pedipalp, prolaterally.

a2 = female, right pedipalp, prolaterally. (After Roewer, 1934. fig.259 a1-2 p.356)

In the author's collection:

Biton sp.: Bir Abraq, near El-Shalateen November 1984; Wadi Sarmatai, near Gebel Elba March 1996; El-Shalateen & Wadi Ramad May 1997; Wadi Feiran April 1998 (Southern Sinai)

Blossia sp.: Bir El-Gahliya, near El-Shalateen October 1995

Daesiidae : Ras Mohammed protectorate April 1994; Nabq protectorate May 1995; Abu Galoum protectorate July, November 1995, May 1995; St.Catherine May 1997, June 1998; Wadi Digla, near Cairo April 1998

Family Galeodidae

Diagnosis of the family :

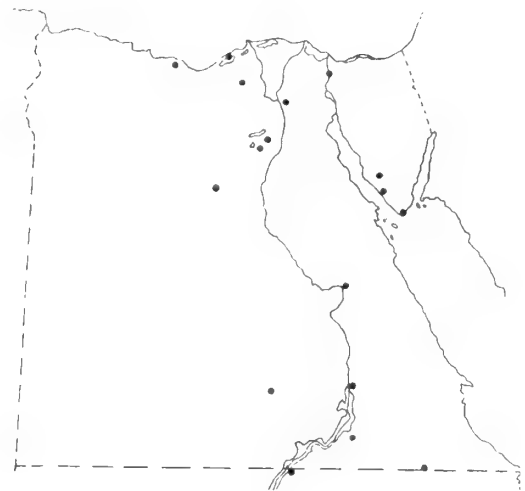
Small to large (12-70 mm), long-legged solpugids with a terminal anus. The exterior lobes of the propeltidium are fused posteriorly. The tarsus of leg 1 has no claws or one or two claws. The tarsal segmentation of legs 1 to 4 is 1-2-2-3, and the tarsal claws of legs 2 to 4 are setaceous. The male cheliceral flagellum is a single, capitate (terminally enlarged), paraxially movable seta located on the mesial surface. The female operculae are not differentiated from other abdominal sternites and are not specifically variable.

Adult and immature galeodids are omnivorous and feed on vertebrates and arthropods, including termites. The mating behaviour of 4 species in 2 genera (*Galeodes* and *Othoes*) involves the submission of the female upon pedipalpal touch or stroking by the male, kneading of the abdomen of the female with the chelicerae of the male, emission of spermatophores by the male, and the opening of the opercula of the female and the introduction of spermatophores into the female with the chelicerae of the male. Galeodids are nocturnal. They construct extensive, shallow burrows utilizing the chelicerae, pedipalpi, and metatarsal and tibial rakes of legs 2 and 3. Both *Galeodes* and *Othoes* have an annular life cycle; that of *Othoes* involves 12 stadia. *Othoes* and certain species of *Galeodes* inhabit sandy soils and sand dunes. Otherwise, their ecology is unknown.

This family is distributed throughout Asia and northern Africa. Although C.F. Roewer recognized 10 genera and 125 species, the most recent reviser of the family, F.A. Turk (1960), recognized only 4 genera, which contain 180 species : *Othoes* (26 species), *Galeodopsis* (3 species), *Paragaleodes* (6 species), and *Galeodes*, with 3 subgenera: (*Galeodellus*, 52 species; *Galeodenna*, 2 species; and *Galeodes*, 91 species).

Thirteen species of 2 genera are recorded from Egypt.

Distribution map II.



Distribution Map of Galeodidae in Egypt

Map II.

Key to Genera :

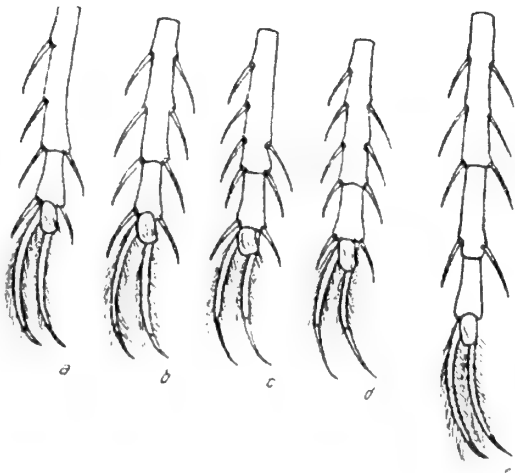
- 1. Tarsi of legs 2 and 3, segments 1 & 2 with 1.1.2/2 or 2.2.2/2 ventral spines (Fig. 15 a,b); Claws: unguiculus never more than one third the length of the pedunculus of the claw and usually only a quarter or less

Galeodes

- Tarsi of legs 2 and 3, segments 1 & 2 with 1.1.2.2/2 or 1.2.2.2/2 ventral spines (Fig. 15 c,d); Claws: unguiculus more or less half the length of the pedunculus of the claw (Fig. 20 a)

Othoes

Fig. 15 Ventral spination of tarsus of : a, b, e = *Galeodes* s.str. c, d, e = *Othoes* (c, d = tarsus 2 or 3; e = tarsus 4). (After Roewer, 1934. fig.313 d-g,k p.504)



Genus *Galeodes* Pallas, 1772

(Plate 1)

Genotype : *Galeodes araneoides* (Pallas, 1772)
Diagnosis: Galeodidae, tarsus 2 and 3 ventrally with 1.1.2/2 or 2.2.2 /2 spines and tarsus 4 ventrally with 2.2.2/2/0 spines (Fig. 15 a,b,e). Unguiculus of claws of tarsus 2-4 reach only nearly 1/4 the length of the pedunculus.
76 species Central and near Asia, in Africa southwards until Sudan; from this 34 males and females, 22 only males and 20 only females are known.

Key to males :

- 1. Movable finger with 1 intermediate tooth; immovable finger the same.. 2
- . Movable finger with 2 or 3 intermediate teeth 3

2. Metatarsus of pedipalp ventrally without cylindrical-bristles; Plantar-bristles of tarsus 4 needle-like (type 1); All sternites of opisthosoma without ctenidia; Colouration uniformly brown, opisthosoma dorsally without distinct, dark median stripe; Body length 44 mm *G. granti*
- . Metatarsus of pedipalp ventrally with cylindrical-bristles; Plantar-bristles of tarsus 4 spindle-shaped or needle-like (type 2 or 1); Sternites of the opisthosoma provided with ctenidia; Sternites 6 and 7 of the opisthosoma with a slanting row of needle-like ctenidia (type 1); Colouration sandy yellow, opisthosoma dorsally only with traces of a dark median stripe; Body length 45 mm *G. rhamses*
3. Movable finger with 2 intermediate teeth. Immoveable finger with 1 intermediate tooth 4
- . Movable finger with 3 intermediate teeth. Immoveable finger with 1 or 2 intermediate teeth 9
4. Metatarsus of pedipalp ventrally without cylindrical-bristles; Only sternite 6 of opisthosoma provided with ctenidia; Malleoli whitish yellow; Opisthosoma dorsally with completely blackish median stripe; Plantar-bristles of tarsus 4 obtusely clavate (type 4); Body length till 34 mm *G. barbarus*
- . Metatarsus of pedipalp ventrally with cylindrical-bristles 5
5. Opisthosoma ventrally without ctenidia; Plantar-bristles of tarsus 4 spindle-shaped (type 2); Colouration rusty yellow, chelicerae and propeltidium sometimes brown; Opisthosoma dorsally with blackish median stripe; Body length 31-34 mm *G. araneoides*
- . At least sternite 6 of the opisthosoma provided with ctenidia 6
6. Sternites 5-7 of the opisthosoma provided with needle-like ctenidia (type 1); Plantar-bristles of tarsus 4 needle-like (type 1); Colouration yellow-brown, opisthosoma dorsally with or without black median stripe; Body length 35-51 mm *G. arabs*
- . Only sternite 6 of the opisthosoma provided with ctenidia 7
7. Sternite 6 of the opisthosoma with spindle-shaped ctenidia (type 2); Plantar-bristles of tarsus 4 obtusely clavate (type 4); Colouration rusty yellow, opisthosoma dorsally with blackish median stripe; Pedipalps and legs with long-silky hairs; Body length 32 mm *G. sericeus*
- . Sternite 6 of the opisthosoma with needle-like ctenidia (type 1) 8
8. Tarsus 4 ventrally without especially differentiated plantar-bristles; Colouration clay-yellow, opisthosoma dorsally with a scarce pilosity, dark median stripe; Body and limbs without long-silky hairs; Body length 24 mm *G. kraepelini*
- . Tarsus 4 ventrally with dense covering, spindle-shaped (type 2); Sternite 6 of the opisthosoma with a few type 1 ctenidia; Metatarsus of pedipalps ventrally with 8-10 pairs of spines with numerous

- cylindrical- bristles; Body length 52 mm *G. medusae*
9. Immovable finger with 1 intermediate tooth; Metatarsus of pedipalps ventrally with 6 pairs of very short stout spines, with cylindrical-bristles; Body length 22 mm *G. theodori*
- . Immovable finger with 2 intermediate teeth 10
10. Metatarsus of pedipalps without cylindrical-bristles; Tarsus 4 ventrally without especially differentiated plantar-bristles; Sternite 6 of the opisthosoma with a slanting row, sternites 5 and 7 with dispersed, needle-like ctenidia (type 1); Colouration rusty yellow, opisthosoma dorsally with distinct blackish median stripe; Body length 35 mm *G. scalaris*
- . Metatarsus of pedipalps with cylindrical-bristles; Tarsus 4 ventrally with differentiated plantar-bristles 11
11. Only sternite 6 of the opisthosoma with a slanting row of needle-like ctenidia (type 1); Plantar-bristles of tarsus 4 spindle-shaped (type 2); Opisthosoma dorsally with blackish median stripe; Body length 34-43 mm *G. graecus*
- . Sternite 6 of the opisthosoma with a slanting row of spindle-shaped ctenidia (type 2); Plantar-bristles of tarsus 4 needle-like (type 1); Metatarsus of pedipalps ventrally with 2.2.2.2.2 spines and cylindrical-bristles border; Colouration golden yellow, opisthosoma with distinct, entire dark black-stripe; Body length 30 mm ... *G. veemi*

Key to females :

1. Movable finger with 1 intermediate tooth, immovable finger alike; Opisthosoma ventrally without ctenidia 2
- . Movable finger with 2 or 3 intermediate teeth; Opisthosoma ventrally with or without ctenidia 3
2. Chelicerae rusty yellow with 2 dark longitudinal stripes; Opisthosoma dorsally without dark median stripe; Body length 53 mm; Arabia, Palestine, Lower Egypt *G. granti*
- . Chelicerae uniformly rusty yellow; Opisthosoma dorsally with blackish median stripe: Body length 50 mm *G. rhamases*
3. Movable finger with 2 intermediate teeth, immovable finger with 1 intermediate tooth 4
- . Movable finger with 3 intermediate teeth, immovable finger with 2 intermediate teeth 7
4. Sternite 6 of the opisthosoma with a slanting row of needle-like ctenidia (type 1); Opisthosoma dorsally with or without dark median stripe; Body length till 51 mm *G. arabs*
- . Opisthosoma ventrally without ctenidia; Opisthosoma dorsally with distinct, seldom unsharp, dark median stripe 5

5. Pedipalps, legs, chelicerae and propeltidium provided with long silky hairs; Body length 34 mm *G. sericeus*
 - . Pedipalps, legs and body without long, dense silky hairs 6
6. Body (Opisthosoma) ventrally rusty to pale yellow; Malleoli uniformly white-yellow; Body length till 31 mm *G. araneoides*
 - . Opisthosomal pleura with dirty grey hairs; Metatarsus 2 and 3 ventrally provided with 1.2 spinous-bristles and metatarsus 4 ventrally provided with 1.22 spinous-bristles; Body length 34 mm *G. barbarus*
7. Opisthosoma upon sternite 6 with a slanting row of needle-like ctenidia (type 1); Opisthosoma dorsally with dark median stripe; Metatarsus of pedipalps ventrally provided with true spines; Metatarsus 2 and 3 ventrally with 1.2 spinous-bristles; Body length till 43 mm *G. graecus*
 - . Opisthosoma ventrally without ctenidia 8
8. Opisthosoma dorsally with entire, till the anal segment, blackish median stripe; All sternites of the opisthosoma yellowish; Body and limbs with yellow hairs; Metatarsus 4 ventrally with 1.2.2 spinous-bristles; Body length 35 mm *G. scalaris*
 - . Colouration golden yellow, opisthosoma with distinct entire dark black stripe; Metatarsus of pedipalps ventrally with 2.2.2.2.2 spines (without cylindrical-bristles); Body length 28-32 mm *G. veemi*

Galeodes arabs C.L.Koch, 1842 (Fig. 1)

C.L.Koch, 1842 p.353; Kraepelin, 1899 p.202 and 1901 p.21 (incl.var. *G. a. syriacus*, only female); Birula, 1905 p.253 - male and female.

Synonyms: *G.a.syriacus* Kraepelin, 1899 [Roewer, 1934 p.532]

World Distribution: Algeria, Arabia, Egypt, Ethiopia, Iraq, Kenya, Libya, Morocco, Palestine, Sudan, Syria, Tunisia, Yemen.

Local Distribution: Egypt: Alamein, Assuan, Cairo, Cantara (Suez canal), Tor (S.Sinai).

Ref: 1. *G.a.* Tullgren, 1909 p.1 N Tor (S.Sinai) (in June)

2. *G.a.* Roewer, 1934 pp.518-519, 522-523, 532 *Dmf* Egypt: Alamein, Assuan, Cairo

3. *G.a.* Roewer, 1941 p.161 *Nmf* Cairo, Cantara (Suez canal)

4. *G.a.* Levy & Shulov, 1964 p.109 N Palestine

Galeodes araneoides (Pallas, 1772) {T} (Figs. 16, 17)

Pallas, 1772 p.37 (sub *Phalangium*); Kraepelin, 1901 p.18 (incl. synonyms); Pocock, 1899 p.402 (= *G. truculentus*); Birula, 1892-93 p.82 and 1905 p.251 (part.) and p.252 - male and female.

Synonyms: *Phalangium a.* Pallas, 1772 [Roewer, 1934 p.526]
Solpuga a. Audouin, 1825
Solpuga intrepida Audouin, 1825 (New synonymy)
G. truculentus Pocock, 1899

World Distribution: Afghanistan, Egypt, Iran, Iraq, Jordan, Palestine, South east Russia & ex USSR, Turkey.

Local Distribution: Egypt: Upper Egypt, Wady Ferran (S.Sinai).

- Ref: 1. *Solpuga a.* Audouin, 1825 pp.176-178 pl.8 fig.7 Dmf
 2. *Solpuga intrepida* Audouin, 1825 p.178 pl.8 fig.8 Dmf
 3. *Solpuga a.* Cambridge, 1870 p.818 N Wady Ferran (Sinai), Upper Egypt [under stones, among debris of old ruins]
 4. *G.a.* Roewer, 1934 pp.516, 523-524, 526-527 Dmf
 5. *G.a.* Roewer, 1941 p.159 Nmf Palestine
 6. *G.a.* Levy & Shulov, 1964 p.109 N Palestine

Habitat : It was found under stones and among debris of old ruins.

Fig. 16 *Galeodes araneoides*, female : 7.1 = dorsal view 7.c = chelicera prolaterally. (After Audouin, 1825. pl.8 fig.7) (right)

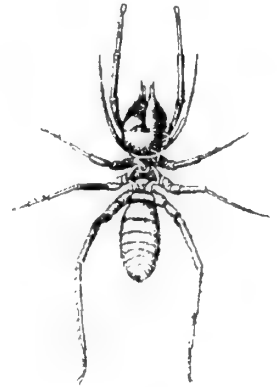
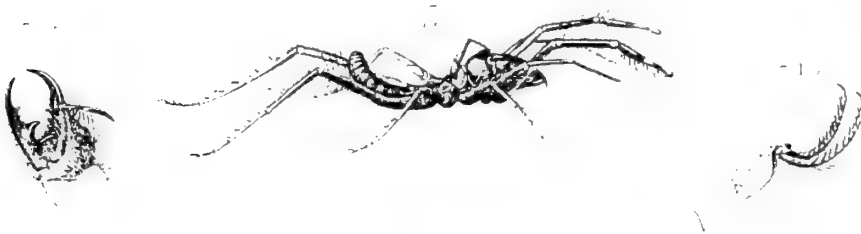


Fig. 17 *Galeodes araneoides*, male : 7.5 = lateral view 7.c = chelicera prolaterally 7.l.f = claws of tarsus IV. (After Audouin, 1825. pl.8 fig.7) (below)



Galeodes barbarus Lucas, 1846

Lucas, 1846 p.279; Kraepelin, 1901 p.24 (sub *Paragaleodes*) - male and female.

Synonyms: *Paragaleodes b.* Kraepelin, 1901 [Roewer, 1934 p.526]

World Distribution: Algeria, Egypt, Ethiopia, Libya, Morocco, Somalia, Sudan, Tunisia.

Local Distribution: Egypt: Alexandria, Cairo, El-Fayum.

Ref: 1. *Paragaleodes b.* Tullgren, 1909 p.2 N Tourah, near Cairo [under stones] (in May)

2. *G.b.* Roewer, 1934 pp.516,523,534 *Dmf* Cairo, Alexandria, Fayum

3. *G.b.* Roewer, 1941 p.162 *Nm* Somalia

4. *G.b.* Benoit, 1964 pp.95-96 N Alexandria, Cairo, El-Fayum

Habitat : It was found under stones.

Galeodes graecus C.L.Koch, 1842

C.L.Koch, 1842 p.353; Kraepelin, 1901 p.20; Werner, 1922 p.144 (sub *G. tölgi*) - male and female.

Synonyms: *G.tölgi* Werner, 1932 [Roewer, 1934 p.531]

World Distribution: Armenia, Balkan countries, Cyprus, Egypt, Greece, Macedonia, North Syria, Turkey.

Local Distribution: Egypt: Wadi Natron.

Ref: 1. *G.g.* Simon, 1899 p.244 N Bir-Hooker (Wadi Natron)

2. *G.g.* Roewer, 1934 pp.521,525,531 *Dmf*

3. *G.g.* Roewer, 1941 p.160 *Nmf* Greece, Turkey

Galeodes granti Pocock, 1903

Pocock, 1903 p.215 - male and female.

World Distribution: Egypt, Palestine, Saudi Arabia, Somalia, Sudan, Syria, Yemen.

Local Distribution: Egypt: El-Fayum.

- Ref: 1. *G.g.* Roewer, 1934 pp.515,522,532 *Dmf* Lower Egypt (Fayum)
 2. *G.g.* Roewer, 1941 p.162 *Nf* Somalia, Syria
 3. *G.g.* Benoit, 1964 p.93 *Dmf* El-Fayum
 4. *G.g.* Levy & Shulov, 1964 p.109 *N* Palestine

Galeodes kraepelini Roewer, 1934

Roewer, 1934 new name for *G. sericeus* male (not female) Kraepelin, 1899 p.205 and 1901 p.27 (sub *Paragaleodes sericeus* male) - only male.

Synonyms: *Paragaleodes sericeus* (*m*) Kraepelin, 1899

[Roewer, 1934 p.533]

World Distribution: Egypt.

Endemic Species

Local Distribution: Egypt: Upper Egypt.

- Ref: 1. *G.k.* Roewer, 1934 pp.517,533 *Dm* Upper Egypt (locality ?)

Galeodes medusae Turk, 1960 (Fig. 18)

World Distribution: Egypt.

Endemic Species

Local Distribution: Egypt: Qena.

- Ref: 1. *G.m.* Turk, 1960 pp.114-115 fig.4 *Dm* Qena (In May)

Fig. 18 *Galeodes medusae*, male, Stridulatory seta.
 (After Turk, 1960. fig.4 p.115)



Galeodes rhamses Roewer, 1934

World Distribution: Egypt.

Endemic Species

Local Distribution: Egypt: Upper Egypt, south of Assuan.

- Ref: 1. *G.r.* Roewer, 1934 pp.515,522,534 *Dmf* Upper Egypt
 (between Assuan and Korosko)

Galeodes scalaris C.L.Koch, 1842

C.L.Koch, 1842 p.353 (female) and p.354 (male ? *leucophaeus*);
Kraepelin, 1899 p.204 and 1901 p. 25 (sub *Paragaleodes*) - male and
female - Arabia, Egypt (Coast of Red Sea); Borelli, 1924 Libya and
Morocco.

Synonyms: *G.leucophaeus* ? C.L.Koch,1842 [Roewer,1934 p.533]
Paragaleodes s. Kraepelin,1899

World Distribution: Arabia, Egypt, Libya, Morocco.

Local Distribution: Egypt: Cairo and coast of the Red Sea.

Ref: 1. *G.s.* Roewer,1934 pp.519-520,526,533 *Dmf* Egypt (coast of
the Red Sea)

2. *G.s.* Roewer,1941 p.162 *Nm* Egypt (Cairo)

Galeodes sericeus (Kraepelin, 1899)

Kraepelin, 1899 p.205 and 1901 p.27 (sub *Paragaleodes* female, not
male ?) - male and female.

Synonyms: *Paragaleodes s. (f)* Kraepelin,1899 [Roewer,1934 p.533]

World Distribution: Egypt.

Endemic Species

Local Distribution: Egypt: Upper Egypt.

Ref: 1. *G.s.* Roewer,1934 pp.517,523,533 fig.317 *Dmf* Upper Egypt
(u. a. Schendi)

Fig. 19 *Galeodes theodori*, male :

1 = Stridulatory seta 2 = Left
chelicera, prolaterally 3 = Flagellum.
(After Turk, 1960. figs.2,3,3a p.114)



Galeodes theodori Turk, 1960 (Fig. 19)

World Distribution: Egypt.

Endemic Species

Local Distribution: Egypt: Qena.

Ref: 1. *G.t.* Turk, 1960 pp.113-114 fig.2,3,3a *Dm* Qena (In May)*Galeodes veemi* Whittick, 1939

World Distribution: Egypt.

Endemic Species

Local Distribution: Egypt: El-Fayum.

Ref: 1. *G.v.* Roewer, 1941 pp.166-167 *Dmf* Egypt (Fayum)**Genus *Othoes* Hirst, 1911**Genotype: *Othoes floweri* Hirst, 1911

Diagnosis: Galeodidae, with tarsus 2 and 3 ventrally with 1.1.2.2 or 1.2.2.2 spines and tarsus 4 ventrally with 2.2.2/2/0 spines (Fig. 15 c,d,e). Unguiculus of the claws of tarsus 2-4 conspicuously long and scarcely shorter than half the length of the pedunculus (Fig. 20 a). Sometimes, the second of the usual 3 front-teeth of the immovable finger is wanting, and tarsus 2 and 3 the last piece only with one spine. Two species, known only as female, one from north-west Africa, while the other is of doubtful origin.

Othoes floweri Hirst, 1911 {T} (Fig. 20 b)

Hirst, 1911 p.120

World Distribution: Egypt.

Endemic Species

Local Distribution: Egypt: Upper Egypt (Wadi Halfa).

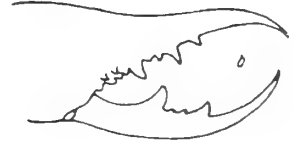
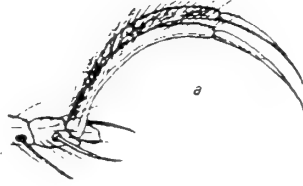
Ref: 1. *O.f.* Roewer, 1934 p.536 fig.318 a,d *Df* Upper Egypt (Wadi Halfa)

Diagnosis: Chelicerae: immovable finger with 2 intermediate teeth (there is a gap between teeth no.1 and 3, then no.2 wanting) and movable finger with 2 intermediate teeth (Fig. 20 b); Pedipalps on tibia and metatarsus

without spines and without cylindrical-bristles; Opisthosoma ventrally without ctenidia; Colouration pale yellow, opisthosoma dorsally without dark median stripe, pedipalps on tibia and metatarsus black, chelicerae and legs pale yellow; Body length 22.5 mm; Only female.

Fig. 20 a = *Othoes*, Claws of tarsus 2 (unguiculus very long and pointed, pilosity of pedunculus very fine and long).

b = *Othoes floweri*, female, left chelicera, prolaterally. (After Roewer, 1934. fig.318 a,d p.535)



In the author's collection:

Galeodes sp. Sharm El-Sheikh June 1985; Wadi El-Raiyan, near El-Fayum June 1990 (El-Hennawy, 1991 p.88); Ras Mohammed protectorate July 1994, May 1995, (Southern Sinai); Bahariya Oasis May, October 1995; Bir Frokit, on the borders with Sudan August 1997; Toshka, south of western desert September 1997

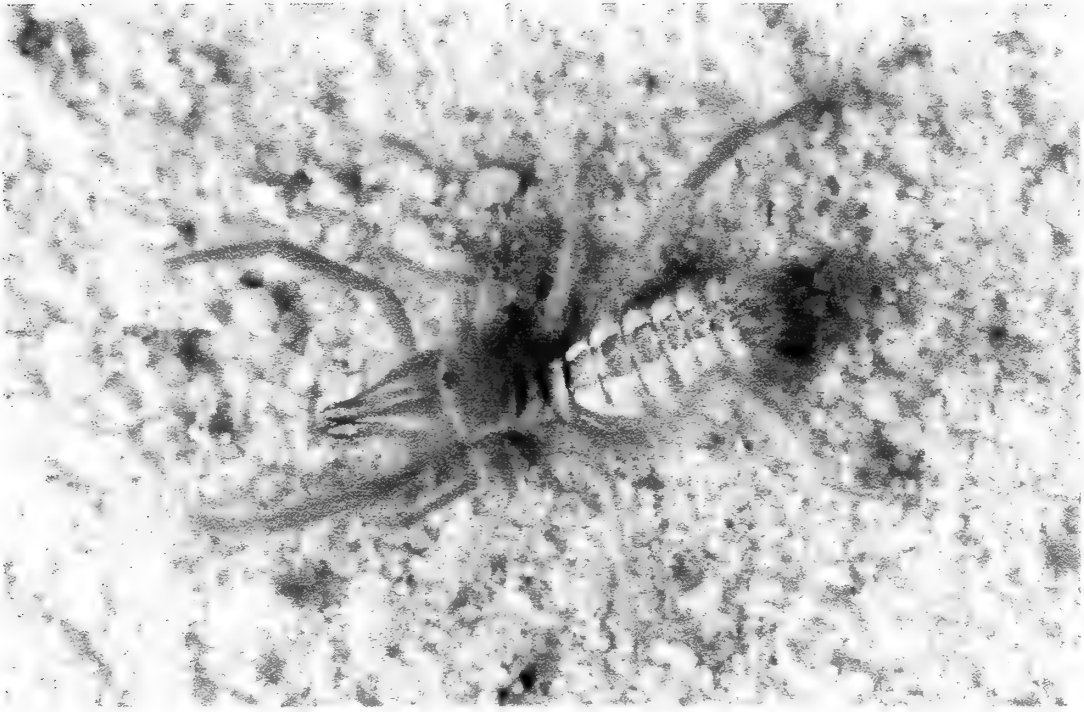


Plate 1. *Galeodes* sp.



Plate 2. *Rhagodes* sp.

Family Karschiidae

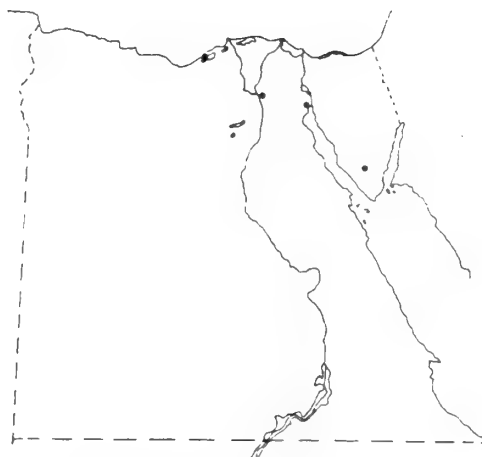
Diagnosis of the family :

Small to moderate-sized (8-20 mm), long-legged solpugids with a terminal anus. The chelicerae are multidentate. The lateral lobes of the propeltidium are posteriorly fused. The tarsi of legs 1 to 4 have one segment and two smooth tarsal claws. The male cheliceral flagellum is a fanlike to coiled, whiplike, paraxially immovable seta located on the mesial surface, with associated modified setae and a dorsal cheliceral horn. The female genital opercula are differentiated from other abdominal sternites and are specifically variable.

The biology and ecology of these solpugids are unknown. The family is distributed throughout Asia and the Near East to southeastern Europe and northwestern Africa. Five genera and 41 species are known : *Karschia* (21 species), *Barrus* (1 species), *Barrella* (4 species), *Eusimonia* (13 species), and *Rhinippus* (2 species).

Three species of 3 genera are recorded from Egypt.

Distribution map III.



Distribution Map of Karschiidae in Egypt

Map III.

Key to Genera :

1. Male flagellum with a strongly differentiated bristles tuft. Female cheliceral movable finger with 2 or more small intermediate-teeth between front and main teeth. ***Karschia***
- Male flagellum more or less oval elongated in shape, without bristles..2
2. Male flagellum, prolaterally, without a horn;
Ocular area with dense tubular hairs (Fig. 21 A). ***Barrus***
- Male flagellum, prolaterally, with a curved, more or less, blunt horn;
Ocular area and also the front edge of male's propeltidium only with normal bristles and hairs. ***Eusimonia***

Genus *Barrus* Simon, 1880

Simon, 1880 p.401; Kraepelin, 1901 p.140

Genotype: *Barrus letourneuxi* Simon, 1880 p.401 (only male).

1 species. Lower Egypt.

Barrus letourneuxi Simon, 1880 {T} (Fig. 21)

World Distribution: Egypt.

Endemic Species

Local Distribution: Egypt: Alexandria (Mex).

Ref: 1. *B.l.* Simon, 1880 pp.401-402 Dm Mex, near Alexandria
[In December]

2. *B.l.* Roewer, 1934 pp.305-306 fig.227 A-D Dm Lower Egypt:
Alexandria (Le Mex)

Diagnosis: Dorsal side of chelicera with 2 long, thin bristles (Fig. 21 A). Pedipalp ventrally: femur prolaterally with a group of 4 spines, tibia without spines, metatarsus with 4 spines and tarsus with 1 spine (Fig. 21 B). Opisthosoma: sternite 4 laterally at both stigma, oblique line of 3 ctenidia (Fig. 21 C), sternite 5 laterally at both stigma, oblique line of 12 ctenidia (Fig. 21 D). Body length 9.5 mm.

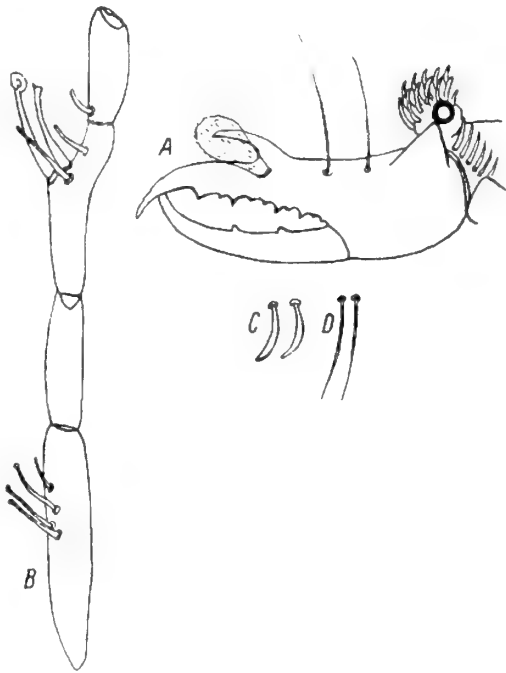


Fig. 21 *Barrus letourneuxi*, male.

A = left chelicera and ocular tubercle with front margin of propeltidium, prolaterally.
B = left pedipalp ventrally. C = 2 ctenidia of opisthosomal sternite 4. D = 2 ctenidia of opisthosomal sternite 5. (After Roewer, 1934. fig.227 A-D p.306)

Genus *Eusimonia* Kraepelin, 1899

Kraepelin, 1899 p.249

Genotype: *Eusimonia furcillata* (Simon, 1872)

7 species (no female). North Africa till Syria, Arabia, Iran, Turkey.

Eusimonia kabiliana (Simon, 1879) (Fig. 22)

Synonyms: *Gluvia k.* Simon, 1879

[Roewer, 1934 p.302]

World Distribution: Algeria, Egypt.

Local Distribution: Egypt: Cairo, Suez ?.

Ref: 1. *E.k.* Roewer, 1934 pp.301-302 fig.224 D Dmf Lower Egypt:
Cairo, Suez ?

Diagnosis: Male flagellum slender, skinny, sloping; Pedipalp: metatarsus only with ventral spines, and tarsus not hairy; Opisthosoma: sternite 4 with 6-8 short roll-shaped ctenidia on both stigma; sternite 5 with one cross row of 10 short, apical spade-shaped ctenidia; Body length 8 mm.

Fig. 22 *Eusimonia kabiliana*, prolateral view of male right chelicera, flagellum complex (after Kraepelin). (After Roewer, 1934. fig.224 D p.300)



Genus *Karschia* Walter, 1889

Genotype: *Karschia cornifera* Walter, 1889

16 species. Near and central Asia.

Karschia sp.

Local Distribution: Egypt: St.Catherine, Southern Sinai.

In the author's collection:

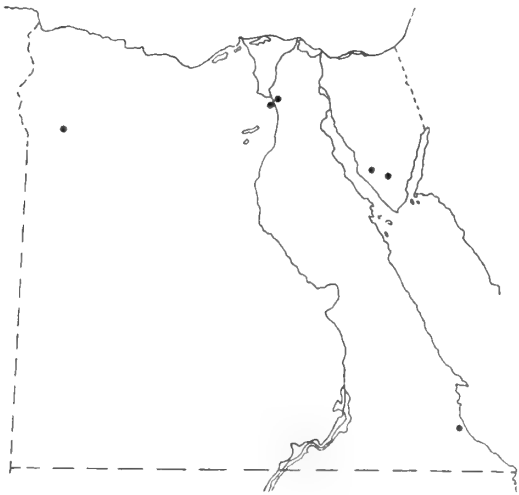
Karschia sp.: St.Catherine, Southern Sinai November 1997

Family Rhagodidae

Diagnosis of the family :

Small to large (10-60 mm), heavy-bodied, short-legged solpugids with a ventrally located anus. The exterior lobes of the propeltidium are free. The tarsi of leg 1 have a pretarsus and two claws. The metatarsi of leg 1 have dense ventral colthing of short spinelike setae. The tarsi of legs 2 to 4 have one segment and two smooth claws. The male cheliceral flagellum is composed of two flattened, curled, paraxially immovable setae that form a nearly complete, slightly curved, truncate, hornlike tube on the mesial surface. The female genital opercula are not differentiated from other abdominal sternites and are not specifically variable.

The biology and ecology of these solpugids are unknown. The family is distributed throughout north-eastern Africa, the near East, and south-western Asia. Although C.F.Roewer recognized 84 species in 26 genera, recent workers have demonstrated that instability exists in his sub-familial, generic, and specific diagnostic characters. More dependable characters are needed. According to Roewer, the largest genera are *Rhagodes* and *Rhagodoca*. Ninety one species are known.



Distribution Map of Rhagodidae in Egypt

Four species of 2 genera are recorded from Egypt. Map IV.
Distribution map IV.

Key to Genera :

- 1. Tarsi of legs 2-4 : without spines *Rhagodes*
- . Tarsi of legs 2-3 : with 1.1 spines; Tarsi of leg 4 : with 1.2.2 spines *Rhagoditta*

Genus *Rhagodes* Pocock, 1897 (Plate 2)

Genotype: *Rhagodes melanus* (Olivier, 1807)
Diagnosis: Tarsi of legs 2-4 without spines.

11 species. North Africa, Tanzania, Near Asia (Palestine till Pakistan).

Key to species :

1. Coxa 1-3 without bacilli; Body, legs, pedipalps and malleoli blackish;
 Body length 20-25 mm *R. melanus*
- . Coxa 1-3 with many bacilli 2
2. Malleoli black; body, chelicerae black; opisthosomal tergites 8 & 9
 only white; pedipalps and leg 1 rusty yellow with dark brown tarsus
 and metatarsus; legs 2-4 rusty yellow with all coxae brownish; Body
 length 14 mm *R. aegypticus*
- . Malleoli white; body, chelicerae dark brown, hairy; all coxae and legs
 pale brown with darker hairs; pedipalpal tarsus and metatarsus black;
 Body length till 30 mm *R. furiosus*

Rhagodes aegypticus Roewer, 1934

World Distribution: Egypt. Endemic Species
 Local Distribution: Egypt: Cairo.

Ref: 1. *R.a.* Roewer,1934 pp.269-270 Df Egypt (Cairo) (only female)

Rhagodes furiosus (C.L.Koch, 1842)

C.L.Koch, 1842 p.354 (sub *Rhax*); Kraepelin, 1901 p.34 - male and female.

Synonyms: *Rhax f.* C.L.Koch,1842 [Roewer,1934 p.270]
 World Distribution: Egypt, Libya.
 Local Distribution: Egypt: Cairo, Gizah.

Ref: 1. *R.f.* Roewer,1934 pp.269-270 N Egypt, Cyrenaica, Audjila
 2. *R.f.* Roewer,1941 p.101 Nm Egypt (Gizeh)
 3. *R.f.* Lawrence,1953 p.955 Nmf Abassia, Cairo

Rhagodes melanus (Olivier, 1807) {T} (Fig. 23)

Olivier, 1807 p.308 (sub *Galeodes*); Kraepelin, 1901 p.34 - male and female.

Synonyms: *Galeodes m.* Olivier, 1807

[Roewer, 1934 p.269]

Solpuga m. Audouin, 1825

World Distribution: Algeria, Egypt, South Palestine.

Local Distribution: Egypt: Siwa, Wady Ferran (Southern Sinai).

Ref: 1. *Solpuga m.* Audouin, 1825 pp.178-179 pl.8 fig.9 Dmf

2. *Solpuga m.* Cambridge, 1870 p.818 N Wady Ferran (Sinai)

3. *R.m.* Roewer, 1934 pp.269-270 Dmf North east Africa: Egypt; Algeria; South Palestine

4. *R.m.* Lawrence, 1953 p.955 Nmf Siwa

5. *R.m.* Levy & Shulov, 1964 p.103 N South Palestine

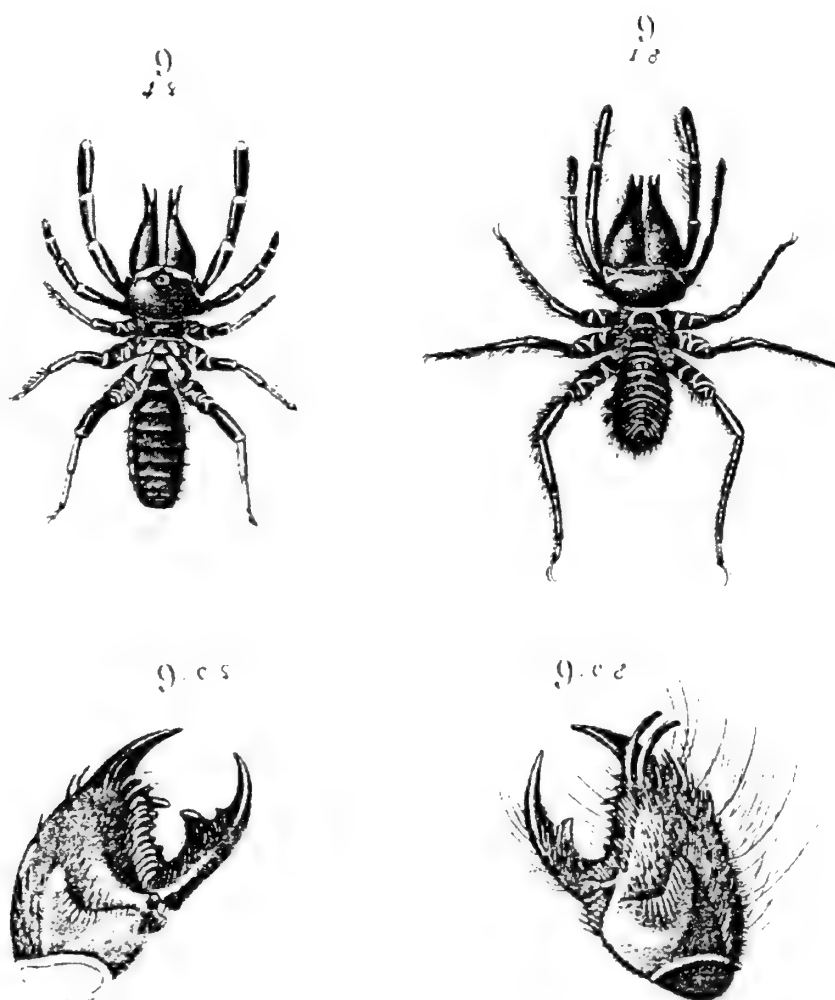


Fig. 23 *Rhagodes melanus* :

male : 9.1 = dorsal view 9.c = chelicera prolaterally

female : 9.4 = dorsal view 9.c = chelicera prolaterally.

(After Audouin, 1825. pl.8 fig.9)

Genus *Rhagoditta* Roewer, 1934

Genotype: *Rhagoditta phalangium* (Olivier, 1807)

Diagnosis: Tarsi of legs 2 and 3 with 1.1 spines; tarsus of leg 4 with 1.2.2 spines.

5 species. North and east Africa, Iran.

Rhagoditta phalangium (Olivier, 1807) {T} (Fig. 24)

Olivier, 1807 p.308 (sub *Galeodes*); Kraepelin, 1901 p.35 (sub *Rhagodes*) and Birula, 1926 p.184 (sub *Rhagodes*) - male and female.

Synonyms: *Galeodes p.* Olivier, 1807 [Roewer, 1934 p.278]

Rhagodes p. Kraepelin, 1901

World Distribution: Egypt, Ethiopia, Somalia.

Local Distribution: Egypt: Upper Egypt.

Ref: 1. *Solpuga p.* Audouin, 1825 p.179 pl.8 fig.10 Dm

2. *R.p.* Roewer, 1934 pp.278-279 Dmf Upper Egypt, Abessinya, Obock

Diagnosis: Tibiae of legs 2 and 3 dorsally with 2 apical spines; coxae 1-3 with a slanting row of 4-6 strong bacilli; metatarsi 2-4 ventrally with 1.1.1.2, 1.1.2.2, 1.1.2.3 spinous-bristles; Malleoli yellowish white; propeltidium black, with frontal margin pale yellow; opisthosoma dorsally with quite black broad longitudinal band, pleura grey with red hairs; chelicerae with red fingers; all coxae and sternites rusty yellow; all legs and pedipalps rusty yellow with tarsus and metatarsus apically red; Body length 22-24 mm.

10 c

1 c

Fig. 24 *Rhagoditta phalangium*, male :
10.1 = lateral view
10.c = chelicera prolaterally.
(After Audouin, 1825. pl.8 fig.10)



In the author's collection:

Rhagodes sp.: St.Katherine December 1987; El-Shalateen March 1996

Family Solpugidae

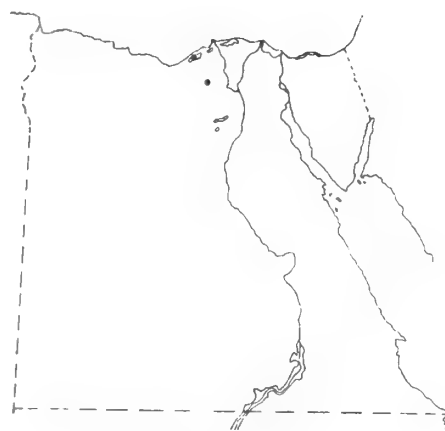
Diagnosis of the family :

Small to large (8-60 mm), long-legged solpugids with a terminal anus. The exterior lobes of the propeltidium are posteriorly fused. The tarsi of leg 1 lack claws. The tarsal claws of legs 2 to 4 are smooth. The tarsal segmentation of legs 1 to 4 varies from 1-4-4-6 to 1-4-4-7. The male cheliceral flagellum is a paraxially immovable, mesodorsal to dorsal, whiplike structure separated from the fixed cheliceral finger by a suture. The female opercula are indistinctly differentiated from other abdominal sternites, and although they are sometimes variable from one genus to another, they are not specifically so.

Adults are omnivorous, but *Solpuga sericea* is known to be termitophagous. The food habits of immature individuals are not known. The mating behaviour is unknown. Both diurnal and nocturnal species are known; the former are capable of climbing shrubs and trees, and the latter often construct long shallow burrows, utilizing the chelicerae, or they hide in natural crevices or cavities. These solpugids are believed to live for several years. Their ecology is unknown.

The family is predominantly African. Twenty three genera and over 200 species have been described. The largest genera are *Zeriassa*, *Solpuga*, and *Solpugema*. Since recent workers have demonstrated that instability exists in the generic and specific diagnostic characters of C.F. Roewer, the family cannot be delineated.

Two species of 1 genus are recorded from Egypt. Distribution map V.



Distribution Map of Solpugidae in Egypt

Map V.

Genus *Oparbella* Roewer, 1934

Genotype: *Oparbella flavescens* (C.L.Koch, 1842)

Diagnosis: Solpuginae, with ocular tubercle with an irregular bush of partly blunt bristles. Deutosternum staff-shaped. Prosoma and pedipalps without spines. Male pedipalp's metatarsus with scopula and cylindrical-bristles. Legs: leg 4 without mane; metatarsus 2-3 dorsally with a long

row of 5 spines (Fig 25 A,d1-d5); tarsus 2, segment 1 of both male and female, dorsolaterally with a long row of 7 spinous-bristles (Fig 25 A,b1-7); tarsus 2-3, segments 2-4 with 2/1/2 spines ventrally (Fig 25 A,v); tarsus 4, segments 1-6 with 2/2/0/2/0/2 spines ventrally; tarsus 2-4 with accessory spines among ventral pairs of spines.

5 species. North and west Africa. (One of them known only as a female).

Key to males :

1. Flagellum shaft long, extends above the basal part till after or over the ocular tubercle (Fig. 25 B), its end bent upwards (Fig. 25 B1); Colouration rusty yellow; propeltidium frontally often brownish; opisthosomal tergites with a dark median stripe; pedipalps rusty yellow, its metatarsus and tarsus dark brown to black; legs and malleoli rusty yellow; Body length till 22 mm *O. flavescens*
- . Flagellum shaft short, scarcely extends after the basal part, its end oblique cut (Fig. 25 C1); Colouration: like *O. flavescens*, nevertheless chelicerae with 2 dark longitudinal stripes and opisthosomal tergites without indication of a dark stripe; malleoli white; Body length 14 mm *O. quedenfeldti*

Key to females :

1. Metatarsus of pedipalp ventrally with numerous cylindrical bristles; Colouration as in male; Body length 28-38 mm *O. flavescens*
- . Metatarsus of pedipalp ventrally nearly without cylindrical bristles; Chelicerae, propeltidium rusty yellow, frontally often brownish; opisthosoma pale yellow, only its anterior tergites lightly brownish; pedipalps rusty yellow, metatarsus and tarsus dark brown to black; legs uniformly rusty yellow; malleoli whitish; Cheliceral immovable finger: the intermediate tooth separated from main tooth by a deep gap (Fig. 25 C2); Body length 22 mm *O. quedenfeldti*

Oparbella flavescens (C.L.Koch, 1842) {T} (Fig. 25 A,B,B1)

C.L.Koch, 1842 p.353; Kraepelin, 1901 p.58 (sub *Solpuga*)

Synonyms: *Solpuga f.* C.L.Koch,1842 [Roewer,1934 p.483]

World Distribution: Algeria, Egypt, Libya, Morocco, Togo, Tunisia.

Local Distribution: Egypt: Alexandria.

Ref: 1. *O.f.* Roewer,1934 pp.481-483 fig.306 A,B,B1 *Dmf* Alexandria

Oparbella quedenfeldti (Kraepelin, 1899) (Fig. 25 C,C1,C2)

Kraepelin, 1899 p.214 and 1901 p.67 (sub *Solpuga*)

Synonyms: *Solpuga q.* Kraepelin, 1899

[Roewer, 1934 p.483]

World Distribution: Egypt, Morocco.

Local Distribution: Egypt: Wadi Natron.

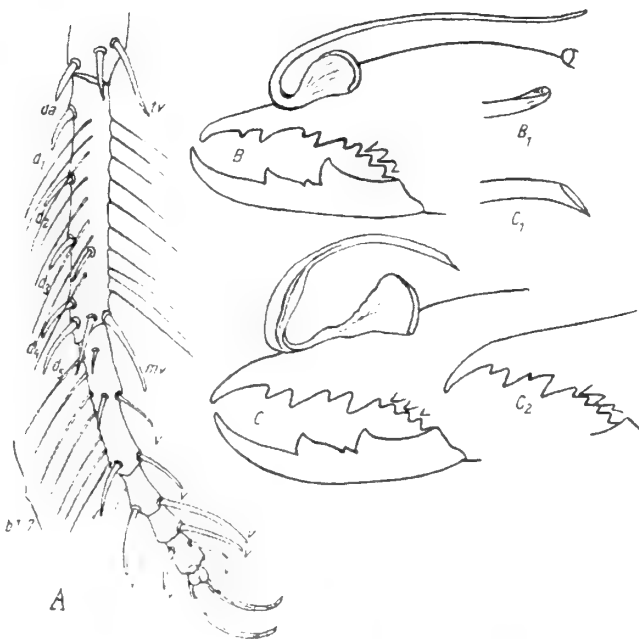
Ref: 1. *O.q.* Roewer, 1934 pp.482-483 fig.306 C,C1,C2 Dmf

2. *O.q.* Roewer, 1941 p.156 Nf Wadi Natron

Fig. 25 *Oparbella flavescens*

A = left leg 2 lateroventrally :
tibia (da = 1 dorsal apical and
tv = 2 ventral apical spines),
metatarsus (d1-d5 = dorsal
longitudinal row of spines, mv =
ventral apical spine pair), and
tarsus (b1-7 = dorsal longitudinal
row of spinous-bristles of
segment 1 and v = ventral
spination of segments 1-4)

B = right male chelicera,
prolaterally, B1 = shaft-tip of
flagellum, enlarged (after Type).



Oparbella quedenfeldti

C = right male chelicera, prolaterally. C1 = shaft-tip of flagellum, laterally, enlarged.

C2 = right immovable finger of female chelicera, prolaterally (after Type). (After Roewer, 1934. fig.306 A,B,C p.481)

Acknowledgments

The author is very grateful to Dr. Jürgen Gruber of Naturhistorisches Museum Wien for the photocopy of Roewer's " Solifugae, ... " (1934) without which the achievement of this work was not possible.

Most of the solpugid specimens in the author's collection were collected by his friends: Mr. Mahmoud S. Abd El-Daiem, Dr. Hassan H. Fadl, Dr. Alaa El-Din A. Moustafa, Mr. Gamal Orabi and Dr. Wolfgang Ullrich. Many thanks to all of them.

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no. 2
1999



SERKET

سرکت



SERKET

Volume 6

Part 2

August, 1999

Cairo, Egypt

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Subscription for volume 6 (1998-1999) :

US \$ 25.00 (personal rate)

US \$ 35.00 (institutional rate)

Back issues :

Volume 1 (1987-1990), Volume 2 (1990-1992),
Volume 4 (1994-1996), Volume 5 (1996-1997):

US \$ 25.00 (p.r.) per volume

US \$ 35.00 (i.r.) per volume

Volume 3 (1992-1993):

US \$ 35.00 (p.r.), US \$ 45.00 (i.r.)

Correspondence concerning subscription, back issues, publication,
etc. should be addressed to the editor.

The editor: Hisham K. El-Hennawy
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 Heliopolis, Cairo 11341, Egypt.

Serket (1999) vol. 6(2): 38-44.

**Review of European scorpions,
with a key to species**

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Introduction

The idea to compile a synoptic table of European scorpions stems from frequent suggestions and requests made by naturalists interested in arachnology but not specializing in scorpions. This table, in combination with a simple key, should help the non-specialist in determination of taxa occurring in Europe. Since this is an overview, locality data are omitted and authors verifying occurrences are not cited. The article summarizes hitherto known data scattered in difficult-to-obtain literature.

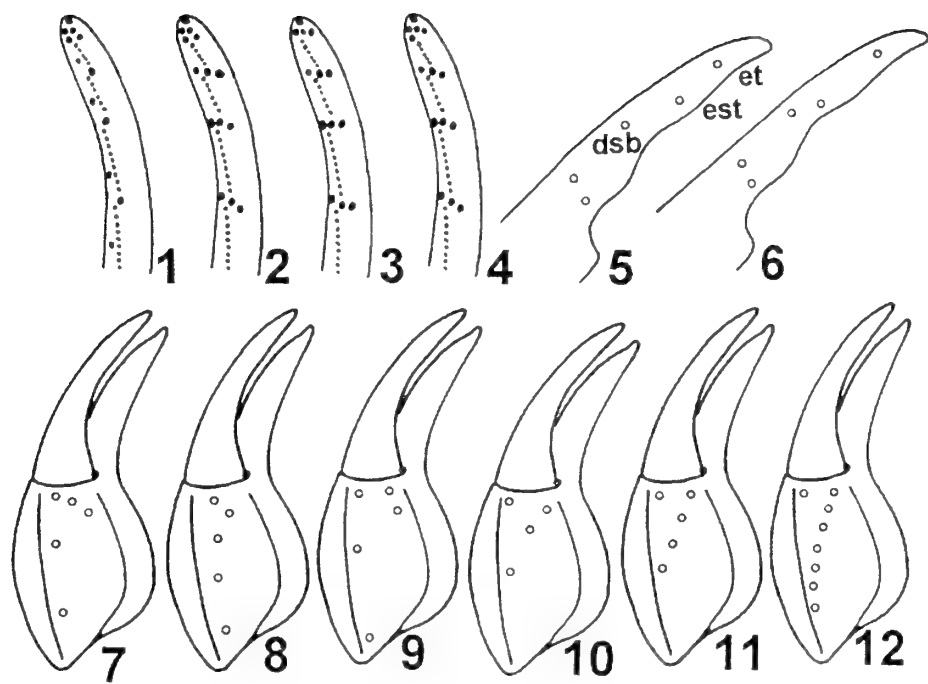
A key to European scorpions

- 1. Patella of pedipalp (Fig. 13E) without ventral trichobothria
..... Buthidae 2
- . Patella of pedipalp with one or more ventral trichobothria 10

2. Animal yellow or yellowish brown 3
- Animal black 8
3. First two segments of mesosoma (Fig. 13F) with five keels
 *Leiurus quinquestriatus* Hemprich & Ehrenberg, 1828*
- First two segments of mesosoma with three keels 4
4. Movable finger of pedipalp (Fig. 13A) with external lateral granules
 (Figs. 2-4) 5
- Movable finger of pedipalp without external lateral granules (Fig. 1)
 *Compsobuthus matthiesseni* Birula, 1905*
5. Movable finger of pedipalp (Fig. 13A) with four principal distal
 granules and one terminal granule (Fig. 2) *Mesobuthus*.... 6
- Movable finger of pedipalp with three principal distal granules and one
 terminal granule (Fig. 3) *Buthus occitanus* (Amoreux, 1789)
6. Fourth segment of metasoma (Fig. 13H) with 10 keels
 *Mesobuthus gibbosus* (Brullé, 1832)
- Fourth segment of metasoma with eight keels 7
7. Movable finger of pedipalp (Fig. 13A) with at most 12 diagonal rows
 of granules. Length from chelicerae (Fig. 13D) to end of metasoma 40
 to 55 mm *Mesobuthus eupeus eupeus* (C. L. Koch, 1839)
- Movable finger of pedipalp with 13 or 14 diagonal rows of granules.
 Length of adult 60 to 80 mm
 *Mesobuthus caucasicus caucasicus* (Nordmann, 1840)
8. Movable finger of pedipalp (Fig. 13A) with four principal distal
 granules and one terminal granule (Fig. 4)
 *Hottentotta judaica* (Simon, 1872)*
- Movable finger of pedipalp with three principal distal granules and one
 terminal granule (Fig. 3) *Androctonus* 9
9. Manus of pedipalp (Fig. 13C) narrow. Dorsal surface of first
 metasomal segment granulated at midline
 *Androctonus bicolor* (Hemprich & Ehrenberg, 1828)
- Manus of pedipalp broad. Dorsal surface of first metasomal segment
 smooth *Androctonus crassicauda* (Olivier, 1807)
10. Manus of pedipalp (Fig. 13C) very broad, entirely rounded, and about
 as wide as long
 *Scorpio maurus fuscus* (Hemprich & Ehrenberg, 1829)*
- . Manus not entirely rounded and longer than wide 11

11. Number and distribution of trichobothria on ventral side of manus (Fig. 13C) corresponds to Fig. 7 *Calchas nordmanni* Birula, 1899
- . Number and distribution of trichobothria on ventral side of manus corresponds to Fig. 8 *Iurus* 12
- . Number and distribution of trichobothria on ventral side of manus corresponds to Fig. 9 *Belisarius xambeui* Simon, 1879
- . Number and distribution of trichobothria on ventral side of manus corresponds to Fig. 10 *Euscorpius* (*Euscorpius*) 13
- . Number and distribution of trichobothria on ventral side of manus corresponds to Fig. 11
..... *Euscorpius* (*Tetratrachobothrius*) *flavicaudis* (De Geer, 1778)
- . Number and distribution of trichobothria on ventral side of manus corresponds to Fig. 12 (a row may be composed of a different number of trichobothria, but no less than six)
..... *Euscorpius* (*Polytrichobothrius*) *italicus* (Herbst, 1800)
12. Adult male has fingers of pedipalps (Fig. 13A and B) bent so that at closure parts of their surfaces do not meet. Occurs only in Turkey *Iurus asiaticus* Birula, 1903
- . Adult male has fingers of pedipalps bent so that at closure their entire surfaces meet. Occurs only in Greece
..... *Iurus dufourei* (Brullé, 1832)
13. Ventral side of fifth metasomal segment (Fig. 13G) smooth and rounded 14
- . Ventral side of fifth metasomal segment bears central granules that usually form a conspicuous keel
..... *Euscorpius* (*Euscorpius*) *carpathicus* (Linné, 1767)
14. Distance between trichobothria dsb - est and est - et on fixed finger of pedipalps is about equal (Fig. 5)
..... *Euscorpius* (*Euscorpius*) *germanus* (C. L. Koch, 1837)
- . Distance between trichobothria est - et on fixed finger of pedipalp is about twice as long as that between trichobothria dsb - est (Fig. 6)
..... *Euscorpius* (*Euscorpius*) *mingrelicus* (Kessler, 1876)

* denotes species that occur only in the Asian parts of Turkey and are included to cover the entire Turkish scorpion fauna.



Figs 1-12. Figs 1-4. Dorsal aspect of movable fingers of pedipalps. Figs 5-6. Dorsal-external aspect of fixed fingers of pedipalps. Figs 7-12. Chela ventral. Schematic drawing of chela in ventral view. Fig. 1. *Compsobuthus matthiesseni*. Fig. 2. *Mesobuthus gibbosus*. Fig. 3. *Buthus occitanus*. Fig. 4. *Hottentotta judaica*. Fig. 5. *Euscorpius* (*Euscorpius*) *germanus*. Fig. 6. *E. (E.) mingrelicus*. Fig. 7. *Calchas nordmanni*. Fig. 8. *Iurus dufourei*. Fig. 9. *Belisarius xambeui*. Fig. 10. *E. (E.) carpathicus*. Fig. 11. *E. (Tetratrachobothrius) flavicaudis*. Fig. 12. *E. (Polytrichobothrius) italicus*.

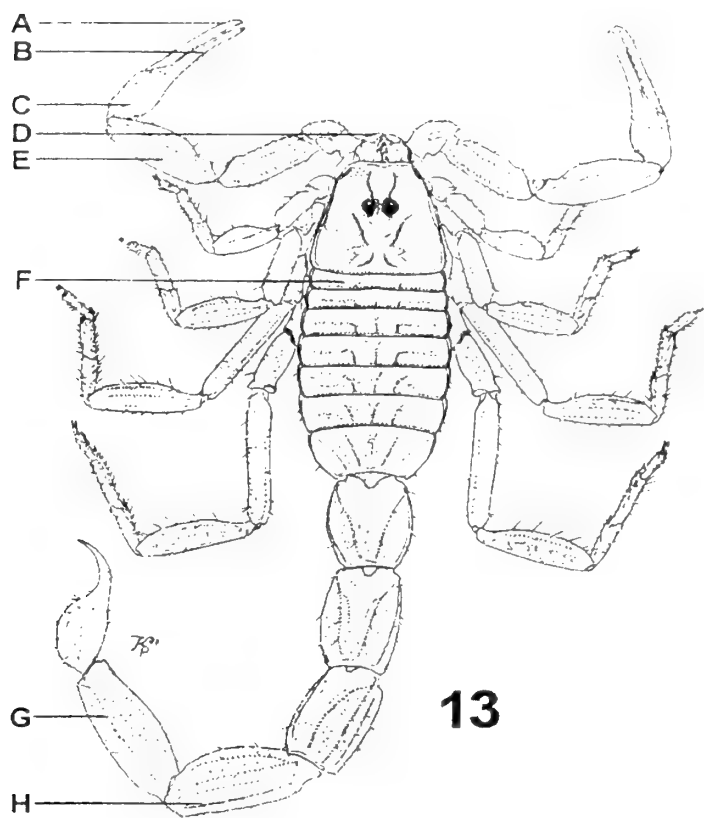


Fig. 13. Dorsal view of a scorpion.
A. Movable finger.
B. Fixed finger.
C. Manus.
A, B, and C. Chela
D. Chelicera.
E. Patella.
F. First mesosomal segment.
G. Fifth metasomal segment.
H. Fourth metasomal segment.

Discussion

Euscorpius Thorell, 1876 is the most characteristic genus for Europe, whose species have otherwise spread only to the northernmost Africa (near the Mediterranean coast) and the Caucasus. Unfortunately the subspecific taxonomy of *Euscorpius* is not entirely clear. The number of hitherto described subspecies is rather large, and some of them do not appear to be justified by geographic distribution. This is true especially for *E. carpathicus* that includes 24 subspecies, of which 16 have been recorded from Italy. A revision of the entire genus is much needed in my opinion. Apart from the subspecies, the status of *E. (E.) mesotrichus* Hadži, 1929 that is usually regarded as a synonym of *E. carpathicus* also needs to be ascertained. For this reason *E. mesotrichus* is not included, although a future revision may well show this species to be valid.

Czech authors have frequently stated that the population of *E. carpathicus* found at an isolated locality near Slapy, central Bohemia, is not autochthonous but introduced from e. g. Bulgaria. Although specimens from this population have been studied by Max Vachon and Viktor Fet, their subspecific affiliation remains to be determined. Viktor Fet (in litt.) agrees with my opinion that the population of *E. carpathicus* at Slapy appears to belong to the same subspecies as the population at the nearest locality in Austria, which favors the thesis of autochthony. At any rate, the possibility of introduction from Bulgaria can be unequivocally rejected.

List of species and subspecies of the genus *Euscorpius* Thorell, 1876

Euscorpius (Euscorpius) carpathicus (Linné, 1767)

- E. (E.) c. aegaeus* Caporiacco, 1950
- E. (E.) c. apuanus* Caporiacco, 1950
- E. (E.) c. aquileiensis* (C. L. Koch, 1837)
- E. (E.) c. argentarii* Caporiacco, 1950
- E. (E.) c. balearicus* Caporiacco, 1950
- E. (E.) c. calabriae* Caporiacco, 1950
- E. (E.) c. candiota* Birula, 1903
- E. (E.) c. canestrinii* Fanzago, 1872
- E. (E.) c. carpathicus* (Linné, 1767)
- E. (E.) c. concinnus* (C.L.Koch, 1837)
- E. (E.) c. corsicanus* Caporiacco, 1950
- E. (E.) c. garganicus* Caporiacco, 1950

- E. (E.) c. hadzii* Caporiacco, 1950
E. (E.) c. ilvanus Caporiacco, 1950
E. (E.) c. lagostae Caporiacco, 1950
E. (E.) c. linosae Caporiacco, 1950
E. (E.) c. niciensis (C. L. Koch, 1841)
E. (E.) c. oglasae Caporiacco, 1950
E. (E.) c. ossae Caporiacco, 1950
E. (E.) c. palmarolae Caporiacco, 1950
E. (E.) c. picenus Caporiacco, 1950
E. (E.) c. sicanus (C. L. Koch, 1837)
E. (E.) c. tauricus (C. L. Koch, 1837)
E. (E.) c. tergestinus (C. L. Koch, 1837)
***E. (Euscorpius) germanus* (C. L. Koch, 1837)**
E. (E.) g. alpha Caporiacco, 1950
E. (E.) g. croaticus Caporiacco, 1950
E. (E.) g. germanus (C. L. Koch, 1837)
E. (E.) g. marcuzzii Valle, Berizzi, Bonino, Gorio, Gimmillaro-Negri & Percassi, 1971
***E. (Euscorpius) mingrelicus* (Kessler, 1876)**
E. (E.) m. caporiaccoi Bonacina, 1980
E. (E.) m. ciliciensis Birula, 1898
E. (E.) m. gamma Caporiacco, 1950
E. (E.) m. histrorum Caporiacco, 1950
E. (E.) m. legrandi Lacroix, 1995
E. (E.) m. mingrelicus (Kessler, 1876)
E. (E.) m. ollivieri Lacroix, 1995
E. (E.) m. phrygius Bonacina, 1980
E. (E.) m. uludagensis Lacroix, 1995
***E. (Polytrichobothrius) italicus* (Herbst, 1800)**
E. (P.) i. awhasicus (Nordmann, 1840)
E. (P.) i. etrusciae Caporiacco, 1950
E. (P.) i. italicus Herbst, 1800
E. (P.) i. oligotrichus Hadži, 1929
E. (P.) i. polytrichus Hadži, 1929
E. (P.) i. zakynthi Caporiacco, 1950
***E. (Tetratrichobothrius) flavicaudis* (De Geer, 1778)**
E. (T.) f. algeriacus (C. L. Koch, 1838)
E. (T.) f. cereris Rivellini, 1986
E. (T.) f. galitae Caporiacco, 1950
E. (T.) f. flavicaudis (De Geer, 1778)
E. (T.) f. massiliensis (C. L. Koch, 1837)

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Catalogue and Bibliography of Family Hersiliidae 1825-1998 (Arachnida: Araneida)

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Introduction

Among the 106 families of spiders, Hersiliidae is one of the interesting families in shape and behaviour. It is firstly noticed by Savigny (Audouin, 1825) who described *Hersilia caudata*. Baehr & Baehr (1987) began the recent taxonomic work on this family. Their continuous works provide us with very important information about the taxonomy, phylogeny, and zoogeography of hersiliids.

This work is a grouping of the data included in the following catalogues (using the same abbreviations) :

Roewer, C.F. 1942

Katalog der Araneae von 1758 bis 1940.

1.Band 1040 pp. Bremen.

[Family Hersiliidae: pp. 381-384]

Brignoli, P.M. 1983

A Catalogue of the Araneae described between 1940 and 1981.

Ed. P.Merrett. 755 pp. Manchester University Press.

(In association with The British Arachnological Society)

[Family Hersiliidae: pp. 430-432]

Platnick, N.I. 1989

Advances in Spider Taxonomy 1981-1987.

Ed. P.Merrett 673 pp. Manchester University Press.

(In association with The British Arachnological Society)

[Family Hersiliidae: pp. 174-176]

Platnick, N.I. 1993

Advances in Spider Taxonomy 1988-1991,

with Synonymies and Transfers 1940-1980.

Ed. P.Merrett 846 pp. New York Entomological Society.

(In association with The American Museum of Natural History)

[Family Hersiliidae: pp. 161-163]

Platnick, N.I. 1997

Advances in Spider Taxonomy 1992-1995,

with Redescriptions 1940-1980.

Ed. P.Merrett 976 pp. New York Entomological Society.

(In association with The American Museum of Natural History)

[Family Hersiliidae: pp. 235-240]

The author is much indebted to Dr. Barbara Baehr for her notices on this work and for her references. Rana, the author's wife helped with typing and correcting several times. Mr. Mohammed A. Mohafez who is studying the biology and behaviour of *Hersilia caudata*, activated me to achieve this work.

=====

Family **Hersiliidae** : 7 genera, 146 species.

Distribution: Tropical and Subtropical.

Gen. **Hersilia** Savigny, 1825 [56 species]

West Africa to Australia

Gen. **Hersiliola** Thorell, 1870 [9 species]

Mediterranean to Central Asia, Nigeria, South Africa

Gen. **Murricia** Simon, 1882 [3 species]

India, Sri Lanka, Singapore

Gen. **Neotama** Baehr & Baehr, 1993 [4 species]

India, Sri Lanka, Java, Sumatra

Gen. **Promurricia** Baehr & Baehr, 1993 [1 species]

Sri Lanka

Gen. **Tama** Simon, 1882 [24 species]

Central and South America, South Africa, Mediterranean, Sri Lanka

Gen. **Tamopsis** Baehr & Baehr, 1987 [49 species]
Australia, New Guinea

=====

CATALOGUE OF GENERA AND SPECIES

Family **HERSILIIDAE**

Gen. **Hersilia** Savigny, 1825 [56 species]

♂♀ **H. albicomis** Simon, 1887 West, Central Africa

H. a. Simon, 1887a: 273 (D ♀).

H. a. Simon, 1893: 443, f. 425 (N ♂).

H. decellei Benoit, 1967: 17, f. 3, 18 (D ♀).

H. a. Benoit, 1967: 26, f. 23, 28, 31 (♂♀).

H. a. Benoit, 1971: 153 (S).

♀ **H. albinota** Baehr & Baehr, 1993 China

H. a. Baehr & Baehr, 1993: 60, f. 9, 39a-b, e-f (D ♀).

♂♀ **H. albomaculata** Wang & Yin, 1985 China

H. a. Wang & Yin, 1985: 47, f. 2A-D (D ♀).

H. a. Song & Chen, 1985: 445, f. 1-2 (D ♂).

H. a. Song, 1987: 114, f. 77 (♂♀).

H. a. Chen & Zhang, 1991: 79, f. 70.1-4 (♂♀).

H. a. Baehr & Baehr, 1993: 19, f. 2, 16a-d (♂).

H. a. Baehr, 1998: 62, f. 1e (♂).

♂♀ **H. alluaudi** Berland, 1919 Central, East Africa

H. a. Berland, 1919: 348, f. 6, 7 (D ♂).

H. a. Berland, 1920: 123, f. 144, 145 (D ♂).

H. a. Benoit, 1967: 22, f. 5, 14-15, 20 (♂, D ♀).

♂♀ **H. arborea** Lawrence, 1928 Namibia

H. a. Lawrence, 1928: 239, pl. 21, f. 23, 24 (♂♀).

H. a. Smithers, 1945: 6, f. 5f, 6a (♂♀).

H. a. Benoit, 1967: 24, f. 16, 21 (♂♀).

♂♀ **H. asiatica** Song & Zheng, 1982 China, Taiwan, Thailand

H. a. Song & Zheng, 1982: 40, f. 1-5 (D ♂♀).

H. a. Hu, 1984: 81, f. 74.1-5 (♂♀).

H. a. Song, 1987: 116, f. 78 (♂♀).

- H. a.* Feng, 1990: 48, f. 23.1-6 (♂♀).
H. a. Chen & Zhang, 1991: 78, f. 69.1-5 (♂♀).
H. a. Baehr & Baehr, 1993: 25, f. 20c-f (♂♀).
H. a. Chen, 1994: 1, f. 1A-F (♂♀).

- ♂♀ ***H. australiensis*** Baehr & Baehr, 1987 Northern Territory
H. a. Baehr & Baehr, 1987: 354, f. 1-2 (D ♂♀). (Australia)
H. a. Baehr, 1998: 62, f. 1f,i (♂♀).

- ♂♀ ***H. baforti*** Benoit, 1967 Zaire (Congo)
H. b. Benoit, 1967: 19, f. 4, 11-12, 19 (D ♂♀).

- ♂♀ ***H. baliensis*** Baehr & Baehr, 1993 Bali
H. b. Baehr & Baehr, 1993: 42, f. 28c-f (D ♂♀).

- ♂ ***H. bifurcata*** Baehr & Baehr, 1998 Northern Territory
H. b. Baehr & Baehr, 1998: 14, f. 1,2,27 (D ♂).

- ♂♀ ***H. caudata*** Savigny, 1825 {T} West Africa to China
H. c. Audouin, 1825: 115, pl. 1, f. 8 (D ♀).
H. c. Audouin, 1827: 318, pl. 1, f. 8 (D ♀).
H. c. Walckenaer, 1837: 371 (D ♀).
H. c. C. L. Koch, 1843: 103, f. 824 (D ♀).
H. c. var diversa O. P.-Cambridge, 1876: 560, pl. 58, f. 6 (N).
H. c. Simon, 1893: 446, f. 418-423 (N).
H. c. Kulczyński, 1901: 18, pl. 1, f. 18, 19 (D ♂♀).
H. hirtiventris Benoit, 1967: 23, f. 6-7 (D ♀).
H. c. Benoit, 1967: 34, f. 37, 40, 44 (♂♀, S).
H. c. Benoit, 1971: 152 (S).
H. c. Baehr & Baehr, 1993: 17, f. 1, 15a-f (♂♀).
H. c. Baehr, 1998: 62, f. 1d (♂).

- ♂ ***H. clarki*** Benoit, 1967 Zambia
H. c. Benoit, 1967: 20, f. 13 (D ♂).

- ♀ ***H. clypealis*** Baehr & Baehr, 1993 Thailand
H. c. Baehr & Baehr, 1993: 45, f. 30e-f (D ♀).

- ♀ ***H. corticola*** Lawrence, 1937 South Africa
H. c. Lawrence, 1937: 226, f. 7 (D ♀).
H. c. Lawrence, 1938: 481 (C ♀).
H. c. Smithers, 1945: 5, f. 5d (♀).

- H. c.* Benoit, 1967: 15, f. 1 (♀).
- ♂♀ ***H. deelemanae*** Baehr & Baehr, 1993 Sumatra
H. d. Baehr & Baehr, 1993: 27, f. 4, 21a-f (D ♂♀).
- ♀ ***H. facialis*** Baehr & Baehr, 1993 Sumatra
H. f. Baehr & Baehr, 1993: 61, f. 10, 40a-b, e-f (D ♀).
- ♂ ***H. feai*** Baehr & Baehr, 1993 Myanmar
H. f. Baehr & Baehr, 1993: 36, f. 25c-d, g (D ♂).
- ♂♀ ***H. flagellifera*** Baehr & Baehr, 1993 Sumatra
H. f. Baehr & Baehr, 1993: 46, f. 6, 31a-f (D ♂♀).
- ♀ ***H. fossulata*** Karsch, 1881 Madagascar
H. f. Karsch, 1881: 195 (D ♀).
- ♀ ***H. hildebrandti*** Karsch, 1878 Zanzibar
H. h. Karsch, 1878: 313, pl. 8, f. 2 (D ♀).
H. h. Benoit, 1967: 31, f. 26 (♀).
- ♀ ***H. impressifrons*** Baehr & Baehr, 1993 Borneo
H. i. Baehr & Baehr, 1993: 57, f. 37e-f (D ♀).
- ♂♀ ***H. incompta*** Benoit, 1971 Ivory Coast
H. i. Benoit, 1971: 154, f. 2, 6 (D ♂♀).
- ♂♀ ***H. insulana*** Strand, 1907 Madagascar
H. i. Strand, 1907a: 728 (D ♂♀).
H. i. Strand, 1907c: 26 (D ♂♀).
- ♀ ***H. kauderni*** Strand, 1908 Madagascar
H. k. Strand, 1908: 457 (D ♀).
- ♂♀ ***H. kinabaluensis*** Baehr & Baehr, 1993 Borneo
H. k. Baehr & Baehr, 1993: 23, f. 3, 19a-f (D ♂♀).
- ♂♀ ***H. longbottomi*** Baehr & Baehr, 1998 Northwestern Australia
H. b. Baehr & Baehr, 1998: 18, f. 7-10, 27 (D ♂♀).
- ♂ ***H. madang*** Baehr & Baehr, 1993 New Guinea
H. m. Baehr & Baehr, 1993: 53, f. 8, 35a-d (D ♂).

- ♀ **H. mainae** Baehr & Baehr, 1995 Western Australia
H. m. Baehr & Baehr, 1995: 107, f. 1a-e (D ♀).
- ♂ **H. martensi** Baehr & Baehr, 1993 Nepal
H. m. Baehr & Baehr, 1993: 21, f. 17c-d (D ♂).
- ♂♀ **H. mimbi** Baehr & Baehr, 1993 Western Australia
H. m. Baehr & Baehr, 1993: 349, f. 1-4 (D ♂♀).
H. m. Baehr, 1998: 62, f. 1g,j (♂♀).
- ♀ **H. mjoebergi** Baehr & Baehr, 1993 Sumatra
H. m. Baehr & Baehr, 1993: 44, f. 29e-f (D ♀).
- ♂♀ **H. nentwigi** Baehr & Baehr, 1993 Java, Sumatra, Krakatau
H. n. Baehr & Baehr, 1993: 40, f. 27c-d (D ♂♀).
- ♂♀ **H. nepalensis** Baehr & Baehr, 1993 Nepal
H. n. Baehr & Baehr, 1993: 34, f. 24c-d, f-g (D ♂♀).
- ♀ **H. nossibeensis** Strand, 1915 Nossibé ?
H. n. Strand, 1915: 55 (D ♀).
- ♂♀ **H. novaeguineae** Baehr & Baehr, 1993 New Guinea
H. n. Baehr & Baehr, 1993: 55, f. 36c-f (D ♂♀).
- ♂♀ **H. occidentalis** Simon, 1907 West, Central Africa, Principe
H. o. Simon, 1907: 248 (D ♂).
H. brevimammillata Strand, 1913a: 339 (D ♀).
H. o. Benoit, 1967: 30, f. 25, 30, 34-35 (♂, S ♀).
- ♂ **H. pectinata** Thorell, 1895 Myanmar, Borneo, Philippines
H. p. Thorell, 1895: 58 (D ♂).
H. p. Pocock, 1900: 241 (D ♂, nec ♀).
H. p. Baehr & Baehr, 1993: 48, f. 32c-d (♂ ; ♀ of Sinha, 1951 =
H. tibialis).
H. p. Baehr, 1998: 62, f. 1a.
- ♂♀ **H. pungwensis** Tucker, 1920 South Africa
H. p. Tucker, 1920: 475, pl. 29, f. 11 (D ♂).
H. p. Smithers, 1945: 6, f. 5e, 6c (♂, D ♀).
H. p. Benoit, 1967: 36, f. 38, 41-42, 45 (♂♀).

♂♀ **H. savignyi** Lucas, 1836 Sri Lanka, India to Myanmar, Philippines

H. s. Lucas, 1836: 8, pl. 13, f. 1 (D ♀).

H. indica Walckenaer, 1837: 372 (D part.).

H. calcuttensis Stoliczka, 1869: 216, pl. 20, f. 9 (D ♀).

H. s. Simon, 1885: 19, pl. 10, f. 18, 19 (N ♂).

H. s. Thorell, 1887: 80 (D ♂♀).

H. s. Simon, 1893: 414, f. 417, 426 (N ♂♀).

H. clathrata Thorell, 1895: 56 (D ♀).

H. s. Pocock, 1900: 241, f. 82 (D ♀).

H. clathrata Pocock, 1900: 242 (D ♀).

H. s. Gravely, 1922: 1050, pl. 5, f. 13 (N).

H. s. Benoit, 1974: 995, f. 1, 4 (♀).

H. s. Tikader & Biswas, 1981: 47, f. 74-76 (♀).

H. clathrata Yaginuma & Wen, 1983: 193, f. 2A-B (♀).

H. clathrata Xu, 1984: 25, f. 1-4 (♀, D ♂).

H. s. Baehr & Baehr, 1993: 29, f. 5, 22a-f (♂♀, S).

H. clathrata Barrion & Litsinger, 1995: 411, f. 246a-k (♂♀).

♀ **H. segregata** Benoit, 1967 Kenya, Tanzania

H. s. Benoit, 1967: 24, f. 22, 27 (D ♀).

♂♀ **H. sericea** Pocock, 1898 South Africa

H. s. Pocock, 1898: 214, pl. 8, f. 9 (D ♀).

H. bicornis Tucker, 1920: 473, pl. 29, f. 10 (D ♂♀).

H. s. Lawrence, 1937: 228 (C ♀).

H. s. Smithers, 1945: 5, f. 5c (♀).

H. bicornis Smithers, 1945: 5, f. 5b, 6d (♂♀).

H. hanströmi Kauri, 1950: 8, f. 5 (D ♀).

H. s. Benoit, 1967: 27, f. 24, 29, 32-33 (♀, S ♂).

♂♀ **H. setifrons** Lawrence, 1928 Namibia

H. s. Lawrence, 1928: 241, pl. 21, f. 25 (D ♂♀).

H. s. Smithers, 1945: 7, f. 5a, 6b (♂♀).

H. s. Benoit, 1967: 32, f. 36, 39, 43 (♂♀).

♂♀ **H. sigillata** Benoit, 1967 Ivory Coast, Zaire (Congo)

H. s. Benoit, 1967: 15, f. 2, 8-10, 17 (D ♂♀).

♂♀ **H. simplicipalpis** Baehr & Baehr, 1993 Thailand

H. s. Baehr & Baehr, 1993: 32, f. 23c-g (D ♂♀).

- ♂♀ **H. striata** Wang & Yin, 1985 China, Myanmar, Thailand, Java,
H. s. Wang & Yin, 1985: 45, f. 1A-E (D ♂♀). Sumatra
H. s. Song, 1987: 117, f. 79 (♂♀).
H. s. Baehr & Baehr, 1993: 37, f. 26c-g (♂♀).
- ♀ **H. stumptfi** Strand, 1915 Nossibé ?
H. s. Strand, 1915: 57 (D ♀).
- ♀ **H. sumatrana** (Thorell, 1890) Malaysia, Sumatra, Borneo
Chalinura s. Thorell, 1890a: 319 (D ♂).
Hersilia s. Thorell, 1890b: 8 (D ♀).
Tama s. Simon, 1893: 445 (N).
Hersilia s. (? = *ceylonica*) Strand, 1907b: 18 (D ♀).
H. stevensi Sinha, 1951: 123, f. 1 (D ♀).
H. s. Baehr & Baehr, 1993: 50, f. 33e-f (T ♀ from Tama, S).
- ♂♀ **H. sundaica** Baehr & Baehr, 1993 Lambok, Sumbawa (Indonesia)
H. s. Baehr & Baehr, 1993: 58, f. 38c-f (D ♂♀).
- ♂♀ **H. tenuifurcata** Baehr & Baehr, 1998 Northwestern Australia
H. b. Baehr & Baehr, 1998: 20, f. 11-14,27 (D ♂♀).
- ♂♀ **H. tibialis** Baehr & Baehr, 1993 India, Sri Lanka
H. pectinata Sinha, 1951: 123, f. 2 (♀, misidentified).
H. t. Baehr & Baehr, 1993: 51, f. 7, 34a-f (D ♂♀).
- ♂♀ **H. vanmoli** Benoit, 1971 Ivory Coast, Togo
H. v. Benoit, 1971: 156, f. 3-5, 7 (D ♂♀).
- ♀ **H. vicina** Baehr & Baehr, 1993 Thailand
H. v. Baehr & Baehr, 1993: 22, f. 18e-f (D ♀).
- ♀ **H. vinsoni** Lucas, 1869 Madagascar
H. v. Lucas, 1869: 160, pl. 11, f. 1-5 (D ♀).
- ♂♀ **H. wellswebberae** Baehr & Baehr, 1998 Northern Territory
H. b. Baehr & Baehr, 1998: 17, f. 3-6,27 (D ♂♀).
- ♂♀ **H. xinjiangensis** Liang & Wang, 1989 China
H. x. Liang & Wang, 1989: 56, f. 1-4 (D ♂♀).
H. x. Hu & Wu, 1989: 78, f. 55.5-8 (♂♀).

- ♀ **H. yunnanensis** Wang, Song & Qiu, 1993 China
H. y. Wang, Song & Qiu, 1993: 33, f. 1-3 (D ♀).

Nomina dubia:

Baehr & Baehr, 1993: 77:

- ♀ *H. celebensis* Thorell, 1877: 472 (D ♀) Celebes, Sumatra, Amboina
 ♀ *H. fletcheri* Sinha, 1951: 125, f. 3a (D ♀) Burma
 ♂ *H. kalimpongensis* Sinha, 1951: 124 (D ♂) India
 ♀ *H. moulmeinensis* Sinha, 1951: 124, f. 3 b (D ♀) Burma

Baehr & Baehr, 1993: 78:

- ♀ *H. peguana* Thorell, 1895: 60 (D ♀) Burma
 ♀ *H. pernix* Kulczyński, 1911: 433, pl. 19, f. 10 (D ♀) New Guinea
 ♀ *H. siamensis* Simon, 1886: 156 (D ♀) Siam
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Gen. **Hersiliola** Thorell, 1870 [9 species]

- ♀ **H. afghanica** Roewer, 1960 Afghanistan
H. a. Roewer, 1960: 48, f. 16 a-d (D ♀).

- ♂♀ **H. australis** Simon, 1893 South Africa
H. a. Simon, 1893: 447 (D ♀).
H. a. Tucker, 1920: 472, pl. 29, f. 9 (D ♂♀).
H. a. Smithers, 1945: 15, f. 10a, 11a-c, 12a-b (♂♀).

- ♂♀ **H. brachyplura** Strand, 1913 Palestine
H. b. Strand, 1913b: 148 (D ♂).
H. b. demaculata Strand, 1914: 182 (D ♀).

- ♀ **H. fragilis** Lawrence, 1928 Namibia
H. f. Lawrence, 1928: 242, pl. 21, f. 26 (D ♀).
H. f. Smithers, 1945: 18, f. 10b (♀).

- ♂o **H. lucasi** (O. P.-Cambridge, 1876) Egypt, Libya
Hersilidia l. O. P.-Cambridge, 1876: 562, pl. 58, f. 5 (D ♂♀).
H. maculata Simon, 1908a: 42 (N).
H. l. Wiehle, 1960: 470, f. 15 (♀).

- ♂♀ **H. macullulata** (Dufour, 1831) {T} Mediterranean to Turkmenistan
Aranea m. Dufour, 1831: 360, pl. 10, f. 2 (D ♀).
Hersilia oraniensis Lucas, 1846: 129, pl. 4, f. 8 (D ♂♀).
Hersilia oraniensis Lucas, 1868: 41 (N).

Hersilidia oraniensis Simon, 1870: 347 (79) (N).

H. m. Simon, 1882: 256 (N).

H. m. Simon, 1893: 447, f. 424 (N).

H. maculata Benoit, 1974: 997, f. 2, 5 (♀).

H. maculata Ribera, Ferrández & Pérez, 1988: 98, f. 1, 4, 7-8 (♂♀).

♀ **H. pallida** Kroneberg, 1875 Turkestan, Karakorum (Mongolia)

H. p. Kroneberg, 1875: 13, pl. 5, f. 41 (D ♀).

H. p. Simon, 1893: 445 (N).

♂♀ **H. simoni** (O. P.-Cambridge, 1872) Mediterranean, Nigeria, Cape Verde Is.

Hersiliada s. O. P.-Cambridge, 1872: 275, pl. 14, f. 9 (D ♂♀).

H. s. Simon, 1893: 445 (N).

H. s. Denis, 1955: 128, f. 29 (♀).

H. s. Benoit, 1974: 997, f. 3, 6 (♀).

H. s. Ribera, Ferrández & Pérez, 1988: 99, f. 2, 5, 9-10 (♂♀).

H. s. Schmidt & Krause, 1995: 356, f. 1-2 (♀).

♀ **H. versicolor** (Blackwall, 1865) Cape Verde Is.

Hersilia v. Blackwall, 1865: 81 (D ♀).

H. v. Simon 1893: 445 (N).

Gen. **Murricia** Simon, 1882 [3 species]

♀ **M. cornuta** Baehr & Baehr, 1993 {T} Singapore

M. c. Baehr & Baehr, 1993: 65, f. 12, 42a-b, e-f (D ♀).

♀ **M. crinifera** Baehr & Baehr, 1993 Sri Lanka

M. c. Baehr & Baehr, 1993: 66, f. 43e-f (D ♀).

♀ **M. triangularis** Baehr & Baehr, 1993 India

M. t. Baehr & Baehr, 1993: 67, f. 44e-f (D ♀).

Nomen dubium:

Baehr & Baehr, 1993: 78:

♂♀ *M. indica* (Lucas, 1836): 7, pl. 13, f. 2 (D ♂) India, Sumatra

Gen. **Neotama** Baehr & Baehr, 1993 [4 species]

♂♀ **N. longimana** Baehr & Baehr, 1993 Java, Sumatra

N. l. Baehr & Baehr, 1993: 71, f. 13. 46a-f (D ♂♀).

N. l. Baehr, 1998: 62, f. 1b.

♀ **N. punctigera** Baehr & Baehr, 1993 India

N. p. Baehr & Baehr, 1993: 73, f. 47e-f (D ♀).

♀ **N. rothorum** Baehr & Baehr, 1993 India

N. r. Baehr & Baehr, 1993: 74, f. 48e-f (D ♀).

♂♀ **N. variata** (Pocock, 1899) {T} Sri Lanka

N. v. Baehr & Baehr, 1993: 69, f. 45c-f (T ♂♀ from Tama).

N. v. Baehr, 1998: 62, f. 1c (♂).

Gen. **Promurricia** Baehr & Baehr, 1993 [1 species]

♀ **P. depressa** Baehr & Baehr, 1993 {T} Sri Lanka

P. d. Baehr & Baehr, 1993: 63, f. 11, 41a-b, e-f (D ♀).

Gen. **Tama** Simon, 1882 [24 species]

♀ **T. albigastra** Mello-Leitão, 1928 Brazil

T. a. Mello-Leitão, 1928: 51 (D ♀).

o **T. americana** (Simon, 1887) Paraguay

Rhadine a. Simon, 1887b: 176 (D o).

T. a. Simon, 1893: 445 (N).

♂ **T. argentina** Mello-Leitão, 1942 Argentina

T. a. Mello-Leitão, 1942: 398, f. 14-15 (D ♂).

♂♀ **T. arida** Smithers, 1945 South Africa

T. a. Smithers, 1945: 10, f. 1-2, 7b, 8 (D ♂♀).

♀ **T. bicava** Smithers, 1945 Namibia

T. b. Smithers, 1945: 14, f. 7d, 9b (D ♀).

- T. brasiliensis** Piza, 1937 Brazil
T. b. Piza, 1937: 14, f. 1 (D).
- ♂♀ **T. catamarcaensis** Carcavallo, 1959 Argentina
T. c. Carcavallo, 1959: 127, f. 1-2 (D ♂♀).
- ♂♀ **T. crucifera** Vellard, 1924 Brazil
T. c. Vellard, 1924: 139, pl. 14, f. 51 (D ♂♀).
- ♂ **T. crulsi** Mello-Leitão, 1930 Amazonas
T. c. Mello-Leitão, 1930: 58, f. 8-10 (D ♂).
- ♂♀ **T. edwardsi** (Lucas, 1846) {T} Spain, Portugal, Algeria
Hersilia e. Lucas, 1846: 128, pl. 4, f. 7 (D ♂♀).
T. e. Simon, 1882: 256 (N).
T. e. Simon, 1893: 446 (N).
T. e. Ribera, Ferrández & Pérez, 1988: 100, f. 3, 6, 11 (♂♀).
- ♂ **T. forcipata** F. O. P.-Cambridge, 1902 Guatemala, Mexico
T. f. F.O.P.-Cambridge, 1902: 351, pl. 33, f. 7 (D ♂).
T. f. Schawaller, 1981: 8, f. 7-8 (♂).
- ♂ **T. guyanensis** Mello-Leitão, 1948 Guyana
T. g. Mello-Leitão, 1948: 155, f. 3 (D ♂).
- ♀ **T. habanensis** Franganillo Balboa, 1935 Cuba
T. h. Franganillo Balboa, 1935: f. 35 (D ♀).
T. h. Franganillo Balboa, 1936: 39, f. 17 (D ♀).
T. h. Bryant, 1940: 274, f. 11, 17 (D ♀).
- ♀ **T. incerta** Tucker, 1920 South Africa
T. i. Tucker, 1920: 476 (D ♀).
T. i. Smithers, 1945: 9, F. 7A (♀).
- ♂ **T. karinae** Carcavallo, 1961 Argentina
T. k. Carcavallo, 1961: 61, f. 1-2 (D ♂).
- ♂ **T. longipes** Carcavallo, 1961 Bolivia
T. l. Carcavallo, 1961: 63, f. 3-6 (D ♂).
- ♂♀ **T. mexicana** (O. P.-Cambridge, 1893) Mexico to Panama
Hersilia m. O. P.-Cambridge, 1893: 107, pl. 14, f. 6, 7 (D ♂♀).

T. m. Banks, 1898: 211 (C).

T. m. F.O.P.-Cambridge, 1902: 351, pl. 33, f. 5, 6 (C ♂♀).

T. m. Wunderlich, 1988: 90, f. 169-171 (♂).

♀ ***T. micrura*** Mello-Leitão, 1928 Pernambuco (Brazil)

T. m. Mello-Leitão, 1928: 52 (D ♀).

♀ ***T. obscura*** Smithers, 1945 South Africa

T. o. Smithers, 1945: 12, f. 7c, 9a (D ♀).

♀ ***T. occidentalis*** Schenkel, 1953 Venezuela

T. o. Schenkel, 1953: 9, f. 9 (D ♀).

♀ ***T. pachyura*** Mello-Leitão, 1935 South Brazil

T. p. Mello-Leitão, 1935: 363, f. 1, 2 (D ♀).

♀ ***T. sasaimae*** Mello-Leitão, 1941 Colombia

T. s. Mello-Leitão, 1941: 244 (D ♀).

♂♀ ***T. variata*** Pocock, 1899 Sri Lanka

T. v. Pocock, 1899: 751 (D ♀).

T. v. Pocock, 1900: 242 (D ♂♀).

♀ ***T. vittata*** (Simon, 1887) Venezuela

Rhadine v. Simon, 1887b: 176 (D ♀).

T. v. Simon, 1893: 445 (N).

Nomina dubia:

Baehr & Baehr, 1987: 391:

o *T. brachyura* Simon, 1908b: 406 (D o) Western Australia

♀ *T. novae-hollandiae* (L. Koch, 1876): 828, pl. 71, f. 1 (D ♀) New South Wales, Queensland

Baehr & Baehr, 1993: 79:

♀ *T. gravelyi* Sinha, 1951: 126, f. 3c (D ♀) India

Gen. ***Tamopsis*** Baehr & Baehr, 1987 [49 species]

♂ ***T. amplithorax*** Baehr & Baehr, 1987 Western Australia

T. a. Baehr & Baehr, 1987: 360, f. 3 (D ♂).

- ♂♀ **T. arnhemensis** Baehr & Baehr, 1987 Northern Territory,
T. a. Baehr & Baehr, 1987: 376, f. 27-28 (D ♂♀). Queensland
T. a. Baehr, 1988: 241, f. 3, 8 (♂).
- ♂♀ **T. brachycauda** Baehr & Baehr, 1987 Queensland, New South
T. b. Baehr & Baehr, 1987: 361, f. 6-7 (D ♂♀). Wales
- ♀ **T. brevipes** Baehr & Baehr, 1987 New South Wales
T. b. Baehr & Baehr, 1987: 375, f. 26 (D ♀).
- ♂♀ **T. brisbanensis** Baehr & Baehr, 1987 Queensland, New South
T. b. Baehr & Baehr, 1987: 365, f. 12-13 (D ♂♀). Wales
- ♂ **T. centralis** Baehr & Baehr, 1987 Queensland
T. c. Baehr & Baehr, 1987: 369, f. 17 (D ♂).
- ♂♀ **T. circumvidens** Baehr & Baehr, 1987 Western Australia
T. c. Baehr & Baehr, 1987: 378, f. 29-30 (D ♂♀).
T. c. Baehr, 1988: 241, f. 1, 9 (♂).
T. c. Baehr & Baehr, 1998: 29 (N).
- ♀ **T. cooloolensis** Baehr & Baehr, 1987 Queensland
T. c. Baehr & Baehr, 1987: 375, f. 25 (D ♀).
- ♀ **T. darlingtoniana** Baehr & Baehr, 1987 Western Australia
T. d. Baehr & Baehr, 1987: 371, f. 20 (D ♀).
- ♂♀ **T. daviesae** Baehr & Baehr, 1987 Queensland
T. daviesi Baehr & Baehr, 1987: 367, f. 14-15 (D ♂♀).
- ♂ **T. depressa** Baehr & Baehr, 1992 Western Australia, Northern
T. d. Baehr & Baehr, 1992: 62, f. 1-3 (D ♂). Territory
- ♂♀ **T. ediacarae** Baehr & Baehr, 1988 South Australia
T. e. Baehr & Baehr, 1988: 15, f. 3a-e (D ♀).
T. e. Baehr & Baehr, 1998: 29, f. 23,24,29 (D ♂).
- ♂♀ **T. eucalypti** (Rainbow, 1900) {T} Queensland to South Australia
Tama e. Rainbow, 1899: 486, pl. 23, f. 2 (D ♂♀).
T. e. Baehr & Baehr, 1987: 364, f. 10-11 (T ♂♀ from Tama).

- ♂♀ **T. facialis** Baehr & Baehr, 1993 Western Australia, New South Wales
T. f. Baehr & Baehr, 1993: 365, f. 7-8 (D ♂).
T. triangularis Baehr & Baehr, 1993: 376, f. 17-18 (D ♀).
T. f. Baehr & Baehr, 1995: 110, f. 2 (♂).
T. f. Baehr & Baehr, 1998: 24, f. 28 (N, S).
- ♂♀ **T. fickerti** (L. Koch, 1876) New South Wales, Queensland
Chalinura f. L. Koch, 1876: 830, pl. 71, f. 2 (D ♀).
Rhadine f. Simon, 1882: 255 (N).
T. f. Baehr & Baehr, 1987: 385, f. 37-38 (T ♀ from Tama, D ♂).
T. f. Baehr, 1988: 241, f. 14 (♂).
- ♂♀ **T. fitzroyensis** Baehr & Baehr, 1987 Western Australia
T. f. Baehr & Baehr, 1987: 389, f. 43-44 (D ♂♀).
T. f. Baehr, 1988: 241, f. 7, 19 (♂).
- ♂♀ **T. forrestae** Baehr & Baehr, 1988 Queensland
T. forresti Baehr & Baehr, 1988: 14, f. 1a-e, 2a-e (D ♂♀).
T. forresti Baehr, 1988: 241, f. 5, 13 (♂).
T. f. Baehr & Baehr, 1995: 110 (emendation of incorrect patronym).
- ♀ **T. gibbosa** Baehr & Baehr, 1993 Western & South Australia
T. g. Baehr & Baehr, 1993: 372, f. 13-14 (D ♀).
T. g. Baehr & Baehr, 1998: 26, f. 27 (N).
- ♂ **T. gracilis** Baehr & Baehr, 1993 Western Australia
T. g. Baehr & Baehr, 1993: 379, f. 19-20 (D ♂).
T. g. Baehr & Baehr, 1998: 31, f. 29 (N).
- ♀ **T. grayi** Baehr & Baehr, 1987 New South Wales
T. g. Baehr & Baehr, 1987: 370, f. 19 (D ♀).
- ♀ **T. harveyi** Baehr & Baehr, 1993 Northern Territory
T. h. Baehr & Baehr, 1993: 370, f. 11-12 (D ♀).
- ♂ **T. hirsti** Baehr & Baehr, 1998 South Australia
T. h. Baehr & Baehr, 1998: 23, f. 15, 16, 27 (D ♂).
- ♂♀ **T. jongi** Baehr & Baehr, 1995 Western Australia
T. j. Baehr & Baehr, 1995: 113, f. 4a-d (D ♂♀).

- ♂ **T. kimberleyana** Baehr & Baehr, 1998 Northwestern Australia
T. k. Baehr & Baehr, 1998: 28, f. 21,22,29 (D ♂).
- ♂ **T. kochi** Baehr & Baehr, 1987 Western Australia
T. k. Baehr & Baehr, 1987: 368, f. 16 (D ♂).
- ♀ **T. leichhardtiana** Baehr & Baehr, 1987 Northern Territory,
T. l. Baehr & Baehr, 1987: 382, f. 35 (D ♀). Queensland
T. l. Baehr & Baehr, 1998: 31, f. 28 (N).
- ♂♀ **T. longbottomi** Baehr & Baehr, 1993 Northern Territory
T. l. Baehr & Baehr, 1993: 382, f. 21-24 (D ♂♀).
- ♀ **T. mainae** Baehr & Baehr, 1993 Western Australia
T. m. Baehr & Baehr, 1993: 374, f. 15-16 (D ♀).
- ♂ **T. mallee** Baehr & Baehr, 1989 Western & South Australia
T. m. Baehr & Baehr, 1989: 316, f. 7-8 (D ♂).
T. m. Baehr & Baehr, 1998: 31, f. 29 (N).
- ♂♀ **T. minor** Baehr & Baehr, 1998 Northwestern Australia
T. m. Baehr & Baehr, 1998: 24, f. 17-20,28 (D ♂♀).
- ♂ **T. nanutarrae** Baehr & Baehr, 1989 Western Australia
T. n. Baehr & Baehr, 1989: 310, f. 1-2 (D ♂).
- ♂♀ **T. occidentalis** Baehr & Baehr, 1987 Western Australia
T. o. Baehr & Baehr, 1987: 387, f. 41-42 (D ♂♀).
T. o. Baehr, 1988: 241, f. 16 (♂).
- ♂♀ **T. perthensis** Baehr & Baehr, 1987 Western Australia
T. p. Baehr & Baehr, 1987: 386, f. 39-40 (D ♂♀).
T. p. Baehr, 1988: 241, f. 15 (♂).
- ♂ **T. petricola** Baehr & Baehr, 1995 Queensland
T. p. Baehr & Baehr, 1995: 115, f. 5a-b (D ♂).
- ♀ **T. piankai** Baehr & Baehr, 1993 Western Australia
T. p. Baehr & Baehr, 1993: 368, f. 9-10 (D ♀).
- ♂♀ **T. platycephala** Baehr & Baehr, 1987 Queensland
T. p. Baehr & Baehr, 1987: 359, f. 4-5 (D ♂♀).

- ♂♀ **T. pseudocircumvidens** Baehr & Baehr, 1987 Northern Territory,
Western & South Australia
T. p. Baehr & Baehr, 1987: 381, f. 34 (D ♀).
T. marri Baehr & Baehr, 1989: 312, f. 3-6 (D ♂♀).
T. p. Baehr & Baehr, 1998: 31, f. 27 (N, S).
- ♂♀ **T. queenslandica** Baehr & Baehr, 1987 Queensland, New South
T. q. Baehr & Baehr, 1987: 372, f. 21-22 (D ♂♀). Wales
- ♂♀ **T. raveni** Baehr & Baehr, 1987 Queensland, South Australia
T. r. Baehr & Baehr, 1987: 373, f. 23-24 (D ♂♀).
T. r. Baehr & Baehr, 1998: 27 (N).
- ♂♀ **T. reevesbyana** Baehr & Baehr, 1987 Western & South Australia
T. r. Baehr & Baehr, 1987: 370, f. 18 (D ♀).
T. distinguenda Baehr & Baehr, 1992: 66, f. 4-8 (D ♂♀).
T. r. Baehr & Baehr, 1998: 27, f. 29 (N, S).
- ♀ **T. riverinae** Baehr & Baehr, 1993 New South Wales
T. r. Baehr & Baehr, 1993: 363, f. 5-6 (D ♀).
- ♀ **T. rossi** Baehr & Baehr, 1987 Western Australia
T. r. Baehr & Baehr, 1987: 383, f. 36 (D ♀).
- ♂ **T. transiens** Baehr & Baehr, 1992 Western Australia, Victoria,
T. t. Baehr & Baehr, 1992: 70, f. 9-11 (D ♂). Northern Territory
T. t. Baehr & Baehr, 1998: 27, f. 28 (N).
- ♂ **T. trionix** Baehr & Baehr, 1987 Queensland
T. t. Baehr & Baehr, 1987: 380, f. 33 (D ♂).
T. t. Baehr, 1988: 241, f. 12 (♂).
- ♂♀ **T. tropica** Baehr & Baehr, 1987 Northern Territory, Queensland
T. t. Baehr & Baehr, 1987: 379, f. 31-32 (D ♂♀).
T. t. Baehr, 1988: 241, f. 4, 11 (♂).
T. t. Baehr & Baehr, 1998: 29 (N).
- ♂♀ **T. tweedensis** Baehr & Baehr, 1987 Queensland, New South
T. t. Baehr & Baehr, 1987: 362, f. 8-9 (D ♂♀). Wales

♀ **T. warialda** Baehr & Baehr, 1998 New South Wales

T. w. Baehr & Baehr, 1998: 32, f. 25,26,28 (D ♀).

♀ **T. wau** Baehr & Baehr, 1993 New Guinea

T. w. Baehr & Baehr, 1993: 75, f. 14, 49a-b, e-f (D ♀).

♀ **T. weiri** Baehr & Baehr, 1995 Western Australia

T. w. Baehr & Baehr, 1995: 111, f. 3a-b (D ♀).

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Volume 6

Part 3

December, 1999

Cairo, Egypt

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Subscription for volume 6 (1998-1999) :

US \$ 25.00 (personal rate)

US \$ 35.00 (institutional rate)

Back issues :

Volume 1 (1987-1990), Volume 2 (1990-1992),

Volume 4 (1994-1996), Volume 5 (1996-1997):

US \$ 25.00 (p.r.) per volume

US \$ 35.00 (i.r.) per volume

Volume 3 (1992-1993):

US \$ 35.00 (p.r.), US \$ 45.00 (i.r.)

Correspondence concerning subscription, back issues, publication, etc. should be addressed to the editor.

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**Sun-spiders of the Arab countries
(Arachnida : Solpugida)**

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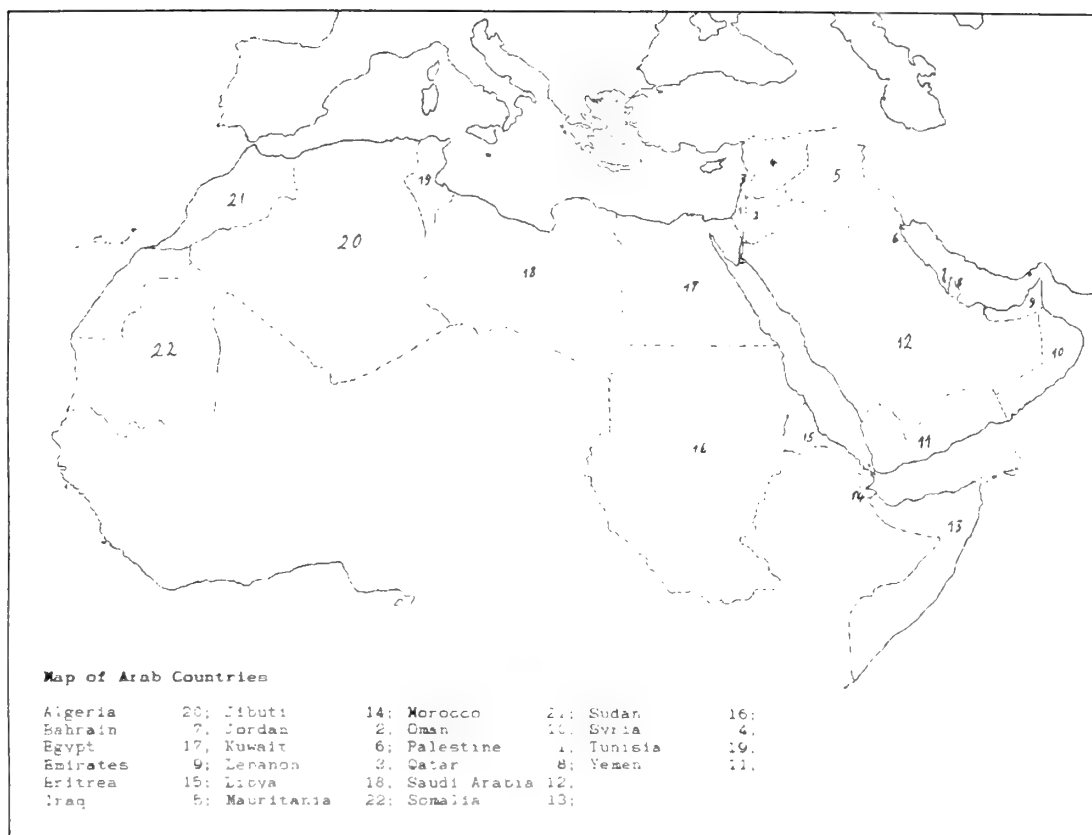
Introduction

Sun-spiders are scarcely studied in the Arab countries. Hence, this work is prepared to present a list of species recorded from these countries with their references and keys to families (from El-Hennawy, 1990) and genera (from Roewer, 1934 & 1941). This will be the base of more detailed studies similar to that of Egyptian species (El-Hennawy, 1998).

In this study, 191 species and 4 subspecies are recorded, classified within 57 genera in 6 families.

Family	Genera	Species	Sub-species
Daesiidae	12	50	4
Galeodidae	9	61	--
Gylippidae	2	5	--
Karschiidae	4	10	--
Rhagodidae	20	44	--
Solpugidae	10	21	--

Abbreviations: Ref = references, {T} = type species



Order Solpugida

Key to Solpugid Families

1. Anus : ventrally located

Family RHAGODIDAE

Tarsal segmentation : 1-1-1-1

Heavy-bodied; short-legged; small to large (10-60 mm)

Leg 1 : tarsi : with a pretarsus + 2 claws,

metatarsi : with a dense ventral clothing of short spinelike setae

Male cheliceral flagellum : paraxially immovable; composed of

2 flattened, curled, setae that form a nearly complete, slightly curved, truncate, hornlike tube on the mesial surface

Female genital opercula : not differentiated from other abdominal sternites and not specifically variable

Distribution : northeastern Africa, southwestern Asia, and Near East
[26 genera, 91 species]

- . Anus : terminally located

...2

2. Tarsal segmentation : 1-4-4-(6-7) **Family SOLPUGIDAE**

Long-legged; small to large (8-60 mm)

Leg 1 : tarsi : without claws

Male cheliceral flagellum : paraxially immovable; mesodorsal to dorsal, whiplike structure separated from the fixed cheliceral finger by a suture

Female genital opercula : indistinctly differentiated from other abdominal sternites, and although they are some-times variable from one genus to another, they are not specifically so

Distribution : predominantly in Africa
[23 genera, >200 species]

-. Tarsal segmentation : 1-1-1-1 to 1-2-2-4 ...3

3. Tarsal claws of legs 2 to 4 : setaceous **Family GALEODIDAE**

Tarsal segmentation : 1-2-2-3

Long-legged; small to large (12-70 mm)

Leg 1 : tarsi : without claws or with 1 or 2 claws

Male cheliceral flagellum : paraxially movable; a single, capitate (terminally enlarged) seta located on the mesial surface

Female genital opercula : not differentiated from other abdominal sternites and not specifically variable

Distribution : northern Africa, and Asia
[4 genera (10 in Roewer, 1934), 180 species]

-. Tarsal claws of legs 2 to 4 : smooth ...4

4. Leg 1 : tarsi : without claws **Family DAESIIDAE**

Tarsal segmentation : 1-1-1-1 to 1-2-2-4

Long-legged; tiny to moderate-sized (6-23 mm)

Male cheliceral flagellum : paraxially movable, ovate to irregular membranous structure attached to the mesial surface by a disk

Female genital opercula : not differentiated from other abdominal sternites and not specifically variable

Propeltidium : exterior lobes : fused

Distribution : Africa, southern Europe, Near East, and South America
[7 subfamilies, 34 genera, 182 species]

-. Leg 1 : tarsi : with 1 or 2 claws ...5

Tarsal segmentation : 1-1-1-1

Small to moderate-sized (8-26 mm); long-legged

Female genital opercula : differentiated from other abdominal

sternites and specifically variable

5. Chelicerae : multidentate

Male cheliceral flagellum : paraxially immovable; fanlike to coiled, whiplike seta located on the mesial surface, with associated modified setae and a dorsal cheliceral horn

Propeltidium : exterior lobes : posteriorly fused

Distribution : Asia and Near East to southeastern Europe and northwestern Africa

[5 genera, 41 species]
- Family KARSCHIIDAE

- . Chelicerae : not multidentate

Male cheliceral flagellum : paraxially immovable; dorsal, more or less membranous process associated with one or more strongly modified setae

Propeltidium : exterior lobes : free

Distribution : central Asia to Near East

[5 genera, 14 species]
- Family GYLIPPIDAE

Family Daesiidae

Key to Sub-Families and Genera :

Tarsal segmentation	Sub-Family	Ventral spination of : Tarsus 2-3 4		Genus
1-1-1-1	Gnosippinae	1.2.2.2	2.2.2.2.2	<i>Tarabulida</i>
		1.2.2.2.2	2.2.2.2.2.2	<i>Gnosippus</i>
1-1-1-2	Blossiinae	1.2.2.2	2.2.2/2.2	<i>Blossiola</i>
		2.2.2.2	2.2.2/2.2	<i>Blossia</i>
1-1-1-3	Gluviopsinae	1.2.2.2.2	2.2/2/2.2.2	<i>Gluviopsis</i>
		1.2.2.2.2	2.2/2/1.2.2	<i>Gluviopsilla</i>
		1.2.2.2.2	2.2/2/2.2	<i>Gluviopsona</i>
1-2-2-2	Triditarsinae	1.1/0	1.1/0	<i>Hodeidania</i>
1-2-2-3	Gluviinae	1.1/0	2.2/2/0	<i>Eberlanzia</i>
		2.2/2	2.2.2/2/2	<i>Gluviema</i>
1-2-2-4	Daesiinae	0/0	0/0/0/0	<i>Bitonota</i>
		1.1/0	2.2/0/2/0	<i>Biton</i>

Genus *Biton* Karsch, 1880*Biton bellulus* (Pocock, 1902)

Distribution: Egypt.

Ref: 1. *B.b.* Roewer, 1934 pp.391,401,402.*Biton brunneus* Roewer, 1934

Distribution: Morocco.

Ref: 1. *B.b.* Roewer, 1934 p.402.*Biton brunnipes* Pocock, 1896

Distribution: Somalia.

Ref: 1. *B.b.* Roewer, 1934 p.403.2. *B.b.* Delle Cave & Simonetta, 1971 pp.43-44.*Biton ehrenbergi* Karsch, 1880 {T}

Distribution: Egypt, Eritrea, Palestine, Saudi Arabia, Somalia, Sudan, Tunisia.

Ref: 1. *B.e.* Roewer, 1934 pp.390,391,400,402.2. *B.e.* Roewer, 1941 p.140.3. *B.e.* Benoit, 1964 pp.96-97.4. *B.e.* Delle Cave & Simonetta, 1971 pp.44-45.*Biton fessanus* Roewer, 1934

Distribution: Libya.

Ref: 1. *B.f.* Roewer, 1934 p.403.*Biton fuscipes* Pocock, 1897

Distribution: Somalia.

Ref: 1. *B.f.* Roewer, 1934 p.403.*Biton fuscus* (Kraepelin, 1899)

Distribution: Algeria.

Ref: 1. *B.f.* Roewer, 1934 p.403.*Biton laminatus* (Pocock, 1903)

Distribution: Arabia, Yemen.

Ref: 1. *B.l.* Roewer, 1934 p.403.*Biton lividus* Simon, 1882

Distribution: Sudan.

Ref: 1. *B.l.* Roewer, 1934 p.403.2. *B.l.* Benoit, 1964 p.97.

Biton lividus aristomenes Delle Cave & Simonetta, 1971

Distribution: Eritrea.

Ref: 1. *B.l.a.* Delle Cave & Simonetta, 1971 pp.53-56.

Biton magnifrons (Birula, 1904)

Distribution: Palestine, Somalia.

Ref: 1. *B.m.* Roewer, 1934 p.403.

2. *B.m.* Levy & Shulov, 1964 p.107.

Biton ragazzii (Kraepelin, 1899)

Distribution: Eritrea, Sudan.

Ref: 1. *B.r.* Roewer, 1934 p.403.

2. *B.r.* Roewer, 1941 p.140.

3. *B.r.* Benoit, 1964 p.97.

4. *B.r.* Delle Cave & Simonetta, 1971 pp.46-48.

Biton sabulosus Pocock, 1903

Distribution: Arabia.

Ref: 1. *B.s.* Roewer, 1934 p.403.

Biton simoni (Kraepelin, 1899)

Distribution: Djibouti.

Ref: 1. *B.s.* Roewer, 1934 p.403.

2. *B.s.* Delle Cave & Simonetta, 1971 pp.48-51.

Biton tarabulus Roewer, 1934

Distribution: Libya.

Ref: 1. *B.t.* Roewer, 1934 p.403.

Biton tunetanus Simon, 1885

Distribution: Algeria, Palestine, Tunisia.

Ref: 1. *B.t.* Roewer, 1934 p.402.

2. *B.t.* Levy & Shulov, 1964 p.107.

Biton velox Simon, 1885

Distribution: Libya, Tunisia.

Ref: 1. *B.v.* Roewer, 1934 p.402.

Biton velox dimitrievi (Birula, 1904)

Distribution: Palestine, Somalia.

Ref: 1. *B.dimitrievi* Roewer, 1934 p.403.

2. *B.dimitrievi* Levy & Shulov, 1964 p.108.

3. *B.v.d. Delle Cave & Simonetta*, 1971 pp.51-52.

Biton villosus Birula, ?

Distribution: Somalia ?.

Ref: 1. *B.v.* Roewer, 1934 p.403.

Biton wicki (Birula, 1915)

Distribution: Somalia, Sudan, Yemen.

Ref: 1. *B.w.* Roewer, 1934 p.403.

2. *Daesia w.* Whittick, 1941 pp.48-49.

3. *B.w.* Benoit, 1964 p.96.

4. *B.w.* Delle Cave & Simonetta, 1971 p.56.

Biton zederbaueri (Werner, 1905)

Distribution: Palestine.

Ref: 1. *B.z.* Roewer, 1934 p.402.

2. *B.z.* Levy & Shulov, 1964 p.107.

Genus *Bitonota* Roewer, 1934

Bitonota kraepelini Roewer, 1934 {T}

Distribution: Somalia.

Ref: 1. *B.k.* Roewer, 1934 pp.388-390.

Genus *Blossia* Simon, 1880

Blossia albocaudata Levy & Shulov, 1964

Distribution: Palestine.

Ref: 1. *B.a.* Levy & Shulov, 1964 pp.106-107.

Blossia electa Roewer, 1934

Distribution: Morocco.

Ref: 1. *B.e.* Roewer, 1934 p.372.

Blossia spinosa Simon, 1880 {T}

Distribution: Algeria, Egypt, Palestine, Sudan.

Ref: 1. *B.s.* Simon, 1880 p.400.

2. *B.s.* Roewer, 1934 pp.371-372.

3. *B.s.* Levy & Shulov, 1964 p.106.

Genus *Blossiola* Roewer, 1934

Blossiola aegyptiaca Roewer, 1934

Distribution: Egypt, Palestine.

Ref: 1. *B.a.* Roewer, 1934 pp.366,370.

2. *B.a.* Levy & Shulov, 1964 p.106.

Blossiola arabica Roewer, 1934

Distribution: Yemen.

Ref: 1. *B.a.* Roewer, 1934 p.370.

Blossiola ebneri Roewer, 1934

Distribution: Morocco, Palestine.

Ref: 1. *B.e.* Roewer, 1934 p.371.

2. *B.e.* Levy & Shulov, 1964 p.106.

Blossiola gluvioides Roewer, 1934

Distribution: Mauritania.

Ref: 1. *B.g.* Roewer, 1934 p.370.

Blossiola laticosta (Hewitt, 1919)

Distribution: Palestine, Somalia.

Ref: 1. *B.l.* Roewer, 1934 p.369.

2. *B.l.* Levy & Shulov, 1964 p.106.

3. *B.l.* Delle Cave & Simonetta, 1971 pp.51-52.

Blossiola maroccana Roewer, 1934

Distribution: Morocco.

Ref: 1. *B.m.* Roewer, 1934 p.370.

Blossiola nigripalpis Roewer, 1934

Distribution: Jordan, Palestine.

Ref: 1. *B.n.* Roewer, 1934 p.370.

2. *B.n.* Levy & Shulov, 1964 p.105.

Blossiola nigripalpis agriope Roewer, 1934

Distribution: Somalia.

Ref: 1. *B.n.a.* Delle Cave & Simonetta, 1971 pp.39-40.

Blossiola occidentalis Roewer, 1934

Distribution: Morocco, Palestine.

Ref: 1. *B.o.* Roewer, 1934 p.370.

2. *B.o.* Levy & Shulov, 1964 p.106.

Blossiola omeri Levy & Shulov, 1964

Distribution: Palestine.

Ref: 1. *B.o.* Levy & Shulov, 1964 p.106.

Genus *Eberlanzia* Roewer, 1941*Eberlanzia ? benedicti* Delle Cave & Simonetta, 1971

Distribution: Somalia.

Ref: 1. *E.b.* Delle Cave & Simonetta, 1971 pp.68-69.**Genus *Gluviema* Caporiacco, 1936***Gluviema migiurtina* Caporiacco, 1936 {T}

Distribution: Somalia.

Ref: 1. *G.m.* Roewer, 1941 p.139.**Genus *Gluviopsilla* Roewer, 1934***Gluviopsilla discolor* (Kraepelin, 1899) {T}

Distribution: Algeria, Syria.

Ref: 1. *G.d.* Roewer, 1934 p.378.**Genus *Gluviopsis* Kraepelin, 1899***Gluviopsis balfouri* (Pocock, 1895)

Distribution: Yemen (Socotra).

Ref: 1. *G.b.* Roewer, 1934 pp.377-378.*Gluviopsis butes* Delle Cave & Simonetta, 1971

Distribution: Somalia.

Ref: 1. *G.b.* Delle Cave & Simonetta, 1971 pp.61-63.*Gluviopsis nigripalpis* (Pocock, 1897)

Distribution: Somalia.

Ref: 1. *G.n.* Roewer, 1934 p.377.2. *G.n.* Delle Cave & Simonetta, 1971 pp.65-67.*Gluviopsis rivaie* (Pavesi, 1897)

Distribution: Somalia.

Ref: 1. *G.r.* Roewer, 1934 p.378.2. *G.r.* Delle Cave & Simonetta, 1971 pp.67-68.*Gluviopsis rufescens* (Pocock, 1897) {T}

Distribution: Djibouti, Yemen.

Ref: 1. *G.* Roewer, 1934 p.377.2. *G.r.* Delle Cave & Simonetta, 1971 pp.64-65.*Gluviopsis rufescens pygmaea* Delle Cave & Simonetta, 1971

Distribution: Djibouti, Somalia.

Ref: 1. *G.r.p.* Delle Cave & Simonetta, 1971 pp.63-64.

Gluiopsis somalica Roewer, 1934

Distribution: Somalia.

Ref: 1. *G.s.* Roewer, 1934 p.377.

Genus *Gluiopsona* Roewer, 1934

Gluiopsona lahavi Levy & Shulov, 1964

Distribution: Palestine.

Ref: 1. *G.l.* Levy & Shulov, 1964 p.107.

Gluiopsona nova Turk, 1960

Distribution: Jordan.

Ref: 1. *G.n.* Turk, 1960 pp.122-123.

Genus *Gnosippus* Karsch, 1880

Gnosippus franchettii Caporiacco, 1936

Distribution: Eritrea.

Ref: 1. *G.f.* Roewer, 1941 pp.130-131.

2. *G.f.* Delle Cave & Simonetta, 1971 pp.59-60.

Gnosippus klunzingeri Karsch, 1880 {T}

Distribution: Egypt, Palestine.

Ref: 1. *G.k.* Roewer, 1934 pp.355-356.

2. *G.k.* Levy & Shulov, 1964 p.105.

Gnosippus styloceros Kraepelin, 1899

Distribution: Palestine.

Ref: 1. *G.s.* Roewer, 1934 p.357.

2. *G.s.* Levy & Shulov, 1964 pp.104-105.

Gnosippus yemenensis (Simon, 1882)

Distribution: Oman, Yemen.

Ref: 1. *G.y.* Roewer, 1934 p.357.

Genus *Hodeidania* Roewer, 1934

Hodeidania brunnipalpis Roewer, 1934

Distribution: Yemen.

Ref: 1. *H.b.* Roewer, 1934 p.383.

Genus *Tarabulida* Roewer, 1934

Tarabulida ehippiata Roewer, 1934 {T}

Distribution: Libya.

Ref: 1. *T.e.* Roewer, 1934 p.354.

Tarabulida fumigata Roewer, 1934

Distribution: Libya.

Ref: 1. *T.f.* Roewer,1934 p.354.

Family Galeodidae

Key to Genera :

Number of ventral spines on :

Segments 1-2 of		Segments 1-3 of Tarsus 4	Genus
Tarsus 2	Tarsus 3		
1.2.2/2	1.2.2/2	2.2/2/0	<i>Galeodibus</i>
2.2/2	2.2/2	2.2.2/2/0	<i>Galeodora</i>
1.1.2/2 or 2.2.2/2	1.1.2/2 or 2.2.2/2	2.2.2/2/0	<i>Galeodes</i> s.str.
1.1.2.2/2 or 1.2.2.2/2	1.1.2.2/2 or 1.2.2.2/2	2.2.2/2/0	<i>Othoes</i>
1.1.2/2 or 1.2.2/2 or 2.2.2/2	1.1.2/2 or 1.2.2/2 or 2.2.2/2	2.2.2/2/1	<i>Galeodila</i>
1.1.2/2 or 2.2.2/2	1.1.2/2 or 2.2.2/2	2.2.2/2/2	<i>Galeodellus</i>
1.1.2.2/2	1.1.2.2/2	2.2.2/2/2	<i>Galeodarus</i>
1.1.2/2.1	1.1.2/2.1	2.2.2/2.2/2	<i>Galeodopsis</i>
2.2.2/2	2.2.2/2	2.2.2.2/2/0	<i>Galeodessus</i>

Genus *Galeodarus* Roewer, 1934

Galeodarus insidiator Roewer, 1934 {T}

Distribution: Yemen.

Ref: 1. *G.i.* Roewer,1934 p.548.

Genus *Galeodellus* Roewer, 1934

Galeodellus darius (Pocock, 1895)

Distribution: Palestine.

Ref: 1. *G.d.* Roewer,1934 p.547.

2. *G.d.* Levy & Shulov,1964 p.111.

Galeodellus lehmanni (Birula, 1890)

Distribution: Palestine.

Ref: 1. *G.l.* Roewer, 1934 pp.543-544.

2. *G.l.* Levy & Shulov, 1964 p.111.

Galeodellus mosconi-bronzii Caporiacco, 1936

Distribution: Somalia.

Ref: 1. *G.m.* Roewer, 1941 pp.170-171.

Galeodellus tarabulus Roewer, 1934

Distribution: Libya.

Ref: 1. *G.t.* Roewer, 1934 p.547.

Genus *Galeodes* Pallas, 1772

Galeodes arabs C.L.Koch, 1842

Distribution: Algeria, Arabia, Egypt, Iraq, Libya, Morocco, Palestine, Sudan, Syria, Tunisia, Yemen.

Ref: 1. *G.a.* Tullgren, 1909 p.1.

2. *G.a.* Roewer, 1934 pp.518-519, 522-523, 532.

3. *G.a.* Roewer, 1941 p.161.

4. *G.a.* Levy & Shulov, 1964 p.109.

5. *G.a.* Benoit, 1964 p.97.

Galeodes araneoides (Pallas, 1772) {T}

Distribution: Egypt, Iraq, Jordan, Palestine.

Ref: 1. *Solpuga a.* Audouin, 1825 pp.176-178.

2. *Solpuga intrepida* Audouin, 1825 p.178.

3. *Solpuga a.* Cambridge, 1870 p.818.

4. *G.a.* Roewer, 1934 pp.516, 523-524, 526-527.

5. *G.a.* Roewer, 1941 p.159.

6. *G.a.* Levy & Shulov, 1964 p.109.

Galeodes atriceps Roewer, 1934

Distribution: Palestine.

Ref: 1. *G.a.* Roewer, 1934 pp.529.

2. *G.a.* Levy & Shulov, 1964 p.109.

Galeodes babylonicus Roewer, 1934

Distribution: Iraq, Palestine.

Ref: 1. *G.b.* Roewer, 1934 p.532.

2. *G.b.* Levy & Shulov, 1964 p.110.

Galeodes bacilliferoides Birula, 1908

Distribution: Iraq.

Ref: 1. *G.b.* Roewer, 1934 p.529.

Galeodes barbarus Lucas, 1846

Distribution: Algeria, Egypt, Ethiopia, Libya, Morocco, Somalia, Sudan, Tunisia.

Ref: 1. *Paragaleodes b.* Tullgren, 1909 p.2.

2. *G.b.* Roewer, 1934 pp.516,523,534.

3. *G.b.* Roewer, 1941 p.162.

4. *G.b.* Benoit, 1964 pp.95-96.

Galeodes bicolor Roewer, 1934

Distribution: Jordan, Palestine.

Ref: 1. *G.b.* Roewer, 1934 p.530.

2. *G.b.* Turk, 1960 p.117.

3. *G.b.* Levy & Shulov, 1964 p.109.

Galeodes birulae Roewer, 1941

Distribution: Iraq.

Ref: 1. *G.b.* Roewer, 1941 pp.162-163.

Galeodes bogojavlenskii Birula, 1906

Distribution: Palestine.

Ref: 1. *G.b.* Roewer, 1934 p.527.

2. *G.b.* Levy & Shulov, 1964 p.109.

Galeodes clavatus Roewer, 1934

Distribution: Tunisia.

Ref: 1. *G.c.* Roewer, 1934 p.534.

Galeodes crassichelis Roewer, 1934

Distribution: Tunisia.

Ref: 1. *G.c.* Roewer, 1934 p.534.

Galeodes dorsalis Roewer, 1934

Distribution: Palestine, Saudi Arabia, Syria.

Ref: 1. *G.d.* Roewer, 1934 p.532.

2. *G.d.* Levy & Shulov, 1964 p.110.

Galeodes edentatus Benoit, 1964

Distribution: Sudan.

Ref: 1. *G.e.* Benoit, 1964 pp.93-95.

Galeodes fulvipes (Birula, 1904)

Distribution: Palestine.

Ref: 1. *G.f.* Roewer, 1934 p.530.

2. *G.f.* Levy & Shulov, 1964 p.109.

Galeodes graecus C.L.Koch, 1842

Distribution: Egypt, Syria.

Ref: 1. *G.g.* Simon, 1899 p.244.

2. *G.g.* Roewer, 1934 pp.521,525,531.

3. *G.g.* Roewer, 1941 p.160.

Galeodes granti Pocock, 1903

Distribution: Egypt, Palestine, Saudi Arabia, Somalia, Sudan, Syria, Yemen.

Ref: 1. *G.g.* Roewer, 1934 pp.515,522,532.

2. *G.g.* Roewer, 1941 p.162.

3. *G.g.* Benoit, 1964 p.93.

4. *G.g.* Levy & Shulov, 1964 p.109.

Galeodes judaicus (Kraepelin, 1899)

Distribution: Palestine.

Ref: 1. *G.j.* Roewer, 1934 pp.531-532.

2. *G.j.* Roewer, 1941 p.160.

3. *G.j.* Levy & Shulov, 1964 p.109.

Galeodes kraepelini Roewer, 1934

Distribution: Egypt.

Ref: 1. *G.k.* Roewer, 1934 pp.517,533.

Galeodes lacertosus Roewer, 1934

Distribution: Yemen.

Ref: 1. *G.l.* Roewer, 1934 p.532.

Galeodes laniator Roewer, 1934

Distribution: Iraq, Palestine.

Ref: 1. *G.l.* Roewer, 1934 p.532.

2. *G.l.* Roewer, 1941 p.161.

3. *G.l.* Levy & Shulov, 1964 p.109.

Galeodes litigosus Roewer, 1934

Distribution: Yemen.

Ref: 1. *G.l.* Roewer, 1934 p.532.

Galeodes lybicus Roewer, 1941

Distribution: Libya.

Ref: 1. *G.l.* Roewer, 1941 p.168.

Galeodes medusae Turk, 1960

Distribution: Egypt.

Ref: 1. *G.m.* Turk, 1960 pp.114-115.

Galeodes minimus Roewer, 1934

Distribution: Morocco.

Ref: 1. *G.m.* Roewer, 1934 p.535.

Galeodes minitor Roewer, 1934

Distribution: Morocco.

Ref: 1. *G.m.* Roewer, 1934 p.535.

Galeodes (Galeodellus) pinkasi Turk, 1960

Distribution: Jordan.

Ref: 1. *G.(G.)p.* Turk, 1960 pp.115-116.

Galeodes nesterovi (Birula, 1916)

Distribution: Iraq.

Ref: 1. *G.n.* Roewer, 1934 p.532.

Galeodes occidentalis Simon, 1885

Distribution: Algeria, Mauritania, Morocco.

Ref: 1. *G.o.* Roewer, 1934 p.534.

Galeodes palpalis Roewer, 1934

Distribution: Tunisia.

Ref: 1. *G.p.* Roewer, 1934 p.534.

Galeodes revestitus Roewer, 1934

Distribution: Morocco.

Ref: 1. *G.r.* Roewer, 1934 p.535.

Galeodes rhamses Roewer, 1934

Distribution: Egypt.

Ref: 1. *G.r.* Roewer, 1934 pp.515,522,534.

Galeodes scalaris C.L.Koch, 1842

Distribution: Saudi Arabia, Egypt, Libya, Morocco.

Ref: 1. *G.s.* Roewer, 1934 pp.519-520,526,533.

2. *G.s.* Roewer, 1941 p.162.

Galeodes schendicus Roewer, 1934

Distribution: Egypt ?, Sudan.

Ref: 1. *G.s.* Roewer, 1934 p.533.

Galeodes sericeus (Kraepelin, 1899)

Distribution: Egypt ?, Sudan.

Ref: 1. *G.s.* Roewer, 1934 pp.517,523,533.

Galeodes simplex Roewer, 1934

Distribution: Tunisia.

Ref: 1. *G.s.* Roewer, 1934 p.534.

Galeodes somalicus Roewer, 1934

Distribution: Somalia.

Ref: 1. *G.s.* Roewer, 1934 p.534.

Galeodes sulfuripes Roewer, 1934

Distribution: Iraq, Palestine.

Ref: 1. *G.s.* Roewer, 1934 p.532.

2. *G.s.* Levy & Shulov, 1964 pp.109-110.

Galeodes theodori Turk, 1960

Distribution: Egypt.

Ref: 1. *G.t.* Turk, 1960 pp.113-114.

Galeodes tunetanus (Kraepelin, 1899)

Distribution: Tunisia.

Ref: 1. *G.t.* Roewer, 1934 p.534.

2. *G.t.* Roewer, 1941 p.162.

Galeodes unicolor (Birula, 1905)

Distribution: Palestine.

Ref: 1. *G.u.* Roewer, 1934 p.530.

2. *G.u.* Levy & Shulov, 1964 p.109.

Galeodes veemi Whittick, 1939

Distribution: Egypt.

Ref: 1. *G.v.* Roewer, 1941 pp.166-167.

Galeodes venator Simon, 1879

Distribution: Algeria, Morocco.

Ref: 1. *G.v.* Roewer, 1934 pp.534-535.

Galeodes ventralis Roewer, 1934

Distribution: Yemen.

Ref: 1. *G.v.* Roewer, 1934 p.533.

Genus *Galeodessus* Roewer, 1934*Galeodessus taurus* Roewer, 1934

Distribution: Iraq.

Ref: 1. *G.t.* Roewer, 1934 p.549.

Genus *Galeodibius* Roewer, 1934*Galeodibius blanchardi* (Simon, 1891) {T}

Distribution: Algeria, Libya, Morocco, Tunisia.

Ref: 1. *G.b.* Roewer, 1934 p.513.

2. *G.b.* Roewer, 1941 p.158.

Galeodibius fessanus Roewer, 1934

Distribution: Libya.

Ref: 1. *G.f.* Roewer, 1934 p.513.

Galeodibius flavivittatus Roewer, 1934

Distribution: Algeria.

Ref: 1. *G.f.* Roewer, 1934 p.513.

Galeodibius olivieri (Simon, 1879)

Distribution: Algeria, Libya, Mauritania, Morocco.

Ref: 1. *G.o.* Roewer, 1934 p.513.

2. *G.o.* Roewer, 1941 pp.157-158.

3. *G.o.* Lawrence, 1953 p.971.

Galeodibius tripolitanus Roewer, 1934

Distribution: Libya, Morocco.

Ref: 1. *G.t.* Roewer, 1934 p.513.

2. *G.t.* Roewer, 1941 p.158.

Genus *Galeodila* Roewer, 1934*Galeodila pusilla* Roewer, 1934

Distribution: Algeria, Palestine.

Ref: 1. *G.p.* Roewer, 1934 p.537.2. *G.p.* Levy & Shulov, 1964 pp.110-111.*Galeodila tillmani* Whittick, 1939

Distribution: Somalia.

Ref: 1. *G.t.* Roewer, 1941 p.159.**Genus *Galeodopsis* Birula, 1903***Galeodopsis tripolitanus* Hirst, 1912

Distribution: Libya.

Ref: 1. *G.t.* Roewer, 1934 p.549.**Genus *Galeodora* Roewer, 1934***Galeodora adamsi* Turk, 1947

Distribution: Iraq.

Ref: 1. *G.a.* Turk, 1947 pp.77-80.*Galeodora distincta* Roewer, 1934 {T}

Distribution: Lebanon.

Ref: 1. *G.d.* Roewer, 1934 p.513.**Genus *Othoes* Hirst, 1911***Othoes floweri* Hirst, 1911 {T}

Distribution: Egypt.

Ref: 1. *O.f.* Roewer, 1934 p.536.*Othoes vittatus* Hirst, 1912

Distribution: Palestine.

Ref: 1. *O.v.* Roewer, 1934 p.536.2. *O.v.* Levy & Shulov, 1964 p.110.

Family Gylippidae**Key to Genera :**

1. Male chelicera with 2 principal spines and 1 digital spine;
Female genital sternites convex in shape

Gylippus

- . Male chelicera with 1 principal spine, without digital spine;
 Female genital sternites concave in shape *Acanthogylippus*

Genus *Acanthogylippus* Birula, 1913

Acanthogylippus judaicus (Kraepelin, 1899) {T}

Distribution: Palestine.

Ref: 1. *A.j.* Roewer, 1934 p.318.

2. *A.j.* Levy & Shulov, 1964 p.104.

Genus *Gylippus* Simon, 1879

Gylippus shulovi Turk, 1948

Distribution: Palestine.

Ref: 1. *G.s.* Levy & Shulov, 1964 p.104.

Gylippus syriacus (Simon, 1872) {T}

Distribution: Iraq, Palestine, Syria.

Ref: 1. *G.s.* Roewer, 1934 p.311.

2. *G.s.* Roewer, 1941 p.114.

3. *G.s.* Levy & Shulov, 1964 p.104.

Gylippus yerohami Levy & Shulov, 1964

Distribution: Palestine.

Ref: 1. *G.y.* Levy & Shulov, 1964 p.104.

Gylippus sp.

Distribution: Yemen (Socotra).

Ref: 1. *G.sp.* Turk, 1960 p.121.

Family Karschiidae

Key to Genera :

1. Male flagellum with a strongly differentiated bristles tuft. Female
 cheliceral movable finger with 2 or more small intermediate-teeth
 between front and main teeth. *Karschia*
- . Male flagellum more or less oval elongated in shape, without bristles..2

2. Male flagellum, prolaterally, without a horn;
 Ocular area with dense tubular hairs. ***Barrus***
- Male flagellum, prolaterally, with a horn ..3
3. Male flagellum, prolaterally, with an acute horn;
 Ocular area and also the front edge of male's propeltidium with soft tubular hairs. ***Barrella***
- Male flagellum, prolaterally, with a curved, more or less, blunt horn;
 Ocular area and also the front edge of male's propeltidium only with normal bristles and hairs. ***Eusimonia***

Genus *Barrella* Hirst, 1910

Barrella walsinghami Hirst, 1910 {T}

World Distribution: Algeria.

Ref: 1. *B.w.* Roewer, 1934 p.304.

Genus *Barrus* Simon, 1880

Barrus letourneuxi Simon, 1880 {T}

Distribution: Egypt.

Ref: 1. *B.l.* Simon, 1880 pp.401-402.

2. *B.l.* Roewer, 1934 pp.305-306.

Genus *Eusimonia* Kraepelin, 1899

Eusimonia arabica Roewer, 1934

Distribution: Yemen.

Ref: 1. *E.a.* Roewer, 1934 p.302.

Eusimonia furcillata (Simon, 1872) {T}

Distribution: Palestine.

Ref: 1. *E.f.* Roewer, 1934 p.302.

2. *E.f.* Levy & Shulov, 1964 p.104.

Eusimonia kabiliana (Simon, 1879)

Distribution: Algeria, Egypt, Palestine.

Ref: 1. *E.k.* Roewer, 1934 pp.301-302.

2. *E.k.* Levy & Shulov, 1964 p.104.

Eusimonia mirabilis Roewer, 1934

Distribution: Libya.

Ref: 1. *E.m.* Roewer, 1934 p.302.

Eusimonia nigrescens Kraepelin, 1899

Distribution: Syria ?.

Ref: 1. *E.n.* Roewer,1934 p.302.

Eusimonia orthoplax Kraepelin, 1899

Distribution: Algeria.

Ref: 1. *E.o.* Roewer,1934 p.302.

Genus *Karschia* Walter, 1889

Karschia kurdistanica Birula, 1935

Distribution: Iraq ?.

Ref: 1. *K.k.* Roewer,1941 pp.110-111.

Karschia sp.

Distribution: Egypt.

Ref: 1. *K.sp.* El-Hennawy,1998 p.28.

Family Rhagodidae

Key to Genera (20) :

Number of ventral spines on :

Tarsus 2 & 3	Tarsus 4	Genus
0	0	<i>Rhagodes</i>
1.1	0	<i>Rhagodax</i>
0	1.1	<i>Rhagoduna</i>
0	1.2	<i>Rhagodula</i>
0	1.1.1	<i>Rhagodoca</i>
0	1.2.2	<i>Rhagodopa</i>
1	1.1.1	<i>Rhagodospus</i>
1.1	1.2.2	<i>Rhagoditta</i>
0	2.2.2	<i>Rhagodinus</i>
1.1	2.2.2	<i>Rhagodira</i>
1.2	2.2.2	<i>Rhagodixa</i>
2.2.2	2.2.2	<i>Rhagodia</i>
0	2.2.2.2	<i>Rhagoderus</i>
1.2	1.2.2.2	<i>Rhagoderma</i>
1.2	2.2.2.2	<i>Rhagodessa</i>
1.1.2	1.2.2.2	<i>Rhagodippa</i>
1.2.2	1.1.2.2	<i>Rhagodeya</i>
1.2.2	2.2.2.2	<i>Rhagodeca</i>

2.2.2	2.2.2.2	<i>Rhagodella</i>
2.2.2.2	2.2.2.2	<i>Rhagodalma</i>

Note. Genus *Rhagodorimus* is not included in the key.

Genus *Rhagodalma* Roewer, 1934

Rhagodalma melanocephala Roewer, 1934 {T}

Distribution: Sudan.

Ref: 1. *R.m.* Roewer,1934 p.288.

Genus *Rhagodax* Roewer, 1941

Rhagodax transjordania Turk, 1960

Distribution: Jordan.

Ref: 1. *R.t.* Turk,1960 pp.120-121.

Rhagodax wadidaba Roewer, 1941 {T}

Distribution: Jordan.

Ref: 1. *R.w.* Roewer,1941 p.102.

Genus *Rhagodeca* Roewer, 1934

Rhagodeca fuscichelis Roewer, 1941

Distribution: Palestine, Syria.

Ref: 1. *R.f.* Roewer,1941 p.108.

2. *R.f.* Levy & Shulov,1964 p.103.

Rhagodeca hirsti Roewer, 1934

Distribution: Palestine.

Ref: 1. *R.h.* Roewer,1934 p.286.

2. *R.h.* Levy & Shulov,1964 p.103.

Rhagodeca impavida (C.L.Koch, 1842) {T}

Distribution: Arabia, Oman.

Ref: 1. *R.i.* Roewer,1934 p.285-286.

Genus *Rhagodella* Roewer, 1934

Rhagodella atra Roewer, 1934

Distribution: Palestine.

Ref: 1. *R.a.* Roewer,1934 p.288.

2. *R.a.* Roewer,1941 p.109.

3. *R.a.* Levy & Shulov,1964 p.103.

Genus *Rhagoderma* Roewer, 1934*Rhagoderma tricolor* Roewer, 1941

Distribution: Palestine.

Ref: 1. *R.t.* Roewer, 1941 p.107.2. *R.t.* Levy & Shulov, 1964 p.103.**Genus *Rhagoderus* Roewer, 1934***Rhagoderus griseopilosus* Roewer, 1934 {T}

Distribution: Palestine.

Ref: 1. *R.g.* Roewer, 1934 pp.281-282.2. *R.g.* Levy & Shulov, 1964 p.103.**Genus *Rhagodes* Pocock, 1897***Rhagodes aegypticus* Roewer, 1934

Distribution: Egypt.

Ref: 1. *R.a.* Roewer, 1934 pp.269-270.*Rhagodes furiosus* (C.L.Koch, 1842)

Distribution: Egypt, Libya.

Ref: 1. *R.f.* Roewer, 1934 pp.269-270.2. *R.f.* Roewer, 1941 p.101.3. *R.f.* Lawrence, 1953 p.955.*Rhagodes melanus* (Olivier, 1807) {T}

Distribution: Algeria, Egypt, South Palestine.

Ref: 1. *Solpuga m.* Audouin, 1825 pp.178-179.2. *Solpuga m.* Cambridge, 1870 p.818.3. *R.m.* Roewer, 1934 pp.269-270.4. *R.m.* Lawrence, 1953 p.955.5. *R.m.* Levy & Shulov, 1964 p.103.*Rhagodes nicotrae* Caporiacco, 1939

Distribution: Somalia.

Ref: 1. *R.n.* Roewer, 1941 p.101.*Rhagodes strandi* Caporiacco, 1939

Distribution: Somalia.

Ref: 1. *R.n.* Roewer, 1941 p.101.*Rhagodes subaureus* Roewer, 1934

Distribution: Somalia.

Ref: 1. *R.s.* Roewer, 1934 p.270.

2. *R.s.* Lawrence, 1953 p.955.

Genus *Rhagodessa* Roewer, 1934

Rhagodessa cloudsleythompsoni Benoit, 1964

Distribution: Sudan.

Ref: 1. *R.c.* Benoit, 1964 pp.91-92.

Rhagodessa judaica (Kraepelin, 1899)

Distribution: Palestine, Syria.

Ref: 1. *R.j.* Roewer, 1934 p.283.

2. *R.j.* Roewer, 1941 p.107.

3. *R.j.* Levy & Shulov, 1964 p.103.

Rhagodessa melanocephala (Simon, 1879) {T}

Distribution: Sudan.

Ref: 1. *R.m.* Roewer, 1934 p.283.

2. *R.m.* Benoit, 1964 p.97.

Rhagodessa sudanensis Roewer, 1934

Distribution: Sudan.

Ref: 1. *R.s.* Roewer, 1934 p.283.

2. *R.s.* Benoit, 1964 p.97.

Rhagodessa zionensis Roewer, 1934

Distribution: Palestine.

Ref: 1. *R.z.* Roewer, 1934 p.283.

2. *R.z.* Roewer, 1941 p.107.

3. *R.z.* Levy & Shulov, 1964 p.103.

Genus *Rhagodeya* Roewer, 1934

Rhagodeya nigra Caporiacco, 1937

Distribution: Libya.

Ref: 1. *R.n.* Roewer, 1941 p.108.

Rhagodeya nubia Roewer, 1934 {T}

Distribution: Sudan.

Ref: 1. *R.n.* Roewer, 1934 p.284.

Genus *Rhagodia* Roewer, 1934

Rhagodia obscurior (Penther, 1913) {T}

Distribution: Iraq.

Ref: 1. *R.o.* Roewer, 1934 p.281.

Genus *Rhagodinus* Roewer, 1934*Rhagodinus caenaeicus* (Penther, 1913) {T}

Distribution: Iraq, Palestine.

Ref: 1. *R.c.* Roewer, 1934 p.279.2. *R.c.* Levy & Shulov, 1964 p.103.*Rhagodinus incertus* Caporiacco, 1936

Distribution: Eritrea.

Ref: 1. *R.i.* Roewer, 1941 pp.105-106.**Genus *Rhagodippa* Roewer, 1934***Rhagodippa albatra* Roewer, 1934 {T}

Distribution: Djibouti.

Ref: 1. *R.a.* Roewer, 1934 p.284.**Genus *Rhagodira* Roewer, 1934***Rhagodira algerica* Roewer, 1934

Distribution: Algeria, Morocco, Tunisia.

Ref: 1. *R.a.* Roewer, 1934 p.280.2. *R.a.* Roewer, 1941 p.106.*Rhagodira ochropus* (Dufour, 1862) {T}

Distribution: Algeria, Tunisia.

Ref: 1. *R.o.* Roewer, 1934 p.280.**Genus *Rhagoditta* Roewer, 1934***Rhagoditta bacillata* Roewer, 1941

Distribution: Tunisia.

Ref: 1. *R.b.* Roewer, 1941 p.105.*Rhagoditta corallipes* (Simon, 1885)

Distribution: Algeria, Tunisia.

Ref: 1. *R.c.* Roewer, 1934 p.279.*Rhagoditta phalangium* (Olivier, 1807) {T}

Distribution: Djibouti, Egypt.

Ref: 1. *Solpuga p.* Audouin, 1825 p.179.2. *R.p.* Roewer, 1934 pp.278-279.**Genus *Rhagodixa* Roewer, 1934***Rhagodixa kurdistan* (Birula, 1936)

Distribution: Iraq.

Ref: 1. *R.k.* Roewer, 1941 p.106.

Genus *Rhagodoca* Roewer, 1934

Rhagodoca macrocephala Roewer, 1934

Distribution: Somalia.

Ref: 1. *R.m.* Roewer, 1934 p.275.

Rhagodoca phillipsi (Pocock, 1896)

Distribution: Somalia.

Ref: 1. *R.p.* Roewer, 1934 p.274.

Rhagodoca picta Roewer, 1934

Distribution: Somalia.

Ref: 1. *R.p.* Roewer, 1934 p.274.

Rhagodoca somalica Roewer, 1934

Distribution: Somalia.

Ref: 1. *R.s.* Roewer, 1934 p.274.

2. *R.s.* Roewer, 1941 p.104.

Genus *Rhagodopa* Roewer, 1934

Rhagodopa jaffana Roewer, 1934

Distribution: Palestine.

Ref: 1. *R.j.* Roewer, 1934 p.277.

2. *R.j.* Levy & Shulov, 1964 p.103.

Rhagodopa jemenensis Roewer, 1934

Distribution: Yemen.

Ref: 1. *R.j.* Roewer, 1934 p.277.

Genus *Rhagodorimus* Turk, 1948 (not included in the key)

Rhagodorimus judaicus Turk, 1948

Distribution: Palestine.

Ref: 1. *R.j.* Levy & Shulov, 1964 p.103.

Genus *Rhagodospus* Roewer, 1941

Rhagodospus babylonicus (Birula, 1935) {T}

Distribution: Iraq.

Ref: 1. *R.b.* Roewer, 1941 p.105.

Genus *Rhagodula* Roewer, 1941

Rhagodula nigra Roewer, 1941

Distribution: Palestine.
Ref: 1. *R.n.* Roewer,1941 p.103.
2. *R.n.* Levy & Shulov,1964 p.103.

Genus *Rhagoduna* Roewer, 1934

Rhagoduna deserticola Roewer, 1941

Distribution: Syria.
Ref: 1. *R.d.* Roewer,1941 p.103.

Rhagoduna nocturna Roewer, 1934 {T}

Distribution: Sudan.
Ref: 1. *R.n.* Roewer,1934 p.271.
2. *R.n.* Benoit,1964 p.97.

Rhagoduna puccionii (Caporiacco, 1927)

Distribution: Syria.
Ref: 1. *R.p.* Roewer,1941 pp.102-103.

Family Solpugidae

Key to Genera (10) :

1. Tarsal segmentation : 1-4-4-6
- Subfamily Ferrandiinae
- Number of ventral spines on :

Segments 2-4 of		Segments 2-6 of Tarsus 4	Genus
Tarsus 2	Tarsus 3		
2/0/0	2/0/1	2/0/2/0/0	<i>Ferrandia</i>
2/0/0	2/0/0	2/0/2/0/0	<i>Enea</i>

- . Tarsal segmentation : 1-4-4-7
- Subfamily Solpuginae ...2

2. First segment of tarsus 2 only dorsally with regular hairs ...3
- . First segment of tarsus 2 dorsally with hairs and besides with distinct stand out long row of 7 spinous-bristles; Number of ventral spines on :

Segments 2-4 of		Segments 2-7 of Tarsus 4	Genus
Tarsus 2	Tarsus 3		
2/1/2	2/1/2	2/2/0/2/0/2	<i>Oparbella</i>
2/1/2	2/1/2	2/2/1/2/0/2	<i>Oparbona</i>
2/2/2	2/2/2	2/2/2/2/0/2	<i>Oparba</i>

3. Deutosternum wedge-shaped; metatarsus of pedipalp ventrally often spiny; ventral spination of : segments 2-4 of tarsus 2 & 3 : 2/0/2, segments 2-7 of tarsus 4 : 2/2/0/2/0/2 *Zeriassa*
- Deutosternum staff-shaped; metatarsus of pedipalp ventrally never spiny; Number of ventral spines on :

Segments 2-4 of		Segments 2-7 of Tarsus 4	Genus
Tarsus 2	Tarsus 3		
2/0/2	2/0/2	2/2/0/2/0/2	<i>Solpugassa</i>
2/1/2	2/1/2	2/2/2/2/0/2	<i>Solpugyla</i>
2/2/2	2/2/2	2/2/0/2/0/2	<i>Solpugeira</i>
2/2/2	2/2/2	2/2/2/2/0/2	<i>Solpuga</i>

Genus *Enea* Roewer, 1934

Enea birulae Roewer, 1934 {T}

Distribution: Somalia.

Ref: 1. *E.b.* Roewer,1934 p.413-414.

Genus *Ferrandia* Roewer, 1934

Ferrandia ferrandii (Kraepelin, 1899) {T}

Distribution: Somalia.

Ref: 1. *F.f.* Roewer,1934 pp.412-413.

Genus *Solpuga* Pocock, 1897

Solpuga funksoni Birula, 1915

Distribution: Sudan.

Ref: 1. *S.f.* Roewer,1934 p.465.

2. *S.f.* Benoit,1964 p.97.

Solpuga neumanni Kraepelin, 1903

Distribution: Somalia.

Ref: 1. *S.n.* Roewer,1934 p.464.

Solpuga parkinsoni Pocock, 1897

Distribution: Somalia.

Ref: 1. *S.p.* Roewer,1934 p.464.

Solpuga persephone (Simon, 1879)

Distribution: Algeria.

Ref: 1. *S.p.* Roewer,1934 pp.464-465.

Solpuga schweinfurthi Karsch, 1880

Distribution: Sudan.

- Ref: 1. *S.s.* Roewer, 1934 p.464.
 2. *S.s.* Lawrence, 1953 p.970.
 3. *S.s.* Benoit, 1964 p.97.

Genus *Solpugassa* Roewer, 1934

Solpugassa dentatidens (Simon, 1879)

Distribution: Somalia.

- Ref: 1. *S.d.* Roewer, 1934 p.431.

Genus *Solpugeira* Roewer, 1934

Solpugeira fusciorufa (Schenkel, 1933) {T}

Distribution: Somalia.

- Ref: 1. *S.f.* Roewer, 1934 p.440.
 2. *S.f.* Roewer, 1941 p.149.

Genus *Solpugyla* Roewer, 1934

Solpugyla maestrii Caporiacco, 1939

Distribution: Somalia.

- Ref: 1. *S.m.* Roewer, 1941 p.149.

Genus *Zeriassa* Pocock, 1897

Zeriassa bicolor (Pocock, 1897) {T}

Distribution: Somalia.

- Ref: 1. *Z.b.* Roewer, 1934 p.428.

Zeriassa longicornis Lawrence, 1953

Distribution: Somalia.

- Ref: 1. *Z.l.* Lawrence, 1953 pp.965-966.

Zeriassa ruspolii (Pavesi, 1897)

Distribution: Somalia.

- Ref: 1. *Z.r.* Roewer, 1934 p.428.

Zeriassa sudanica Roewer, 1934

Distribution: Sudan.

- Ref: 1. *Z.s.* Roewer, 1934 p.428.
 2. *Z.s.* Benoit, 1964 p.97.

Genus *Oparba* Roewer, 1934*Oparba maroccana* (Kraepelin, 1899) {T}

Distribution: Morocco.

Ref: 1. *O.m.* Roewer, 1934 p.485.2. *O.m.* Roewer, 1941 p.157.**Genus *Oparbella* Roewer, 1934***Oparbella aciculata* (Simon, 1879)

Distribution: Algeria, Palestine, Tunisia.

Ref: 1. *O.a.* Roewer, 1934 pp.483.2. *O.a.* Levy & Shulov, 1964 p.108.*Oparbella bicolor* Roewer, 1934

Distribution: Tunisia.

Ref: 1. *O.b.* Roewer, 1934 pp.483.*Oparbella flavescens* (C.L.Koch, 1842) {T}

Distribution: Algeria, Egypt, Libya, Morocco, Tunisia.

Ref: 1. *O.f.* Roewer, 1934 pp.481-483.2. *O.f.* Roewer, 1941 p.156.*Oparbella quedenfeldti* (Kraepelin, 1899)

Distribution: Egypt, Morocco.

Ref: 1. *O.q.* Roewer, 1934 pp.482-483.2. *O.q.* Roewer, 1941 p.156.*Oparbella weneri* (Birula, 1914)

Distribution: Algeria, Palestine, Tunisia.

Ref: 1. *O.w.* Roewer, 1934 pp.483.2. *O.w.* Roewer, 1941 p.157.3. *O.w.* Levy & Shulov, 1964 p.108.**Genus *Oparbona* Roewer, 1934***Oparbona simoni* Roewer, 1934 {T}

Distribution: Algeria.

Ref: 1. *O.s.* Roewer, 1934 pp.484.

Acknowledgments

The author is very grateful to Dr. Jürgen Gruber of Naturhistorisches Museum Wien for the photocopy of Roewer's " Solifugae, ... " (1934) without which the achievement of this work was not possible.

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VOLUME 6
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February, 2000

Cairo, Egypt

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The Spider (Araneae) fauna of the cotton fields located in the western part of Turkey

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Abstract

In this study, the spider (Araneae) fauna of cotton (*Gossypium hirsutum* Linnaeus) fields located in the western part of Turkey was studied. The spiders were collected from the ground and field zones. The aspirator and the whole plant bag sampling technique were used in the collection. A total of 613 spiders were obtained from eight cotton fields during the period of July-September in 1997 and 1998. Forty one species belonging to 31 genera and 12 families were determined. Numerically, Lycosidae was the dominant family, representing 29.0 % of all spiders collected, and *Pardosa proxima* (C.L.Koch) the dominant species, representing 21.1 % of the adult specimens. The four most common spider families were Lycosidae, Linyphiidae, Salticidae and Philodromidae.

Key Words: Araneae, cotton, fauna, Turkey, spiders.

Introduction

Natural living areas and agricultural ecosystems are important habitats for spiders. The ecological and faunistic studies which have been done on agricultural ecosystems showed that spiders are important agents in the pest control. For this reason, spiders in the agroecosystem have been the subject of numerous studies (Luczak, 1975, 1979; Nyffeler, 1982; Riechert & Lockley, 1984; Ruzicka, 1986; Clausen, 1986; Mansour, 1987; Sunderland, 1987; Nyffeler & Benz, 1987, 1988). Recently, a great deal of research has been undertaken to study the spider fauna of cereals such as soybean (Ferguson *et al.*, 1984), winter wheat (Jones, 1976; Sunderland, 1987), paddy (Zheng *et al.*, 1985) or alfalfa (Yeargan & Dondale, 1974), tobacco (Bayram *et al.*, 1998) and cotton fields (Leigh & Hunter, 1969; Mansour, 1987; Van den Berg *et al.*, 1990).

The present study constitutes an attempt to study the spider fauna of the cotton fields located in the western part of Turkey.

Material and Methods

1. Study area :

This study was performed on cotton fields located in the Aegean Region of Turkey in 1997 and 1998 (Fig. 1). The field work was started in early June and stopped in late September of each year. Eight cotton fields were investigated in the provinces of Manisa, İzmir and Aydın. Four fields are located in the vicinity of Saruhanlı (Manisa, *Field1=3 km east of Saruhanlı, about 25 acres; F2=8 km north of Saruhanlı, about 60 acres; F3=6 km west of Saruhanlı, about 30 acres; F4=5 km south of Saruhanlı, about 20 acres*), one field is located in the vicinity of Kemalpaşa (İzmir, *F5=5 km east of Kemalpaşa, on the Armutlu road, about 40 acres*), one field is located in the vicinity of Ödemiş (İzmir, *F6=2 km west of Ödemiş, on the Seyrekli road, about 30 acres*), one field is located in the vicinity of Nazilli (Aydın, *F7=10 km west of Nazilli, on the Aydın road, about 30 acres*), and one field is located in the vicinity of Söke (Aydın, *F8=5 km southeast of Söke, on the Bağarası road, about 60 acres*). The cotton fields were surrounded by abandoned grasslands, wheat fields, tobacco fields or another cotton fields or hedgerows.

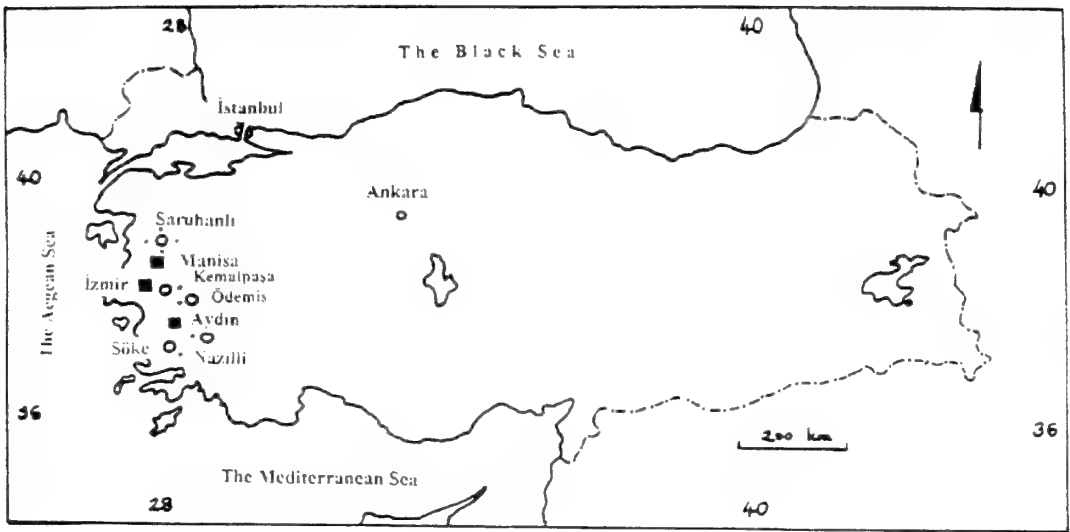


Fig. 1: The locations of the cotton fields where spiders collected in the Aegean Region of Turkey.

2. Climate :

The study area is in the range of the Mediterranean climate. The summer is hot and dry, and the winter is moderate and rainy. The hottest month is August (for instance, the average monthly temperature is 27°C in Kemalpaşa, and 28°C in Ödemiş), and the coldest month is January. In this area, the average annual temperature does not fall under 16°C. The average annual rainfall ranges between 700 and 1200 mm. Natural vegetation of the area is maquis. This area has got an important role in the agricultural development of Turkey.

3. Sampling techniques :

Four collections were made in each field (one a month) in each year. The spiders were collected from the vegetation and ground surface with aspirator. However, the Whole Plant Bag Sampling (WPBS) technique was used as well. About ten cotton plants were taken from each field in each collection. In the plant bag samplings, the plants were chosen at random. The transparent polithene bag (70 cm wide x 120 cm long) was pulled over the plant chosen, and the lower open mouth of the bag was tied around the stem of the plant with a string. Then the stem of the plant was cut off with a vineyard knife below the tied area, and the bag was labelled and the whole plant removed to the laboratory. In the laboratory, the plants inside their bags were placed in a deep-freezer to immobilize the spiders and insects. Then the plants were thoroughly searched, the spiders were collected with aspirator, and placed in glass tubes

containing 70 % alcohol. The spiders collected with aspirator during the field work were taken directly to the alcohol tubes. All sampling was done during the day-time. The laboratory work was carried out at the Zoological Research Center in Kırıkkale University. A total of 613 spiders were investigated during this study. The species, sex and stage of development of each specimen were recorded. Taxonomic determinations of the spiders were made with a Nikon AFX-MX model stereo binocular microscope. The juvenile spiders were identified to the family level only. All the adult spiders collected were deposited in the Zoological Research Center in Kırıkkale University.

Results and Discussion

During the four month collecting period for 1997 and 1998, a total of 613 spiders were collected. The spiders collected representing 12 families (Tables 1,2), 31 genera and 41 species. The collection was made in cotton fields located in Manisa, İzmir and Aydın provinces, in the western part of Turkey. Species numbers, individual numbers according to sex and stage, total numbers and percentage of each family are given in Table 1.

Table 1. Spider families collected from the cotton fields: Species numbers, individual numbers according to sex and stage, total numbers and percentage of each family

F a m i l y	Species Number	Male	Female	Adult	Immature	Total	%
Lycosidae	8	41	75	116	62	178	29.03
Linyphiidae	8	17	46	63	33	96	15.66
Salticidae	5	21	26	47	36	83	13.53
Philodromidae	3	13	20	33	35	68	11.09
Thomisidae	4	7	13	20	45	65	10.60
Gnaphosidae	4	14	12	26	23	49	7.99
Theridiidae	3	15	7	22	15	37	6.03
Oxyopidae	2	1	3	4	7	11	1.79
Amaurobiidae	1	0	2	2	9	11	1.79
Pisauridae	1	2	4	6	4	10	1.63
Araneidae	2	0	2	2	2	4	0.65
Hahniidae	-	0	0	0	1	1	0.16
Total	41	131	210	341	272	613	--
%	--	38.4	61.6	55.6	44.4	--	--

In this study, the most encountered spiders were wolf-spiders (Lycosidae, 29.03 %); followed by linyphiids (15.66 %) and salticids (13.53 %). Among these spiders lycosids were mostly collected from the ground surface and vegetation zone under 15 cm in height ("the ground zone", Duffey, 1966). However, some lycosids were captured from the upper part of the cotton plants as well. Most of the money spiders (linyphiids) were collected from the "field zone". On contrary, some of them were taken from the ground zone. Most of the salticid spiders were found on the leaves. The philodromid (11.09 %) and thomisid (10.60 %) spiders were collected from the two zones. Spiders belong to other families were captured in low percentages (under 10 percent). Of these, the gnaphosids were mostly found on soil surface. The theridiids were collected from the webs either in the field zone or the ground zone.

The species recorded from the cotton fields (arranged according to caught number of individuals) are as in the following :

Lycosidae: *Pardosa proxima* (C.L.Koch, 1847), *Pardosa morosa* (L.Koch, 1870), *Pardosa amentata* (Clerck, 1757), *Pardosa hortensis* (Thorell, 1872), *Pardosa pullata* (Clerck, 1757), *Alopecosa pulverulenta* (Clerck, 1757), *Trochosa ruricola* (De Geer, 1778), *Arctosa perita* Latreille, 1799. **Linyphiidae:** *Walckenaeria atrotibialis* (O.P.-Cambridge, 1878), *Lepthyphantes leprosus* (Ohlert, 1865), *Lessertia denticelis* (Simon, 1884), *Linyphia triangularis* (Clerck, 1757), *Nerienne furtiva* (O.P.-Cambridge, 1870), *Taranucnus setosus* (O.P.-Cambridge, 1863), *Erigone dentipalpis* (Wider, 1834), *Lepthyphantes tenuis* (Blackwall, 1852). **Salticidae:** *Salticus scenicus* (Clerck, 1757), *Phlegra fasciata* (Hahn, 1826), *Marpissa nivoyi* (Lucas, 1846), *Evarcha falcata* (Clerck, 1757), *Synageles dalmaticus* (Keyserling, 1863). **Philodromidae:** *Tibellus oblongus* (Walckenaer, 1802), *Thanatus formicinus* (Clerck, 1757), *Thanatus striatus* C.L.Koch, 1845. **Thomisidae:** *Xysticus cristatus* (Clerck, 1757), *Thomisus onustus* Walckenaer, 1806, *Xysticus robustus* (Hahn, 1831), *Xysticus bifasciatus* C.L.Koch, 1837. **Gnaphosidae:** *Haplodrassus signifer* (C.L.Koch, 1839), *Gnaphosa lugubris* (C.L.Koch, 1839), *Drassyllus pusillus* (C.L.Koch, 1833), *Aphantaulex seminigra* Simon, 1878. **Theridiidae:** *Steatoda bipunctata* (Linnaeus, 1758), *Steatoda albomaculata* (De Geer, 1778), *Euryopis flavomaculata* (C.L.Koch, 1836). **Oxyopidae:** *Oxyopes lineatus* Latreille, 1806, *Oxyopes nigripalpis* Kulczyński, 1907. **Amaurobiidae:** *Callobius claustrarius* (Hahn, 1833). **Pisauridae:**

Table 2. List of the species recorded from the cotton fields according to the caught numbers.

Species	Male	Female	Total
1. <i>Pardosa proxima</i> (Lycosidae)	22	50	72
2. <i>Salticus scenicus</i> (Salticidae)	19	17	36
3. <i>Walckenaeria atrotibialis</i> (Linyphiidae)	7	18	25
4. <i>Tibellus oblongus</i> (Philodromidae)	10	9	19
5. <i>Thanatus formicinus</i> (Philodromidae)	2	11	13
6. <i>Pardosa morosa</i> (Lycosidae)	6	5	11
7. <i>Xysticus cristatus</i> (Thomisidae)	4	7	11
8. <i>Steatoda bipunctata</i> (Theridiidae)	8	3	11
9. <i>Pardosa amentata</i> (Lycosidae)	6	4	10
10. <i>Steatoda albomaculata</i> (Theridiidae)	6	4	10
11. <i>Lepthyphantes leprosus</i> (Linyphiidae)	0	9	9
12. <i>Haplodrassus signifer</i> (Gnaphosidae)	6	3	9
13. <i>Gnaphosa lugubris</i> (Gnaphosidae)	3	5	8
14. <i>Drassyllus pusillus</i> (Gnaphosidae)	5	3	8
15. <i>Pardosa hortensis</i> (Lycosidae)	4	3	7
16. <i>Lessertia denticelis</i> (Linyphiidae)	0	7	7
17. <i>Linyphia triangularis</i> (Linyphiidae)	4	2	6
18. <i>Pardosa pullata</i> (Lycosidae)	2	4	6
19. <i>Pisaura mirabilis</i> (Pisauridae)	2	4	6
20. <i>Thomisus onustus</i> (Thomisidae)	2	3	5
21. <i>Neriene furtiva</i> (Linyphiidae)	5	0	5
22. <i>Alopecosa pulverulenta</i> (Lycosidae)	0	4	4
23. <i>Trochosa ruricola</i> (Lycosidae)	1	3	4
24. <i>Taranucnus setosus</i> (Linyphiidae)	1	3	4
25. <i>Erigone dentipalpis</i> (Linyphiidae)	0	4	4
26. <i>Phlegra fasciata</i> (Salticidae)	1	3	4
27. <i>Marpissa nivoyi</i> (Salticidae)	1	2	3
28. <i>Evarcha falcata</i> (Salticidae)	0	3	3
29. <i>Lepthyphantes tenuis</i> (Linyphiidae)	0	3	3
30. <i>Oxyopes lineaus</i> (Oxyopidae)	1	2	3
31. <i>Xysticus robustus</i> (Thomisidae)	1	2	3
32. <i>Arctosa perita</i> (Lycosidae)	0	2	2
33. <i>Callobius claustrarius</i> (Amaurobiidae)	0	2	2
34. <i>Xysticus bifasciatus</i> (Thomisidae)	0	1	1
35. <i>Synageles dalmaticus</i> (Salticidae)	0	1	1
36. <i>Thanatus striatus</i> (Philodromidae)	1	0	1
37. <i>Aphantaulax seminigra</i> (Gnaphosidae)	0	1	1
38. <i>Euryopis flavomaculata</i> (Theridiidae)	1	0	1
39. <i>Oxyopes nigripalpis</i> (Oxyopidae)	0	1	1
40. <i>Araneus quadratus</i> (Araneidae)	0	1	1
41. <i>Argiope lobata</i> (Araneidae)	0	1	1
Total	131	210	341

Pisaura mirabilis (Clerck, 1757). **Araneidae:** *Araneus quadratus* Clerck, 1757, *Argiope lobata* (Pallas, 1772). **Hahniidae:** unidentifiable immature individual.

Among these species, *Pardosa morosa* (L.Koch) (Lycosidae), *Tarantulus setosus* (O.P.-Cambridge), *Lepthyphantes tenuis* (Blackwall), *Linyphia triangularis* (Clerck) (Linyphiidae), *Gnaphosa lugubris* (C.L.Koch), *Euryopis flavomaculata* (C.L.Koch) (Theridiidae), *Marpissa nivoyi* (Lucas) (Salticidae) and *Callobius claustrarius* (Hahn) (Amaurobiidae) are new records for the Turkish spider fauna.

Pardosa proxima was the most encountered species (21.1 %) among the recorded spiders from the cotton fields. *Salticus scenicus* took the second row (10.5 %), and followed by *Walckenaeria atrotibialis* (7.3 %), *Tibellus oblongus* (5.6 %), *Thanatus formicinus* (3.8 %), *Pardosa morosa* (3.2 %), *Xysticus cristatus* (3.2 %), *Steatoda bipunctata* (3.2 %), respectively (Table 2). The other species were represented by lower percentages.

In the studies which have been done in terrestrial ecosystems by our team so far, the wolf spiders were more or less the most encountered spiders. Even they were found as dominant spiders among the ground-zone spiders (Bayram, 1993; Bayram & Luff, 1993; Bayram & Allahverdi, 1994; Bayram, 1994; Bayram *et al.*, 1998; Bayram & Varol, 1999; Bayram *et al.*, *in press*). These findings fit with some other records (Yeargan & Dondale, 1974; Nyffeler & Benz 1987, 1988). However, among the field-zone spiders the linyphiids, salticids, philodromids and thomisids were more encountered. In this work, the vegetation zone spiders such as linyphiids could be dominant if the whole plant bag sampling technique had heavily been used. Of course, the catch numbers of the philodromid, thomisid, theridiid and araneid spiders would be perhaps higher. Likewise in the works of Van den Berg *et al.* (1990) and Mansour (1987), the spiders such as araneids, dictynids, linyphiids, clubionids and gnaphosids took place in the upper rows. On the other hand, especially day active spiders were collected in this work. In order to collect the nocturnal spiders, the pitfall traps should be used as well in the cotton fields. Consequently, to investigate the spider fauna of cotton fields in a better way the combination of the methods of aspirator sampling, whole plant bag sampling and pitfall trapping may be more effective.

Acknowledgment

I would like to thank Hisham K. El-Hennawy for checking the English language of the paper, and constructive criticism to eliminate the errors.

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**The first landmark in the route of Egyptian
Arachnology : "*Explication Sommaire des Planches
d'Arachnides de l'Égypte et de la Syrie*" (1825)**

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Introduction

The first scientific record of a spider species from Egypt was that of Linnaeus (1758) in his 10th edition of "Systema Naturae" Tomus I. Regnum Animale. p.622 where he described, in a few words, the species *Aranea flavissima* : "A. abdomine oblongo flavissimo lævi. M. L. U. Habitat in Ægypto. Hasselqvist. Thorax fulvus. Pedes glabri." = "Spider with a light yellowish oblong abdomen. M. L. U. (Coll. S : æ Lovisæ Ulricæ Reginæ Museum) Habitat in Egypt. F. Hasselquist, 1749. Thorax deep yellow. Legs smooth." A brief description which is not enough to identify such a species or "qu'il sera toujours impossible d'identifier" as Simon (1910) stated.

The second record was that of Forskål (1775), who described four spider species from the region of Cairo : *Aranea citricola*, *A.insidiatrix*, *A.rivulata* and *A.trifasciata*. His descriptions were more detailed and, at least, enough for identification. His work was published after his death by Carsten Niebuhr in Copenhagen.

The third work and the real scientific study of arachnida of Egypt began with the "*Description de l'Égypte*". In this historical work, we can find the first landmark in the study of Arachnida of Egypt. It includes the work of Savigny on Egyptian arachnids, completed and edited by Audouin. The story of this work was summarized by Simon (1910):

In 1798, J.-C. Savigny, attached as a zoologist to the great French expedition to Egypt, worked on the magnificent drawings of the nine plates of Arachnids, of which the engraving, entrusted to the best artists of his time, were accomplished about 1811.

The publication was adjourned, nobody knew why, and the author became completely blind in 1824, the editing of the text was entrusted to Victor Audouin.

The 1st edition in-4° is a gathering of memoirs on diverse subjects of natural history, under the general title of *Description de l'Égypte ou Recueil des observations et des recherches qui ont été faites en Égypte pendant l'Expédition de l'armée française. Histoire naturelle, t. I, 1809*, a date which may deceive because all the memoirs which compose this volume were not published simultaneously; the part of Arachnids, which is incorporated in no. 4, p. 99, under the title : *Explication sommaire des planches d'Arachnides de l'Égypte et de la Syrie, publiée par Jules-César Savigny, membre de l'Institut; offrant un exposé des caractères naturels des genres, avec la distinction des espèces, par Victor Audouin*, is very subsequent, because Audouin had not been charged by editing until 1825, as proved by a ministerial letter inserted in the same book, p. 5, and beared the date 19 March 1825.

The 2nd edition in-8°, in 24 volumes, had been published few years later; volume 22, relating to natural history, beared the date of 1827, this last edition, was the only cited by Walckenaer and all the authors who followed him, nevertheless it was a literal reprint of the first under another format.

The share of Audouin in editing the text appears very minute, due to a note in the 1st edition: «M. Audouin se fait un devoir de déclarer qu'il a mis à profit la description des Arachnides commencée par M. Savigny, mais dont ce savant n'avait pu revoir les épreuves. Cette description, qui s'arrête à la pl. IV, a été souvent restreinte afin d'être mise en rapport avec l'explication sommaire des Mollusques, Annélides, Crustacés, etc.»

Indeed, the explanations relating to the 42 figures of plates I to IV were accompanied by summarized descriptions, and indications to localities and habitats, certainly by the hand of Savigny, while for the other plates, these indications defaulted and the descriptions, in all cases were reduced to minimum, sometimes also missing, notably for the Salticids.

The two authors : *Savigny* and *Audouin*

The life of the two authors of this work can be summarized from Pallary (1926), in his "Notice Biographique Sur J.C.Savigny", pp.1-6, as:

Marie-Jules-César-Lelorgne de Savigny was born at Provins in 5 April 1777. He was son and grandson of judges, and his mother belonged to a rich noble family. He departed to Egypt because Rouen lorsque Cuvier proposed him to make a trip for the commission which Bonaparte proposed to be incorporated in the expeditionary corps of Egypt (1798), with Geoffroy Saint-Hilaire as colleague for natural history. He was nominated as a member of the Institut (Académie des Sciences) in 1821.

Unfortunately during his stay in Egypt, Savigny contracted an illness of eyes which, under the influence of tiredness occasioned by the examination of the microscopic creatures which he studied, evolved rapidly and turned, in 1824, to complete blindness. A nervous illness also came, alas! to increase his state and to render all his intellectual work impossible, then he could not publish the text which must accompany his plates. Savigny died in 5 October and buried in 14 October 1851.

Victor Audouin, was born in 1797, distinguished naturalist, founder of the *Annales des sciences naturelles* and the Entomological Society. After having been deputy of Lamarck in the Museum, he was nominated as professor of entomology, instead of Latreille, who died in 1823. He had important observations on crustacea, silkworms, pyralid moths of vine, and *Histoire naturelle du littoral de la France*, in collaboration with Milne Edwards. Audouin had admired by the Academy of Sciences in 1838 and died early in the age of 44 (in 1841).

The problem of the authorship and the date of publication

Sherborn (1897) depended mainly on : (1) Engelmann, *Bibl.Hist.Nat.* p.340 and (2) *Ann.Soc.Entom.France*, xi., 1842, p.99, to state : "I have no doubt myself that all the parts enumerated above (vol. I., parts 1-4) may be safely regarded as dated 1826."

Tollitt (1986) followed the guidelines of Sherborn and stated : "In general, most workers tend to cite Audouin alone as the author of *Explication sommaire des planches*, probably as a result of Sherborn's original paper. Thus, in following Sherborn's analysis, the author and date of volume 1, part 4, *Explication sommaire des planches* may be cited as J. V. Audouin, [1826]."

Both Sherborn and Tollitt attributed volume 1, part 4 "solely to Jean Victor Audouin as he apparently had to begin the work again from the

beginning, because Savigny did not pass any of his manuscripts on to him after he went blind."

Two examples of the scientists who accepted "Audouin, 1826" are : Millidge (1988) following Tollitt and Baehr & Baehr (1995) following Sherborn.

Date of publication

The cover of the first edition of *Histoire Naturelle*, t.I carried the date M.DCCC.IX. (1809). This date is "Date incorrect" (ICZN, Art.21 d - I.U.B.S., 1985) as explained by Simon (1910). There is an evidence at the end of the *Note concernant l'Explication Sommaire des Planches dont les dessins ont été fournis par M. J.C.Savigny, pour l'Histoire Naturelle de l'ouvrage*. p.6 (Anonymous, 1825), as following :

Note

Distribution des planches de Zoologie dont les dessins ont été fournis par M. Savigny N.B. L'Histoire Naturelle de l'ouvrage et terminée par la Botanique, soixante-deux planches, et par la Minéralogie, quinze planches, formant la seconde partie du 2.e volume.

Paris, le 1.er novembre 1825.

Signé Lafont, Président; Girard, Fourier, Desgenettes, Geoffroy-Saint-Hilaire, Devilliers, Jollois, Le Père, Jomard.

Hence, the year of publication is 1825, depending on this evidence printed inside the work itself.

Authorship

There are two footnotes in p.99 (the first page of *Explication sommaire des planches d'Arachnides de l'Égypte et de la Syrie*) :

(1) M. Savigny observe que les dessins des Arachnides, exécutés sous ses yeux et dans son cabinet par MM. Meunier, Huet et Prêtre, ont été commencés en 1804, et qu'ils étoient tous terminés et donnés à la gravure en 1812. C'est pour ce motif que toutes les planches, même celles qui ont été terminées dans ces derniers temps, porteront cette date. Nous insistons sur cette déclaration, afin qu'on ne suppose pas que les travaux de M. Savigny ont pu être modifiés par des découvertes assez importantes dont la science s'est enrichie depuis peu, et qui n'avoient pas échappé à la sagacité de notre auteur : telle est, entre autres, l'observation curieuse de M. Treviranus, qui a démontré que les pièces situées à l'extrémité des palpes du mâle n'étoient autre chose qu'un appareil d'excitation, et que les organes propres à ce sexe existoient au-dessous du premier segment de l'abdomen, c'est-à-dire, à la même place que dans la femelle. Notre savant ami M. Savigny avoit remarqué ce fait avant l'anatomiste allemand.

(2) Voyez ci-dessus, page 3, la *Note concernant L'Explication sommaire des planches dont les dessins ont été fournis par M. J. C. Savigny, pour L'Histoire Naturelle de L'Ouvrage*. M. Audouin se fait un devoir de déclarer qu'il a mis à profit la description des Arachnides commencée par M. Savigny, mais dont ce savant n'avoit pu revoir aucune épreuve. Cette description, qui s'arrête à la planche 4, a été souvent restreinte et modifiée, afin d'être mise en rapport avec L'Explication sommaire des Mollusques, Annelides, Crustacés, &c.

They elucidate the role of Savigny and that he described the specimens figured in plates 1-4, and stopped because of blindness. Then, Audouin completed the work after being ordered by Son Exc.le Ministre Secrétaire d'état au département de l'intérieur, as stated in the note mentioned above (p.3) : Déterminé par ces diverses considérations, le Ministre a écrit à la Commission d'Égypte, en même temps qu'à M. Savigny, qu'il chargeoit M. Audouin, naturaliste et son élève, de mettre ces planches en état de paroître, en y inscrivant les noms généraux et toutes les désignations nécessaires, et en rédigeant une explication très-sommaire, suffisante pour l'intelligence des figures; mais en même temps Son Excellence a réservé à M. Savigny tous ses droits comme auteur; son intention étant que les explications fournies par M. Audouin ne puissent nuire aux travaux scientifiques du premier, et qu'elles se rapportent constamment aux parties déjà faites, de manière que M. Savigny puisse un jour donner à ses recherches la suite qu'il jugera convenable.

Simon (1910) also noticed that the description of specimens are accompanied by localities, only in 42 figures (in pl.1-4, except the last two specimens in pl.4). I think that Savigny depended on his notices or his memory to remember the locality of every specimen figured to state its locality, where the specimen was collected. After being blind, it was impossible to him to remember the locality of any figured specimen. It was not possible too, to know the relation between any of Savigny's written notices and any specimen figure to be of use to Audouin.

Therefore, it is evident that Savigny is responsible for the description of the specimens figured in plates 1-4, which are accompanied by localities, except the last two specimens in pl.4 (42 figures). Savigny is responsible too for the names of new species and genera included in the text (pp.106-150). While Audouin is responsible for the names of new species included in the text (pp.151-186) (70 figures).

In addition, there are only 6 new valid genera of spiders in this work. They are : *Nemesia*, *Ariadna*, *Hersilia*, *Erigone*, *Argiope*, and *Ocyale*. All of them are described in the section of Savigny (pl.1-4). Also, Savigny's descriptions are twice longer than those of Audouin.

The Arachnid species published in "*Description de l'Égypte*"

The nine plates of Arachnida includes 112 figures. Seven plates were devoted to spiders : 89 figures (81 species of 30 genera classified in 20 groups) = 71 species + 6 synonyms + 5 nomina dubia (in the most recent classification).

Plate 8 : figs. 1-3 scorpions : 3 species of *Scorpio* = 3 species;

figs. 4-6 pseudoscorpions : 3 species of *Chelifer* = 2 species + 1 n.d.;

figs. 7-10 solpugids : 4 species of *Solpuga* = 3 species + 1 syn.

Plate 9 : figs. 1-3 opilionids : 3 species of *Phalangium* = 1 sp. + 2 n.d.;
figs. 4-13 acarids : 10 species of 3 genera = 6(+2) spp. + 1 syn. + 1 n.d.

Every figure group of a species includes a habitus drawing and minor details drawings.

In the following table : 1) all the arachnid species described in "*Description de l'Égypte. Histoire naturelle, t. I, p. 4*" are mentioned, noting 2) the sex of the specimen figured, with 3) number of pages in both the first (1825) and the second (1827) editions, 4) the locality of the first 42 figured specimens, and 5) the valid name currently used.

All species names are revisioned and synonymized according to :
Order Araneida (El-Hennawy, 1990; Platnick, 1993 & 1997), Order Scorpionida (El-Hennawy, 1992), Order Pseudoscorpionida (Harvey, 1990), Order Solpugida (El-Hennawy, 1998), Order Opilionida Cokendolpher, 1990), Order Acarida (Nuttall, *et.al.*, 1908).

A note on the misspellings in the second edition is provided. The drawing of the male palpal organ in spiders is noted in 21 species (see footnote (1) above in "Authorship" section). Also, all male solpugid species figured are accompanied by flagellum drawing. It was a great addition in the time of Savigny.

The locality was mentioned for the first 42 figured specimens [40 species (now 37), Egypt 36 (33), Palestine 5]. There were 7 localities in Egypt : Alexandrie (15 species), Rosette (11), Kaire (8), Damiette (2), Menzaleh (2), Sâlahyeh (1), Delta (1); and 3 localities in Palestine : Acre (3), Carmel (1), Jaffa (1).

There are 3 figures which were described as male spiders while they were subadult or juvenile as shown in the figure. They are :

Pl. 2 fig. 5 *Argyope aurelia*, subadult male.

Pl. 4 fig. 12 *Eresus Dufourii*, juvenile.

Pl. 6 fig. 11 *Thomisus hirtus* ?, subadult male.

Note 1 : Planche 1-7, 9 Dessiné et gravé en 1805-12; pl. 8 en 1806-11.

2 : In Edit. II (1827) p.328 the currently used spelling *Argiope* is used once instead of *Argyope*.

Acknowledgments

I am grateful to Dr. J.Gruber (Vienna) and Dr. M.Fouda (Cairo) for the photocopies of the papers of Sherborn (1897) and Tollitt (1986) respectively.

Table of Arachnid species published in "Description de l'Égypte"

Plate 1

Fig.	Species	♂♀	Edit. I	Edit. II	Locality	Valid Name
1	<i>Nemesia cellicola</i> Sav.	♀	107-108	304-305	Des environs d'Alexandrie	<i>Nemesia cellicola</i> Savigny, 1825
2	<i>Segestria perfida</i> Walck. *	♂♀	108-109	306-307	De l'intérieur des maisons et des caves d'Alexandrie	<i>Segestria florentina</i> (Rossi, 1790)
3	<i>Ariadna insidiatrix</i>	♀	109-110	308-309	De l'intérieur des maisons d'Alexandrie	<i>Ariadna insidiatrix</i> Savigny, 1825
4	<i>Lachesis perversa</i> *	♂	111	311-312	Des environs du Kaire	<i>Lachesana perversa</i> (Savigny, 1825)
5	<i>Tegenaria domestica</i> Walck.	♀	112-113	312-314	Des maisons d'Alexandrie	<i>Tegenaria parietina</i> (Fourcroy, 1785)
6	<i>Arachne familiaris</i>	♀	113-114	315-316	Des maisons de Rosette	<i>Tegenaria domestica</i> (Clerck, 1757)
7	<i>Arachne timida</i> *	♂	114	316	Des jardins de Rosette	<i>Agelena timida</i> (nomen dubium)
8	<i>Hersilia caudata</i>	♀	115	318	Des environs du Kaire	<i>Hersilia caudata</i> Savigny, 1825
9	<i>Erigone vagans</i>	♂	116-117	320-321	Des jardins du Kaire	<i>Prinerigone vagans</i> (Savigny, 1825)

Plate 2

1	<i>Uloborus flavus</i> Sav.	♀	117-118	322-323	Des environs de Rosette et des jardins du Kaire	<i>Tetragnatha flava</i> (Savigny, 1825)
2	<i>Eugnatha nitens</i>	♀	118-119	323-325	Des environs de Rosette	<i>Tetragnatha nitens</i> (Savigny, 1825)
3	<i>Eugnatha pelusia</i> *	♂♀	119-120	325-327	De l'île de Rosette et des îlots du lac Menzaleh	<i>Tetragnatha nitens</i> (Savigny, 1825)
4	<i>Eugnatha filiformis</i>	♀	120-121	327	De l'intérieur du Delta	<i>Tetragnatha filiformis</i> (Savigny, 1825)
5	<i>Argiope aurelia</i>	s♂♀	122-124	331-334	Des environs du Kaire et des environs d'Alexandrie	<i>Argiope trifasciata</i> (Forskål, 1775)

6	<i>Argiope sericea</i>	♀	124-125	334-335	Des environs du Kaire et des environs d'Alexandrie	<i>Argiope lobata</i> (Pallas, 1772)
7	<i>Argiope splendida</i>	♀	125-126	335-336	Des environs d'Acre	<i>Argiope lobata</i> (Pallas, 1772)
8	<i>Epeira armida</i>	♀	126-127	337-338	Des environs d'Acre	<i>Aculepeira armida</i> (Savigny, 1825)
9	<i>Epeira circe</i>	♀	127-128	338-339	De l'intérieur des maisons d'Alexandrie	<i>Araneus circe</i> (Savigny, 1825)
10	<i>Epeira apoclis</i> Walck.	s♂♀	128-129	339-341	Des bords du Nil, aux environs de Rosette et de l'île de Rosette	<i>Larinioides cornutus</i> (Clerck, 1757)

Plate 3

1, 2	<i>Epeira apoclis</i> *	♂	130-132	341-344	De l'île de Rosette	<i>Larinioides folium</i> (Schrank, 1803)
3	<i>Epeira umbratica</i>	♂	132	345	Des environs de Damiette	<i>Nuctenea umbratica</i> (Clerck, 1757)
4	<i>Epeira lucina</i>	♀	132-133	345-347	De l'île de Rosette	<i>Singa lucina</i> (Savigny, 1825)
5	<i>Epeira chloris</i>	♂	133	347	Des environs d'Acre	<i>Larinia chloris</i> (Savigny, 1825)
6	<i>Clotho Durandii</i> Walck.	♀	134-135	348-349	Du mont Carmel	<i>Uroctea durandi</i> (Latreille, 1809)
7	<i>Enyo nitida</i>	♀	135-136	350-351	Des environs d'Alexandrie	<i>Zodarion nitidum</i> (Savigny, 1825)
8	<i>Enyo longipes</i> *	♂	136	351-352	Des environs du Kaire	<i>Zodarion nitidum</i> (Savigny, 1825)
9	<i>Latrodectus erebus</i>	♀	137	352-353	Des environs de Sâlahyeh	<i>Latrodectus tredecimguttatus</i> (Rossi, 1790)
10	<i>Latrodectus argus</i>	♀	137-138	353-354	Des environs d'Alexandrie	<i>Latrodectus tredecimguttatus</i> (Rossi, 1790)
11	<i>Latrodectus venator</i>	♀	138	354-355	Des environs d'Alexandrie	<i>Steatoda venator</i> (Savigny, 1825)

12	<i>Pholcus rivulatus</i> *	♂	140-141	358-359	Dans l'intérieur des maisons du Kaïre	<i>Holocnemus pluchei</i> (Scopoli, 1763)
13	<i>Pholcus phalangioides</i> Walck. *	♂♀	141-142	360	Dans l'intérieur d'une maison d'Alexandrie	<i>Pholcus phalangioides</i> (Fuesslin, 1775)

Plate 4

1	<i>Sphasus alexandrinus</i>	♀	142-143	361-362	Du désert aux environs d'Alexandrie	<i>Oxyopes heterophthalmus</i> (Latreille, 1804)
2	<i>Lycosa tarentulina</i> *	♂♀	143-145	363-367	Des environs d'Alexandrie	<i>Allocosa tarentulina</i> (Savigny, 1825)
3	<i>Lycosa arenaria</i>	♀	146	367-368	Du désert aux environs de Rosette	<i>Evippa arenaria</i> (Savigny, 1825)
4	<i>Lycosa peregrina</i>	♀	146	368	Des environs de Rosette	<i>Lycorma peregrina</i> (Savigny, 1825)
5	<i>Lycosa pelliona</i>	♀	146-147	368-369	Des environs de Rosette	<i>Ocyale pelliona</i> (Savigny, 1825)
6	<i>Lycosa agretyca</i> Latr.	♀	147	369	Des rives du canal d'Alexandrie	<i>Geolycosa urbana</i> (Cambridge, 1876)
7	<i>Lycosa Nilotica</i>	♀	147-148	369-370	Des rives du canal d'Alexandrie	<i>Lycosa nilotica</i> Savigny, 1825
8	<i>Lycosa pelusiaca</i>	♀	148	370	Des bords du lac Menzaleh	<i>Alopecosella pelusiaca</i> (Savigny, 1825)
9	<i>Dolomede hippomene</i> ¹	♀	148-149	371	Des environs de Damiette	<i>Dolomedes hyppomene</i> Savigny, 1825
10	<i>Ocyale atalanta</i>	♀	150	374	Des environs de Jaffa	<i>Ocyale atalanta</i> Savigny, 1825
11	<i>Eresus Petagnae</i>	♀	151	375-376	---	<i>Eresus petagnae</i> Audouin, 1825
12	<i>Eresus Dufourii</i>	o	151-152	376-377	---	<i>Stegodyphus dufouri</i> (Audouin, 1825)

Plate 5

Fig.	Species	♂♀	Edit. I	Edit. II	Valid Name
1	<i>Scytodes thoracica</i> Latr.	♀	152-153	378-379	<i>Scytodes thoracica</i> (Latreille, 1802)
2	<i>Scytodes rufescens</i> Duf.	♀	153-154	379-380	<i>Loxosceles rufescens</i> (Dufour, 1820)
3	<i>Dysdera erythrina</i> Latr.	♀	154	380-381	<i>Dysdera erythrina</i> (Walckenaer, 1802)
4	<i>Drassus Listeri</i>	♀	155	382-383	<i>Zelotes listeri</i> (Audouin, 1825)
5	<i>Drassus Schaefferi</i> ²	♀	156	383	<i>Pterotricha schaefferi</i> (Audouin, 1825)
6	<i>Drassus Lyonnnetii</i> *	♂	156	383-384	<i>Trachyzelotes lyonneti</i> (Audouin, 1825)
7	<i>Drassus Linnaei</i>	♀	156	384	<i>Pterotricha linnaei</i> (Audouin, 1825)
8	<i>Clubiona Albini</i>	♀	157	385-386	<i>Aphantaulax albini</i> (Audouin, 1825)
9	<i>Clubiona Listeri</i>	♀	157-158	386-387	? (nomen dubium)
10	<i>Philodromus Clerckii</i>	♀	159	388-389	? (nomen dubium)

Plate 6

1	<i>Philodromus Walckenaërii</i> ³	♀	159-160	390	<i>Eusparassus walckenaeri</i> (Audouin, 1825)
2	<i>Philodromus Linnaei</i> *	♂	160-161	390-391	<i>Eusparassus walckenaeri</i> (Audouin, 1825)
3	<i>Philodromus Fabricii</i> *	♂	161	392	<i>Thanatus fabricii</i> (Audouin, 1825)
4	<i>Philodromus Albini</i> *	♂	161	392	<i>Thanatus albini</i> (Audouin, 1825)
5	<i>Philodromus rhombiferens</i> Walck.	♀	161-162	392-393	<i>Thanatus formicinus</i> (Clerck, 1757)
6	<i>Selenops Aegyptiaca</i>	♀	162-163	394-395	<i>Selenops radiatus</i> Latreille, 1819
7, 8	<i>Thomisus Peronii</i>	♀	163	395-396	<i>Thomisus onustus</i> Walckenaer, 1805
9	<i>Thomisus Martyni</i> *	♂♀	163-164	396	<i>Pistius truncatus</i> (Pallas, 1772)
10	<i>Thomisus Buffonii</i> *	♂	164	396-397	<i>Heriaeus buffoni</i> (Audouin, 1825)
11	<i>Thomisus hirtus</i> ? Latr.	s♂♀	164-165	397-398	<i>Xysticus bliteus</i> (Simon, 1875)
12	<i>Thomisus Lalandii</i> *	♂♀	165	398	<i>Xysticus lalandei</i> (Audouin, 1825)

13	<i>Thomismus Clerckii</i>	♀	165	398	<i>Xysticus clercki</i> (Audouin, 1825)
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Plate 7

1, 2	<i>Thomismus Diana</i> ? Walck.	♂	165-166	399	<i>Synema diana</i> (Audouin, 1825)
3, 4, 5	<i>Thomismus rotundatus</i> Walck. *	♂♀	166-167	399-401	<i>Synema globosum</i> (Fabricius, 1775)
6, 7	<i>Platyscelum Savignyi</i>	♂	167-168	402-403	<i>Palpimanus gibbulus</i> Dufour, 1820
8	<i>Attus Adansonii</i> *	♂	169	404-405	<i>Hasarius adansonii</i> (Audouin, 1825)
9	<i>Attus Dorthesii</i>	♀	170	405-406	<i>Aelurillus dorthesi</i> (Audouin, 1825)
10	<i>Attus Druryi</i>	♂	170	406	? (nomen dubium)
11	<i>Attus Frischii</i>	♀	170	406	<i>Pellenes frischii</i> (Audouin, 1825)
12	<i>Attus Gesneri</i>	♀	170	406	<i>Menemerus gesneri</i> (Audouin, 1825)
13	<i>Attus tardigradus</i> Walck.	♀	170	406-407	<i>Hasarius adansonii</i> (Audouin, 1825)
14	<i>Attus Bonnetii</i>	♀	170	407	<i>Mogrus bonneti</i> (Audouin, 1825)
15	<i>Attus cupreus</i> Walck. *	♂	171	407	<i>Heliophanus cupreus</i> (Walckenaer, 1802)
16	<i>Attus Mouffettii</i>	♀	171	407	? (nomen dubium)
17, 18	<i>Attus Soldanii</i> *	♂♀	171	407-408	<i>Menemerus soldani</i> (Audouin, 1825)
19	<i>Attus Hunterii</i>	♀	171	408	<i>Menemerus hunteri</i> (Audouin, 1825)
20	<i>Attus Illigerii</i>	♀	172	408	<i>Menemerus illigeri</i> (Audouin, 1825)
21	<i>Attus Redii</i>	♂	172	408-409	<i>Langona redii</i> (Audouin, 1825)
22	<i>Attus Paykullii</i> *	♂	172	409	<i>Plexippus paykulli</i> (Audouin, 1825)

Plate 8

1	<i>Scorpio occitanus</i> ? Am.	♂	173	410-411	<i>Leiurus quinquestriatus</i> (Hemprich & Ehrenberg, 1828)
2	<i>Scorpio Amoreuxii</i>	♀	173-174	411-412	<i>Androctonus amoreuxi</i> (Audouin, 1825)
3	<i>Scorpio australis</i> Herbst.		174	412	<i>Androctonus bicolor</i> Hemprich & Ehrenberg, 1828

4	<i>Chelifer sesamoides</i>			174-175	413-414	<i>Chelifer cancroides</i> (Linnaeus, 1758)
5	<i>Chelifer Hermannii</i>			175	414	<i>Olpium savignyi</i> Simon, 1879 (nomen dubium)
6	<i>Chelifer Beauvoisii</i>			175	414	<i>Garypus beauvoisi</i> (Audouin, 1825)
7	<i>Solpuga araneoides</i> Olivier	♂♀		176-178	416-419	<i>Galeodes araneoides</i> (Pallas, 1772)
8	<i>Solpuga intrepida</i> Duf.	♂		178	419	<i>Galeodes araneoides</i> (Pallas, 1772)
9	<i>Solpuga melanus</i> Oliv.	♂♀		178-179	419-420	<i>Rhagodes melanus</i> (Olivier, 1807)
10	<i>Solpuga phalangium</i> Oliv.	♂		179	420	<i>Rhagoditta phalangium</i> (Olivier, 1807)

Plate 9

1	<i>Phalangium Aegyptiacum</i> Sav.	♂		180-181	422-424	<i>Phalangium aegyptiacum</i> Savigny, 1816 (nomen dubium ?)
2	<i>Phalangium Copticum</i> Sav. ⁴	♀		181-182	424	<i>Phalangium copticum</i> Savigny, 1816 (nomen dubium ?)
3	<i>Phalangium Savignyi</i>	♂		182	424	<i>Phalangium savignyi</i> Audouin, 1825
4	<i>Acarus Savignyi</i>	♂		182-183	425-426	?
5	<i>Argas Savignyi</i>			183-184	426-427	<i>Ornithodoros savignyi</i> (Audouin, 1825)
6	<i>Argas Fischeri</i>			184	427-428	<i>Argas vespertilionis</i> (Latreille, 1796)
7	<i>Argas Hermannii</i>			184	428	<i>Argas hermanni</i> Audouin, 1825
8	<i>Argas persicus</i> Fisch. ⁵			184	428	<i>Argas persicus</i> (Oken, 1818)
9	<i>Ixodes Leachii</i>			185	428	<i>Haemaphysalis leachi</i> (Audouin, 1825)
10	<i>Ixodes Aegyptius</i> Latr.			185	428-429	<i>Hyalomma aegyptium</i> (Linnaeus, 1758)
11	<i>Ixodes Fabricii</i>			186	429	<i>Hyalomma aegyptium</i> (Linnaeus, 1758)
12	<i>Ixodes Linnaei</i>			186	429	<i>Rhipicephalus</i> sp. ?
13	<i>Ixodes Forskaelii</i>			186	430	<i>Argas</i> sp. ?

In "Edit. II" : 1. *Dolomedes hyppomene*. 2. *Drassus Scaefferi*. 3. *Philodromus Walckenaerii*. 4. *Phalangium qobiticum*. 5. *Argas Persicus*.

* With drawing of male palpal organ.

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SERKET

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